

CHAPTER I

INTRODUCTION

Nepal is roughly rectangular in shape. The country landmass stretches 885 km from east to west and has a non-uniform width of 193 km north to south. It lies within the sub-tropical to the mountainous region at 26° 22' to 30° 27' N latitudes and 80° 4' to 88° 12' E longitudes, with an altitude that ranges from 90m to 8,848 m. the country is landlocked and is bordered by India in the East, West and South, and China in the North. Geographically, Nepal represents a transitional mountain area between the fertile Gangetic Plain of India and the arid plateau of China.

It has a total land area of 147, 181 sq.km. Topographically, Nepal is divided into three ecological regions terai, hill and mountain. The narrow strip of flat alluvial terrain along the southern border, known as the terai, is an extension of the Gangetic Plain and comprises about 23% of the country, including most of the fertile and forest areas. The terai the Churia and Mahabharat Ranges punctuate the terai plains and constitutes the fertile with alluvial soil, with a good water holding capacity. Its northern edge is the Bhabar. The broad flat valleys or Duns found between successive hill ranges. The first elevation next to the terai is the Siwaliks (also known as Churia Range). There are a number of terai-like valleys lying between the Siwaliks and the Mahabharat range, commonly called the Dun Valleys (inner terai plains), such as Chitwan and Dang.

Running parallel with the Churia range is the middle mountain zone, also known as the middle hills or the mahabharat range. This zone also includes the so-called "middle hills" which extend northwards in a somewhat confused maze of ridges and valleys to the foot of the great Himalayas. It has great rivers such as the Karnali, Narayani and Saptakoshi flow through this area into the broad plains of the terai. The high mountain zone, located north of the middle mountain region. This differs from the previous type in its northward, on the long, straight and steep slopes, and narrow valleys which are sensitive to erosion. and spurs of the great Himalayas. The high mountain zone, located north of the Middle Mountain Region. The high himal zone occupies about 23 per cent of the

kingdom and is mostly snow covered. The snow line is at 5,000 m in the east and 4,000 m in the west. This zone is an area of rocky, ice-covered massifs, rolling uplands, snow-fields, valley glaciers, and sweeping meadow lands.

The terai area receives heavy precipitation, ranging between 180 and 225 cm. The relative humidity varies between 80 percent and 90 percent during the monsoon but declines in the other months. The climatic conditions in the different parts of the country especially in terai region is favorable for the breeding of *Culex* mosquitoes, the proven vectors of Japanese Encephalitis and Filariasis and for the availability of the various amplifying hosts of the disease.

In Nepal, four mosquito-borne diseases are prevalent and cause much morbidity and mortality. They are Malaria (Peters et al., 1955, Brydon et al., 1961, Johnson 1966, Pradhan et al., 1970, Shrestha and Parajuli 1980, Parajuli et al., 1981, Shrestha et al., 1988), Japanese Encephalitis (JE) (Pradhan 1981, Khatri et al., 1983, Henderson et al., 1983), Filariasis (Kessel 1966, Jung 1973) and Dengue Fever.

1.1 *Culex* species as vector

Japanese Encephalitis (JE) and Filariasis are principally transmitted by *Culex* mosquitoes, although anophelines do play a secondary role in the transmission. *Culex tritaeniorhynchus* is a major vector of JE virus in many part of the Oriental Region, including Nepal (Hammon et al., 1949, Hale et al., 1957, Buescher et al., 1959, Reuben et al., 1971a, 1971b; Leake et al., 1986). JE has been isolated from *Culex fuscocephala* in Thailand (Gould et al., 1974) and is a suspected vector in Nepal. *Culex gelidus* is a suspected vector of JE and was the most abundant mosquitoes collected in Sunsari district during September-October 1985 (Leake et al., unpublished data). This virus has been isolated from *Cx. gelidus* in Thailand (Gould et al., 1962). *Cx. quinquefasciatus* is the principle vector of *Wuchereria bancrofti* in Nepal, which is within the endemic zone of Filariasis (Jung 1973). *Culex thelieri* has been found naturally infected with Sindbis and West Nile Viruses (Mcintosh et al., 1967).

Studies of encephalitis before and after World War II, several groups of Japanese and foreign workers supported the view that the mosquito *Culex*

tritaeniorhynchus plays an important role as the vector of JEV in Northern Asia and wherever JEV occurs. In Japan and Okinawa *Culex tritaeniorhynchus* was the only mosquito consistently infected and was showed to be the principal vector of JE. Studies of encephalitis before and after World War II several groups of Japanese and foreign workers supported the view that the mosquito *Culex tritaeniorhynchus* plays an important role as the vector of JEV in northern Asia and wherever JEV occurs.

In Japan and Okinawa *Culex tritaeniorhynchus* was the only mosquito consistently infected and was showed to be the principal vector of JE. In India *Culex vishnui* and *Culex bitaeniorhynchus* and *Culex annulorostris* were the probable vector of JEV. Furthermore, in Japan Okinawa, Korea and China, JEV has been frequently recorded from other *Culex* such as *Culex pipiens*, *Culex quinquefasciatus*, *Culex tritaeniorhynchus*. *Aedes onicus* and *Aedes togoi*. Isolation from *Aedes vexans nipponi* has also been reported from Japan and from *Anopheles vagus*, *Anopheles annularis* were isolated virus in India.

Culex tritaeniorhynchus and other possible vectors of JEV have been recorded from almost all the problem areas of Nepal, which are proven vectors of Indian subcontinent. Historically encephalitis cases were recorded in the hospitals located in the southern plain terai belt of Nepal bordering India. However, these cases were never investigated, and the etiology of the disease was not known until the first epidemics of Japanese encephalitis occurred in a few districts of the western region in July 1978. At that time, a few cases were serologically confirmed as Japanese encephalitis virus (JEV).

Since the first epidemics of Japanese Encephalitis (JE) in 1978, epidemic is occurring every alternate year, and the case incidence is increasing in regular increments up. Until now, the prevalence of three different structures of JEV (Nep. 1/90, B-2524, and B-9548) has been reported. Now 24 districts of terai and inner terai of the region are affected. However, no detailed study of the vector species has been carried out in Nepal.

The total 8874 cases and 1264 death have been reported in Nepal with an average case fatality rate of 14.2 percent in aggregate since 1998. During six years period of time, highest number of cases were reported in 1999 (2924 cases) and the second highest were in 2001 (1888 cases). The lowest number of cases (330 cases) with highest mortality (CFR 20.9 percent) has

been reported during the year 2003. The overall mortality of JE varies from 9.77 percent during the year 2000 to 20.9 percent during the year 2003. Comparative assessment of disease in different regions showed that Far western and Mid-western Development Regions have reported the highest number of JE cases during the years 1998 to 2003. The reporting districts based on the abundance of number of cases per 100,000 populations are in the order of Banke, Kailali, Kanchanpur, Parsa, Rupandehi and Morang. Cases started to appear in April-May and reached their peak during late August to early September. Cases started to decline from October EDCD (2001).

Filariasis has been known to be endemic in different areas of Nepal since a long time. It is a public health problem in Nepal. Filariasis is much more prevalent in Nepal due to the presence of suitable environmental condition and mosquito breeding sites. In Nepal, very few researches on Lymphatic filariasis have been undertaken. Out of three species of lymphatic parasites; *Wuchereria bancrofti*, *Brugia malayi* and *Brugia timori*, only one species i.e. *Wuchereria bancrofti* is recorded in Nepal. The previous works on filariasis were done by Jung (1973), Pradhan et al., (2003) studied mapping of filarial infection in 37 districts of total 75 districts of Nepal. The study showed that 33 districts of them were endemic for lymphatic filariasis. Of the 33 districts 11 districts were having above 20% prevalence rate, 15 districts between 6-19% and 7 districts between 1-5%. The average 13% of the whole population of Nepal was found to be affected (Sherchand, 2003).

The achievement of MDA in the country by the year 2007 was found to be covered 11 millions of population (DOHS, 2006) and followings are the rounds of MDA conducted in the different districts of Nepal: 5th Round-1 district (Parsa), 3rd Round-4 districts (Makwanpur, Chitwan, Nawalparasi and Rupandehi) and 1st Round-15 districts (Nuwakot, Dhading, Gorkha, Tanahun, Shyangja, Palpa, Kapilbastu, Kavrepalanchok, Ramechhap, Sindhuli, Dhanusa, Mahottari, Sarlahi Rautahat and Bara) (DoHS, 2006) .

Dengue fever is commonly referred to as breakbone fever. This is another viral disease transmitted by mosquitoes. *Aedes aegypti*, *Ae. albopictus*, *Ae. Scutellaris* are suitable vectors, *Ae. albopictus* is prevalent in Japan, New Guinea, Norther Australia, Malagasy Republic, the Philippines, and Hawaii.

Ae.albopictus is distinguishable by a silvery stripe on its mesonotum and whitish irregular patches on the lateral aspects of its thorax. Adult are aggressively anthropophilic and are important vectors of dengue viruses (Gould et.al., 1968, Huang 1972). Larvae occur in tree holes in forested areas, also rain barrels and artificial containers. *Aedes sculellaris* is a closely related species characterized by whitish wavy lines composed of scales down each side of the thoracic pleuron.

The Government of Nepal has identified Japanese encephalitis as a priority program. In this relation, government has shown its commitment to implement different disease prevention and control activities and strengthen the diagnostic capacity for different established JE diagnostic laboratories. Moreover, national protocol has been developed for JE diagnosis and government is committed for better clinical management and treatment of disease. For this, government has planned to immunize 250 thousand populations of different JE endemic areas in current fiscal year.

A sharp rise in the JE and Filaria mortality rate has been observed in Chitwan district. JE and Filaria are transmitted by *Culex* mosquito vectors. JE and Filaria control strategies can be more successful if the distribution and abundance of mosquito vectors is predicted. This study aims to provide a better understanding of the distribution and abundance of *Culex* mosquitoes in Chitwan district including vectors.

1.2 Breeding site of vector species

Immature stages of *Culex fuscocephala* have been collected in rice fields, ground pools, foot prints and marshes. In Nepal larvae have been dipped from shallow pools in swampy ground and irrigation ditches. Immatures *Culex gelidus* live in puddles, pools, rice fields and marshy depressions having abundant vegetation. Immature stages of *Culex quinquefasciatus* are found in any type of habitat from fresh and clear to brackish, turbid and polluted waters. It is common in ground pools, ditches, drains, sewage, latrines, septic tanks and artificial containers. *Culex theileri* breed in permanent and temporary bodies of fresh or foul water. They are commonly found in slow moving streams, irrigation ditches, ground pools, ponds, springs and cisterns. *Culex tritaeniorhynchus* is a common rural species in rice fields, shallow marshes, pools, ponds and ditches containing fresh or polluted water

with grass or aquatic vegetation in partial shade or full sun. This species becomes dominant in rice paddies when plants reach 0.3m in height (Darsie and Pradhan 1990).

1.3 Previously Reported Culex species in Chitwan District

Culex bitaeniorhynchus was reported in Chitwan, Rapti Valley (Peters and Dewar, 1956). *Culex tritaeniorhynchus* was reported in Chitwan, Rapti Valley, VI-63, 6 .

1.4 Aims and Objectives

The aims and objectives of this study are as follows:

- 1.4.1 To determine the distribution of different Culex mosquito species on selected villages of Chitwan district;
- 1.4.2 To determine the indoor and outdoor species present in the study area and their relative density;
- 1.4.3 To assess seasonal changes;
- 1.4.4 To determine vector abundance and species composition in the study area;
- 1.4.5 To determine resting habit of mosquito species in the study area.

1.5 Formulation of Hypothesis

The expected results of this study are as follows:

- 1.5.1 The number of mosquitoes depends on temperature i.e. there is positive relationship between the number of mosquitoes and temperature.
- 1.5.2 The number of mosquitoes collected in post-monsoon survey and pre-monsoon survey is independent.

CHAPTER II

LITERATURE REVIEW

Moore et al., (1978) states that Mosquito density was positively correlated with rainfall, the relationship being more marked in the dry, south-coastal part of the island in Puerto Rico.

Takashima et al., (1989) reported at the time of the out breaks of abortion, *Culex pipiens*, *Anopheles* species, *Aedes vexans hipponii*, and *Aedes japonicus* were predominant over *Culex tritaeniorhynchus*. *Culex tritaeniorhynchus*, almost a solve vector species of JE virus in the Southern part of Japan, is probably not a vector of the virus in Hokkaido.

Gingrich et al., (1992) states that, vector abundance was high in monsoon (May-October), moderate in transition (March-April and November-December) and low in dry (January-February) seasons in Bangkok. More pigs sero-converted in monsoon and transition seasons than in dry seasons. Indices of JE transmission activity (vector abundance, pig sero-conversions, and MIRS) increased proportionately with rainfall. The risk of human infection appeared greatest at the site with moderate vector abundance because of its greatest human population density.

Geevarghese et al., (1994) reported very high incidence of JE cases were in extensively irrigated areas and a low incidence in some of the taluks with less or no irrigation system in Mandya District, Karnataka, India. Mosquito populations were peak during March-April and September.

Gajanana et al; (1997) reported vector abundance was lowest in the hot and dry season (April-June) and highest in the cool and wet season (October-December) in the South Arwt district in Tamil Nadu. The probability of a child receiving an infective bite was 0.53 per JE transmission season.

Prakash et al., (1997) studied the indoor biting behaviour of *Anopheles dirus* Peyton and Harrison, 1979 in upper Assam, India and recorded a high

level of endophagy of *Anopheles dirus* with a mean biting rate of 8.7 ± 9.0 . Indoor biting of *An. dirus* 5% started after 19.00hr, increased gradually, was more pronounced between 21.00 and 04.00hr, and ceased after 05.00hr.

Mwandawiro et al; (1999) studied on the host preference of Japanese encephalitis vectors in Chiang Mai, North Thailand and states that, there was no difference in the proportions of fed females of *Culex tritaeniorhynchus* and *Cx. vishnui* between the cowshed and pigsty collections, and *Cx. gelidus* fed significantly more on the cows than the pigs.

Hanna et al; (1999) reported JE virus activity was more widespread in North Queensland in the 1998 wet season than in the three previous wet seasons, but ecological circumstances (e.g., less intensive pig husbandry, fewer mosquitoes) appear to have limited transmission on the mainland.

Das et al; (2000) studied on the prevalence of *Aedes aegypti* at the international part and airport, Kolkata (West Bengal), India

Guimaraes et al., (2000) reported *Anopheles cruzii* and *Aedes scapularis* feeding on blood inside and around the residence in areas of Serra do Mar state park state of Sao Paulo, Brazil. Both species were vectors of malaria and arbovirus respectively, may be involved in the transmission of such disease in rural areas.

According to Tandon and Ray (2000), a high bovine blood index of *Aedes aegypti* and *Aedes albopictus* in the cattle sheds and a high human blood index in the human dwelling in the highly congested residential areas of the city, and from outdoor situations in urban garden in Kolkata India. A majority of members of both the species had fed on one host and a small percentage on more than one host, and that both the species were mainly anthropophilic in nature.

Johansen et al., (2001) conducted an entomological investigation of an outbreak of JEV in the Torres Strait, Australia, in 1998, and recovered 43 isolates of JE virus from adult mosquitoes (42 isolates from *Culex sitiens* and one from *Ochlerotatus vigilax*). They also reported two

confirmed human JE cases in that area and Cape York Peninsula, in Northern Queensland.

Maier (2002) studied the effect of environmental changes on vectors of disease in Germany and neighboring countries with special reference to mosquitoes

According to Myrty et al., (2002) Ninety-three cases of Japanese encephalitis (27 confirmed serologically) were reported in an endemic district of Andhra-Pradesh, India, Mainly during the monsoon months of November and December. A significant positive correlation between densities of mosquitoes of the *Culex vishnui* subgroup and occurrence of Japanese encephalitis cases ($r=0.765$, $P<0.01$) was observed.

In Delhi and environs the feeding behaviour of Anopheline and Culicine mosquitoes was found to be highly influenced by biotypes. Village and areas, different climatic sleeping behaviour of human beings and availability of alternative hosts particularly animals population. Overall anthrophilic index was found to be 24.66% for *Anopheles culicifacies*, 20.96% for *An. annularis*, 26.33% for *Culex quinquefasciatus* and 36.39% for *Cx. vishnui* (Kumar et al., 2002).

Apperson et al., (2002) studied on Host feeding habits of *Culex* and other mosquitoes in the Brough of Queens in New York City and reported, *Culex pipiens* L. and *Cx. restuans* Theobald fed primarily on birds, and their feeding habits support their implication as enzootic vectors West Nile Virus. *Culex salinarius coquilleti* and *Coquilletidia perturbans* (walker) fed mainly on mammals with fewer blood meals taken from birds, and these two species are potential bridge vectors of West Nile Virus.

Eighteen mosquito species recorded by Ronald Ross since 1898, all of them are vector in mature, was noted within next 50 years in Kolkata. After 1950, 19 mosquito species, 8 recorded prior to and 11 arrived after 1950, b are regular visitors of human dwellings and cattle sheds of Kolkata 10 vector and 9 non vector mosquito species were sucking blood from man and cattle in

Kolkata. *Culex quinquefasciatus* was dominant (62.96%) on average of 492 mosquitoes (Pramanik and Raut, 2002).

Fakoorziba et al., (2003) reported *Cx. tritaeniorhynchus* from different breeding places in and around Mysore city in South India can be different varieties. One variety comparatively has shorter siphon index ratio and longer basal hair tufts on siphon than the one. Mating behaviour of these varieties show that, variety from the city pools was moderate eurygamous and the one from mainly paddy fields moderate stenogamous.

Kanojia et al., (2003) studied on long-term vector abundance and seasonal prevalence in relation to the occurrence of Japanese encephalitis in Gorakhpur district, and reported that *Cx. tritaeniorhynchus* the most likely vector of JE together with other known vector species remained more active during the period of paddy cultivation.

Dutta et al., (2003) conducted a survey on medically important mosquitoes found in Mizoram and recorded forty-seven species in the monsoon season and 48 in the post-monsoon season. Both the primary vectors of malaria, viz, *Anopheles dirus* and *An. minimus* were recorded. Potential vectors of dengue and Japanese encephalitis were also detected; fifteen species reported earlier were seen missing in the present survey.

In Paraiba valley region, state of Sao Paulo Brazil, *Aedes albopictus* commonly found to be fed on a wide range of host, and *Cx. quinquefasciatus* presented similar behaviour but humans and dogs were the most common (Gomes et al., 2003).

Barbosa et al., (2003) studied on the culicidae activity in a restrict forest in Curitiba urban area (Parana, Brazil) & reported that 15 species were found out of 312 Culicidae specimens. The predominant species belong to *Culex (Culex) coronation* Dyer and knob, 1906 group. Others species found with potential epidemiological importance were: *Haemagogus (Conopostegus) leucocelaenus* (dyar and Shannon, 1924), *Ochlerotatus (Ochlerotatus) flaviatilis* (IUTZ, 1904) *Anopheles (Nyssorhynchus)* (Strodei, 1926).

Rajendran et al., (2003) reported average vector abundance per man hour for *Culex tritaeniorhynchus* was 324.5 per month for the period June 1998- May 2000 in south Indian villages. The average minimum infection rate (MIR) per month in *Cx. tritaeniorhynchus* was 1.4 (range 0.0-5.6). Seroconversions (SCs) were recorded in 14 goats (70%) in the first year and 23 goats (74%) in the second year.

Taibe-Lagos et al., (2003) reported twenty-five species or genetic groups were identified among 53,496 specimens collected in a preserved metropolitan area. *Ochlerotatus scapularis*, *Culex quinquefasciatus* and *Culex declarator* were the most frequent and abundant species have been implicated in disease transmission to man in other regions.

Jeong et al., (2003) studied on prevalence and seasonal abundance of the dominant mosquito species in a large marsh near coast of Ulsan and reported that, *Anopheles sinensis* was most abundant (53.4% in species ratio), followed by *Culex tritaeniorhynchus* (43.0%), *Cx. inatomii* (1.6%) *Ochlerotatus dorsalis* (1.3%) and *Cx. pipiens pallens* (0.5%). A malaria vector *An. sinensis* and a Japanese encephalitis vector. *Cx. tritaeniorhynchus* were collected 3,663.3 females and 3,142.5 females per trap night from June to September for the years, respectively. According to the biweekly population changes at the area, *Cx. inatomii* which was dominant species in 1997 was the most abundant in the early July during 1999-2001.

According to Zhou et al., (2004), in western Kenya, *Anopheles gambiae* Giles was the predominant malaria vector species, constituting 84.6% of the specimens, whereas *Anopheles funestus* Giles constituted 15.4% of the vector populations. *An. gambiae* abundance increased by six to eight-folds in the long rainy season over the dry seasons, but *An. funestus* abundance peaked 3mo after the long rainy season. *An. gambiae* distribution was more aggregated in the wet seasons than in the dry seasons. Degree of aggregation of *An. funestus* was similar in all four seasons.

Kumar et al., (2004) conducted a study on mosquito diversity in Rajiv Gandhi National Park (Nagarahole), Karnataka state, India and recorded a

total of 60 species belonging to 10 genera. Genus *Culex* was predominant. Maximum number of species was found to breed in ponds of the total number of species recorded, 14 are known to be vectors of different mosquito-borne disease in India.

Fettene et al., (2004) studied the behaviour of *Anopheles arabiensis* and *An. quadriannulatus sp.B* mosquitoes and malaria transmission in south western Ethiopia and reported majority of indoor-resting *An. arabiensis* (79.6%) and *An. quadriannulatus sp.B* (94.8%) were caught in cattle sheds. The human blood index of *An. arabiensis* was 7.3% and only one specimen was positive for *Plasmodium* parasites, having both *P. falciparum* and *P. vivax* sporozoites and giving a sporozoite rate of 0.24%. The human blood index of *An. quadriannulatus sp. B* was 1.1% which was significantly different from that of *An. arabiensis*, positive reaction for *Plasmodium* circumsporozoite proteins were not observed in the *An. quadriannulatus sp. B* that were tested.

Dos et al., (2004) reported the density of *Anopheles albitarsis* females showed a marked seasonal variation, with peaks as high as 629 individuals per human per night in the wet season. The low daily survival probability of 0.61 is an important factor limiting the vectorial capacity of *An. albitarsis s.l.* in Southeastern Brazil.

Enrih and Boca (2004) conducted a study on seasonal dynamics of the *Anopheles maculipennis* complex in Osijek, Croatia. During an eight-year investigation of *Anopheles maculipennis*, using CDC traps with Co₂ as an attractant, a total of 3,508 mosquitoes were collected. Long term high water level created better conditions for continuous breeding of mosquitoes, which contributed to a significant increase in abundance rates of all species in the complex.

Sharma et al., (2005) studied on seasonal fluctuation of dengue fever vector, *Aedes aegypti* in Delhi, India and reported water cooler and tires were preferred breeding habitats of *Aedes* mosquitoes in the city out of 103,778 houses surveyed, 20,513 houses and 3,547 containers were reported positive for *Aedes aegypti*. The house container and Breteau indices were very high

during the post-monsoon season. The container indices was very high (17.7%) in the defence area in September 2000. The house index for *Aedes aegypti* ranged from 0.1 to 7.4, 0.1 to 11.3 and 0.1 to 11.1 in the municipal corporation of Delhi (MCD), New Delhi Municipal committee (NDMC) and defence areas, respectively.

Arunachalam et al., (2005) conducted observation on the multiple feeding behavior of *Culex tritaeniorhynchus* in Eerala in southern India and states that, *Culex tritaeniorhynchus* had fed mainly (56.6%) on cattle. Pig feeding accounted 6.3% of the total samples. Some samples (n=980, 38.3%) were of serologic mixed origin. Mixed blood meals were mostly (96.7%) from cattle and goats.

Russell (2005) studied on the species diversity of mosquito on Florida. Most of the mosquito species were collected during "high season" month (June to September) than in low season (January to March).

Ulloa et al., (2006) studies on host selection and gonotrophic cycle length of *Anopheles punctimacula* in southern Mexico and states that, *Anopheles punctimacula* was most abundant during the dry season and preferred animals to humans. The daily survival rate in mosquitoes collected in animal traps was 0.96 (Parity rate=0.80; gonotrophic cycle=4 days). The minimum time estimated for developing mature eggs after blood feeding was 72lt. The proportion of mosquitoes living enough to transmit *Plasmodium vivax* malaria during the dry season was 0.35.

Richards et al., (2006) studied the host feeding patterns of *Aedes albopictus* in relation to availability of human and domestical animals in Suburban Landscapes of central north, Carolina and states that, *Aedes albopictus* fed predominately on mammalian hosts (83%), common mammalian hosts included humans (24%), cats (21%) and dogs (14%), about 7% of blood meals was also taken from avian host (chickens and a northern cardinal). These mosquitoes took mixed blood meals from avian and non avian hosts.

Harbison et al., (2006) studied on sampling method for indoor resting malaria mosquitoes in Africa and reported that more mosquitoes were collected when a resting box and a ceiling net used together than a single collector using a hand-held aspirator but only one-third the number collected by pyrethrum spray catches (PSCS). At sites where PSCS are impractical, a resting box and ceiling net can effectively used as an alternative to hand catches in malaria surveillance.

Quiroga-Elizondo et al., (2006) state that, mosquitoes feed on humans with less frequency in comparison with chickens horses and pigs in Monterrey, Northeastern Mexico. The FRS for chickens were the highest of all available hosts (1.7 and 3.2) and they were the most abundant hosts in Eswbedo, and the second most abundant in Guadalupe, indicating a selective bias of *Cx. quinquefasciatus* for chickens i.e. Ornithophilic.

Parida et al., (2006) studies host feeding patterns of malaria vectors of Orissa, India, and reported 0.2% *Anopheles culicifacies* blood meals contained blood from humans and cattle. *An. flaviatilis* and *An. culicifacies* revealed seasonality in their anthropophilic index (AI). *An. flaviatilis* showed a human forage ratio of more than 1 where as *An. culicifacies*, *An. annularis*, and *An. varuna* had forage ratios of 2.6, 2.5 and 2.4 respectively, for bovine. There was a correlation between the AI of *An. flaviatilis* and malaria slide positively rate.

Kent et al., (2007) reported transmission of *Plasmodium falciparum* was nearly zero during the 2004-2005 rainy seasons because of widespread drought. During 2005-2006, the estimated Entomologic inoculation rate values were 1.6 and 18.3 infective bite per person per transmission season in each of the two village areas, respectively. *An. arabiensis* throughout Africa was the primary vector responsible for transmission of *P. falciparum*.

Kanojia Pc. (2007) reported *Cx. tritaeniorhynchus* have a major role in the transmission of JE virus in Bellary district Karnataka, India Risk of transmission of JEV to humans can be reduced by house spraying with residual insecticides and intermittent paddy irrigation.

Howell et al., (2007) studied the influence of house construction on the indoor abundance of mosquito and reported the mean number of *Culex quinquefasciatus* mosquitoes was greater in cement homes than in either wood or other poorer quality homes [Trinidad (TT) cement 17.43, others 14.43; Dominican Republic (DR) cement 4.24, others 3.41]. High abundance of mosquitoes resting indoors with painted interiors than without painted. But a painted exterior was not significant. Indoor resting mosquitoes in the TT (interior OR 2.90, CL 1.09-8.72, exterior OR 2.14, CL 0.89-6.67) and DR (interior OR 3.13, CL 1.41-6.92, exterior OR 1.97 CL.91-4.26).

Molaei et al; (2007) conducted a study on host feeding pattern of *Culex quinquefasciatus* and its role in transmission of west Nile virus in Harris country, Texas. About 672 blood-engorged *Culex quinquefasciatus*, collected during 2005, were identified by nucleotide sequencing PCR product of the cytochrome b gene 39.1% had required blood from birds, 52.5% from mammals, and 8.3% were mixed avian and mammalian blood meals. Most frequent vertebrate hosts were dog (41.0%) mourning dove (18.3%), domestic cat (8.8%) white-winged dove (4.3%), house sparrow (3.2%), house finch (3.0%), gray cat bird (3.0%) and American robin (2.5%) *Culex quinquefasciatus* is an opportunistic feeder and principal mosquito vector of WNV in this Metropolitan area; however, transmission by other mosquito species or by other modes of infection, such as ingestion, must account for the high WNV infection rates among local blue jays and American crows.

In Nepal, Pradhan (1988) reported seven new country records which include: *Aedes pulchriventer*, *Ae. subalbopictus*, *An. fragilis*, *An. dravidicus*, *Armigeres durhami*, *Culex infula* and *Culex pseudovishnui*.

Burgess (1990) found out that in Nepal the study of mosquitoes begins only recently, he recorded 130 species and subspecies in 14 genera of mosquitoes known from Nepal.

Darsie and Pradhan (1990) reported the taxon *Anopheles (Anopheles) gigas* Glies (Complex), as *Anopheles gigas* var *bailey*. *Anopheles (Celiia)*

filipinae Manalang. This species was also reported by Pradhan and Brydon (1960) from Lamjung district in the North Central Nepal.

Peters and Dewar (1956) were the first to record certain culicine species occurring in Nepal. Joshi et al., (1965) made a major contribution by reporting 59 species of Culicine, including 28 new country records. Shrestha (1966) reported 97 species, including 36 anophelines and 61 culicines.

Darsie and Pradhan (1994) reported 167 species and subspecies in 16 genera including 54 species of *Aedes*, 44 species of *Anopheles*, 31 species of *Culex*, 12 species of *Armigeres*, 12 species of *Uranotaenia*, 4 species *Mimomyia*, 3 species of *Mansonia* and *Heizmannia*, 2 species of *Malaya* and *Tripteroides* mosquitoes in Nepal. Out of 44 species of *Anopheles* mosquitoes five species has been identified incriminated as vector of primary importance. These are *Anopheles mimimus*, *A. fluviatilis*, *A. maculates*, *A. willmori* and *A. anullaris*.

Darsie (1994) found out that since 1987, a study of species of mosquitoes occurring in Nepal has been under way. No previous work has been understood to record the total mosquitoes of this Himalayan Kingdom of the Nepal.

Gautam et. al., (2009) reported first record of *Aedes aegypti* (L.) vector of Dengue Virus from Kathmandu, Nepal.

Parajuli et al., (1992) carried out an epidemiological study of JE in all epidemic districts of Nepal during the year 1989 and reported that of 868 total JE cases, 227 died. All ages and both sex groups were affected from the disease.

Joshi et al., (1995) carried out an epidemiological survey of JE in all endemic areas of Nepal through National Zoonoses and Food Hygiene Research Centre (NZFHRC) from 1990 to 1993. The CFR were 36.0%, 38.0%, 35.2% and 31.7% in 1990, 1991, 1992 and 1993, respectively.

Zimmerman et al., (1997) reported a first outbreak of JE in Kathmandu Valley, Nepal during September and October, 1995 and treated 15 patients with meningo-encephalitis.

Akiba et al., (2001) conducted an epidemiological study of JE outbreak in the South-Western part of Nepal in 1997 and reported a high density of JE infections. It was estimated that 27.9% of the total population were infected with JE virus in the study area.

Gurung and Singh (2003) studied on factors associated with JE in Nepal. Total of 142 numbers of confirmed JE cases and 142 controls from Banke, Bardia and Dang district were interviewed. They concluded that non-immunization status, presence of households pigs, non-use of mosquito-net and out-door sleeping are the risk factors related with the occurrence of JE.

CHAPTER III

METHODOLOGY

3.1 Description of Study Area

Nepal is a land of extremes and its topography varies from 60m to highest peak in the world. The country is a landlocked mountainous lying between China and India with an area of 1,47,181 sq. Km. About 38 percent of the population lives below the absolute poverty line and lack of Arable Land indicates that the population pressure is high in Nepal. On an average, seven persons are depending on each hectare of arable land for their livelihood (APP, 1997-2017).

Nepal's topographical and Socio-ecological diversification helps to promote periodic epidemics of infectious diseases, epizootics and natural disasters. Millions of people are at risk of infection and hundreds of them die every year due to communicable diseases, malnutrition and other health related events. As also majority of the population is rural area based and illiterate, economic and demographic changes coupled with sudden occurrence of epizootics might possibly have contributed to the disease out breaks. Japanese Encephalitis (JE), Filaria, Malaria, Kala-azar etc. have been affecting large number of people in successive years.

Chitwan District, a western part of Narayani Zone, is one of the seventy-five districts of Nepal, a landlocked country of South Asia. Bharatpur (seventh largest city of Nepal) is the district headquarters of Chitwan District. Bharatpur is the commercial and service center of central South Nepal; it is the merger destination for higher education, health and transportation of the region.

It is an inner Terai district widened from Bavor region of Terai to Mahabharat hill of north, bordered with Makawanpur and Parsa to the east, Nawalparasi and Tanahun to the west, Dhading and Gorkha to the north and Bihar of India to the south. It covers an area of 2,218 sq. Km and has a population (2001) of 472,048. Ninety percent of population is concentrated at plane valley, while the rest 10.00% in Mahabharat Hill.

It is situated at 27⁰35' N latitude and 84⁰30' E longitude and 144m to 1947m altitude range from sea level. The area enjoys two types of climate: Sub-tropical at Mahabharat range and tropical at foothill of Churia range and at valley areas. Temperature ranges from minimum 7⁰c to maximum 42⁰c.

Narayangadh, on the bank of Narayani River is the main town with numerous shopping Zones. At the foot of the Himalayas, Chitwan is one of the few remaining undisturbed vestiges of the Terai region which formerly extended over the foothills of Nepal.

Now there are 40 village development committees (each of which has nine wards or villages) and municipality Bharatpur and Ratnanagar each of which has more than nine wards or urban areas. Chitwan is very rich in beautiful natural scenes, lakes, forest, and river with well-known Chitwan National Park.

3.2 Study area and sampling sites (Number of sites)

The study was conducted on the ten villages among five VDC of Chitwan district. Gitanagar, Kesherbag, Parasnagar, Champanagar village of Gitanagar VDC; Bijayanagar, Rampur villages of Mangalpur VDC; Bramhapuri village of Patihani VDC of Western Chitwan and Suryapur, Kholesimal village of Jutpani VDC; Dobato Village of Pithuwa VDC of Eastern Chitwan.

3.3 Map of Chitwan District Showing Mosquitoes Sampling Sites



Figure 1 Map of Chitwan district

3.4 Agriculture Sector of Chitwan

Chitwan is very famous in Nepal because of its dominant production of mustard from which mustard oil is produced. This popularity of the mustard plantation in Chitwan is attributed to the predominant soil type of the place. The soil in Chitwan is mostly of the silt type. The silt nature of the soil is in turn attributed to the flooding over the ages in the past from the rivers like Gandaki. Chitwan is also profusely spotted with lands with soil type clay which are very good for use as paddy fields. The paddy crop is available in two period, pre-monsoon (March-April) and post-monsoon (August-September).

Maize and wheat is the second and third most cultivated crop of Chitwan. Wheat is cultivated in winter season but Maize is cultivated in all

season in Chitwan. Though in some leisure time, farmers used to grow some varieties of grains and pulses such as millet, buck wheat etc., millet is planted in Shrawan (July-August) but buck-wheat is winter crop of Chitwan.

The soil is also very good for growing various types of vegetables such as cabbage, cauliflower, radish, potato, broccoli, carrot, capsicum (winter vegetable) cucumbers, pumpkins, lady's finger, (summer vegetable).

Gitanagar's rice and potatoes of Pithuwa are most popular in Chitwan. Chitwan product 80% of the country poultry industry, and it is also famous for floriculture, Mushrooms cultivation, Bee forms and Honey production etc.

3.5 House structure

Most of the human dwellings were made by bricks with mud; the roof was made by Zinc. Only few were made by mud with hedge roof, and some were cemented house.

3.6 Cattle-Shed

Majority of animal shelter was open type and made up of wooden block with hedge roof and some are closed type made by brick with wooden block with zinc roof.

3.7 Sampling Techniques

3.7.1 Study Design

The study was conducted from August-September 2007 (Post monsoon/rainy season). A unique house code (Like G, P, K and so on) was given on the front of each house. Households were surveyed in sequence daily along black from the start house between 6:00 am to 10:00 am. Un-occupied or closed houses and houses where resident did not provide permission for the survey, offices and schools were not sampled. Continuing surveys of un-sampled households an attempt was made to inspect houses that were previously closed or where access had been refused. About four houses were sampled per day per village and total of forty houses were sampled per season. Mosquitoes were not collected on the rainy and windy day. The number of human occupants as well as cattle occupants during the previous night was recorded for each survey house. The indoor collection

(human and cattle shed) was completed in 15 / 15 minute and outdoor collection in 30 minute.

The temperature of indoor collection (human and cattle shed) and outdoor collection of each house was noted separately using thermometer. The appeared specimen was instantly sucked by aspirator and they were transported to the plastic cup. The collected mosquitoes were identified using identification key by Darsie and Pradhan (1990).

Immediately after termination of first survey, the sampling procedure was repeated. The second survey was completed on March-April 2008. To differentiate data associated with the different collections, the two survey were referred as 'a' and 'b'.

3.7.2 Entomological Surveys

Briefly, after asking permission to survey the household, a demographic survey designed to determine the number of occupants, dimension of the property, house construction materials, method of cooking, water use patterns type of sewage disposal, and insecticide use etc. were administered. Indoor collections of adult mosquitoes were attempted in all rooms of the house (when permitted) including walls, under furniture and inside closets and other likely adult mosquito resting sites (cattle sheds).

Outdoor collections similarly attempted outside the house from outside wall, under eaves, vegetation and bushes around cattle sheds and around outdoor-stored material etc.

3.8 Requirements

3.8.1 Materials Required

1. Aspirator
2. Ocular Microscope
3. Plastic cup
4. Ivory paper

5. Entomological pin
6. Thermocol
7. Torch light
8. Mosquito net
9. Cotton
10. Forceps
11. Plastic bag

3.8.2 Chemical required

- 1 Nephthalene ball
- 2 Ethylacetate
- 3 Nail polish (colourless)

3.9 Killing Method

The collected mosquitoes were killed by using the ethylacetate. For this, a plastic cup was mounted with mosquito net with hole on the center. The hole was covered with a piece of cotton. The mosquitoes were transferred into the cup from the aspirator through this hole. A small piece of cotton was soaked with one or two drops of ethylacetate and placed into this cup and the cup was made airtight, after few second the mosquitoes were killed.

3.10 Fixing method

Only *Culex* mosquitoes were fixed on the Ivory paper. Ivory paper was cut into triangular (one end pointed) card. A drop of colorless nail polish was placed at the apex of the triangular card. Each collected specimen was fixed with the help of colorless nail polish; the apex of the card is slightly bent at an angle to make the specimen up right. The bent tip was attached to the pleuron of thorax. The other end of the card was picked with Entomological pin after giving the code number, such specimen were fixed in thermocol kept in box. Such stag specimens were kept in special care to prevent the damage of wings, legs, maxillary palps, proboscis and abdomen. Nephthalene ball was kept inside the box by attaching with glue. Complete information about

collected locality, time, site, number of such stag specimens were noted in a questionnaire paper.

3.11 Identification

Each are ever-stag specimen i.e., *Culex* group were identified in the lab to the species level using the identification key by Pradhan and Darsie (1990).

3.12 Tools for Analysis and Presentation:

After identifying these collected mosquitoes, it was processed, arranged and analyzed. The analysis has been made more effective with the help of tables, bar diagrams, graphs wherever necessary. But for testing of hypothesis, Correlation and Hypothesis testing model have been used.

3.13 Analytical Framework:

As far as possible, the analysis has been made quantitative. For Correlation of number of mosquitoes on temperature, the following variables have been defined.

- I. Number of Mosquitoes: These are total number of mosquitoes collected in each village in one season. This variable; say (X).
- II. Temperature: The temperature recorded in field i.e. average temperature of each village in one season (Post Monsoon and Pre Monsoon season). This variable; say (Y).

'n' is the total number of observation.

After having done this, simple Correlation Coefficient is calculated. The formula for calculating simple Correlation Coefficient is

$$\text{Correlation Coefficient}(r) = \frac{n \sum XY - \sum X \sum Y}{\sqrt{[n \sum X^2 - (\sum X)^2][n \sum Y^2 - (\sum Y)^2]}} \dots\dots \text{Annex-1}$$

Again, for testing, the second hypothesis, that is, the number of mosquitoes collected in Post-Monsoon and Pre-Monsoon survey is independent. The following concepts and method have been followed:

The data has been arranged in to (rxc) manifold contingency table where 'r' represents the number of rows and 'c' represents the number of columns. The season divided into two groups Post Monsoon season and Pre

Monsoon season and there is 12 species of culex mosquitoes. Now for testing hypothesis χ^2 test is used as a test of independence of attributes at 5% level of significance. Symbolically,

$$t^2 = \sum \frac{(O - E)^2}{E} \dots\dots\dots \text{Annex-2}$$

Where,

O= Observed Frequency

E= Expected Frequency

E can be calculated as:

$$E_{ij} = \frac{\text{Total of } i^{\text{th}} \text{ row} \times \text{Total of } j^{\text{th}} \text{ column}}{\text{Grand total}}$$

CHAPTER IV

RESULTS

A total of 1197 *Culex* mosquitoes belonging to 12 species were collected during 4 months period (August and September in post-monsoon and March and April in pre-monsoon) all total of 20 days (80 man-hours) period.

5.1 Number and Density of Mosquitoes collected in Indoor and Outdoor Collection

5.1.1 Post-monsoon survey

The total of 718 *Culex* Mosquitoes belong to 12 species (12 species in cattle shed, 11 species in human dwelling and outdoor collection) were recorded during ten day period in August and September. *Culex quinquefasciatus* is the predominant species about 121/121 samples were recorded from human dwelling and cattle shed and 80 from Outdoor collection. *Culex mimulus* is the least dominant species only one sample was collected from cattleshed. The density of *Cx. mimulus* was highest i.e, 1 in Cattleshed. In total highest density 4.91 was recorded in Cattleshed. Most of the samples (n=279) were recorded from human dwelling. See table 1.

Table 1
Number and Density of Mosquitoes

| Name of species | INDOOR | | | | OUTDOOR | | TOTAL | |
|----------------------------------|----------------------|--------|-----------------------|--------|----------------------|-----------------------|----------------------|-----------------------|
| | Number of Mosquitoes | | Density of Mosquitoes | | Number of Mosquitoes | Density of Mosquitoes | Number of Mosquitoes | Density of Mosquitoes |
| | Human | Cattle | Human | Cattle | | | | |
| <i>Cx. quinquefasciatus</i> | 121 | 121 | 0.37 | 0.37 | 80 | 0.24 | 322 | 0.98 |
| <i>Cx. fuscocephala</i> | 103 | 87 | 0.41 | 0.34 | 60 | 0.24 | 250 | 0.99 |
| <i>Cx. tritaeniorhynchus</i> | 21 | 21 | 0.44 | 0.44 | 5 | 0.10 | 47 | 0.98 |
| <i>Cx. gelidus</i> | 9 | 7 | 0.32 | 0.25 | 12 | 0.42 | 28 | 0.99 |
| <i>Cx. bitaeniorhynchus</i> | 5 | 9 | 0.25 | 0.45 | 6 | 0.3 | 20 | 1 |
| <i>Cx. whitmorei</i> | 6 | 4 | 0.5 | 0.33 | 2 | 0.16 | 12 | 0.99 |
| <i>Cx. pseudovishnuivis hnoi</i> | 4 | 5 | 0.36 | 0.45 | 2 | 0.18 | 11 | 0.99 |
| <i>Cx. whitei</i> | 4 | 3 | 0.44 | 0.33 | 2 | 0.22 | 9 | 0.99 |
| <i>Cx. theileri</i> | 2 | 2 | 0.28 | 0.28 | 3 | 0.42 | 7 | 0.98 |
| <i>Cx. barraudi</i> | 2 | 3 | 0.28 | 0.42 | 2 | 0.28 | 7 | 0.98 |
| <i>Cx. sinensis</i> | 2 | 1 | 0.5 | 0.25 | 1 | 0.25 | 4 | 1 |
| <i>Cx. mimulus</i> | 0 | 1 | 0 | 1 | 0 | 0 | 1 | 1 |
| Grand Total | 279 | 264 | 4.15 | 4.91 | 175 | 2.81 | 718 | 11.87 |

Figure 2
Number of Mosquitoes collected in indoor and outdoor
collection in Post Monsoon

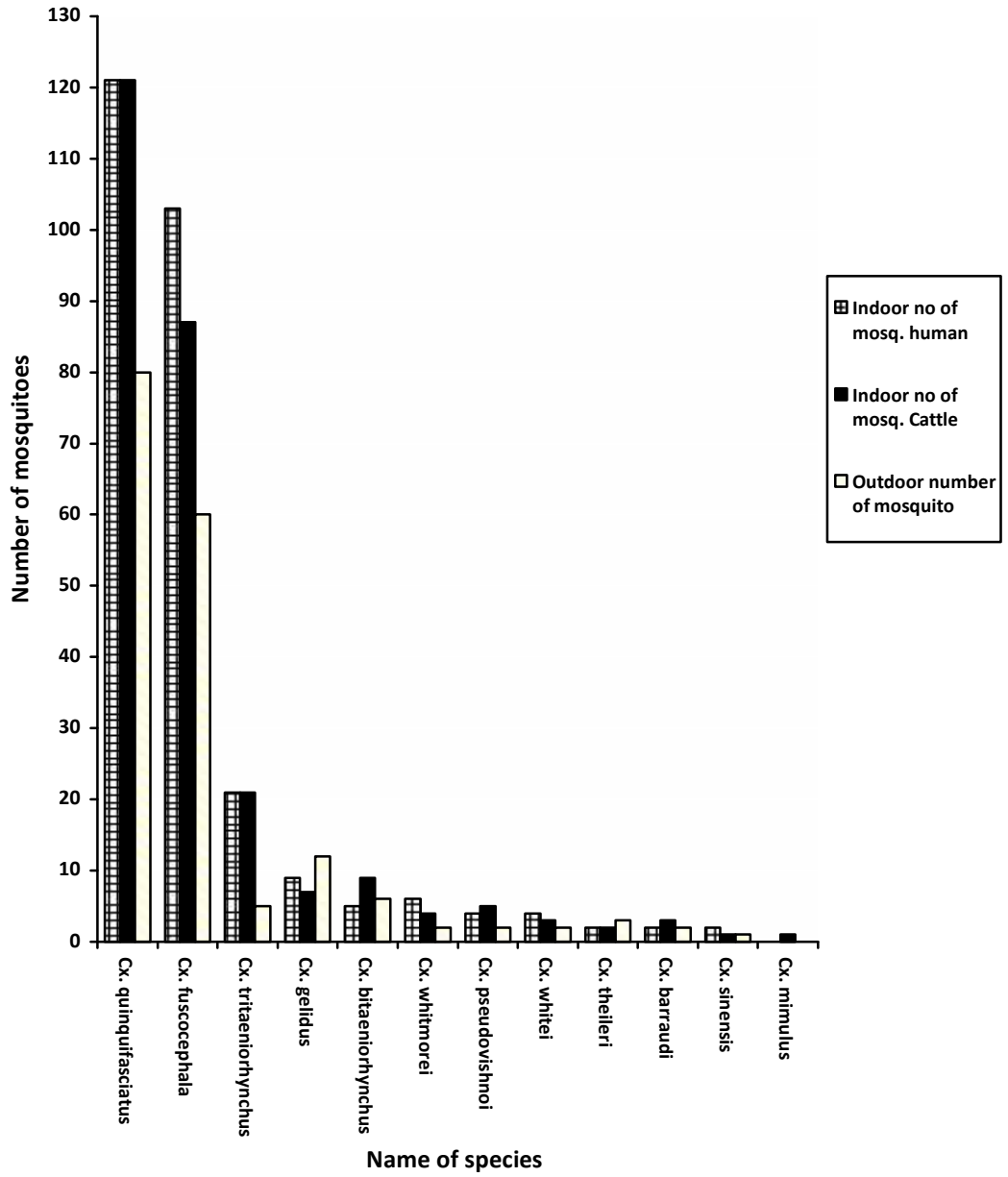
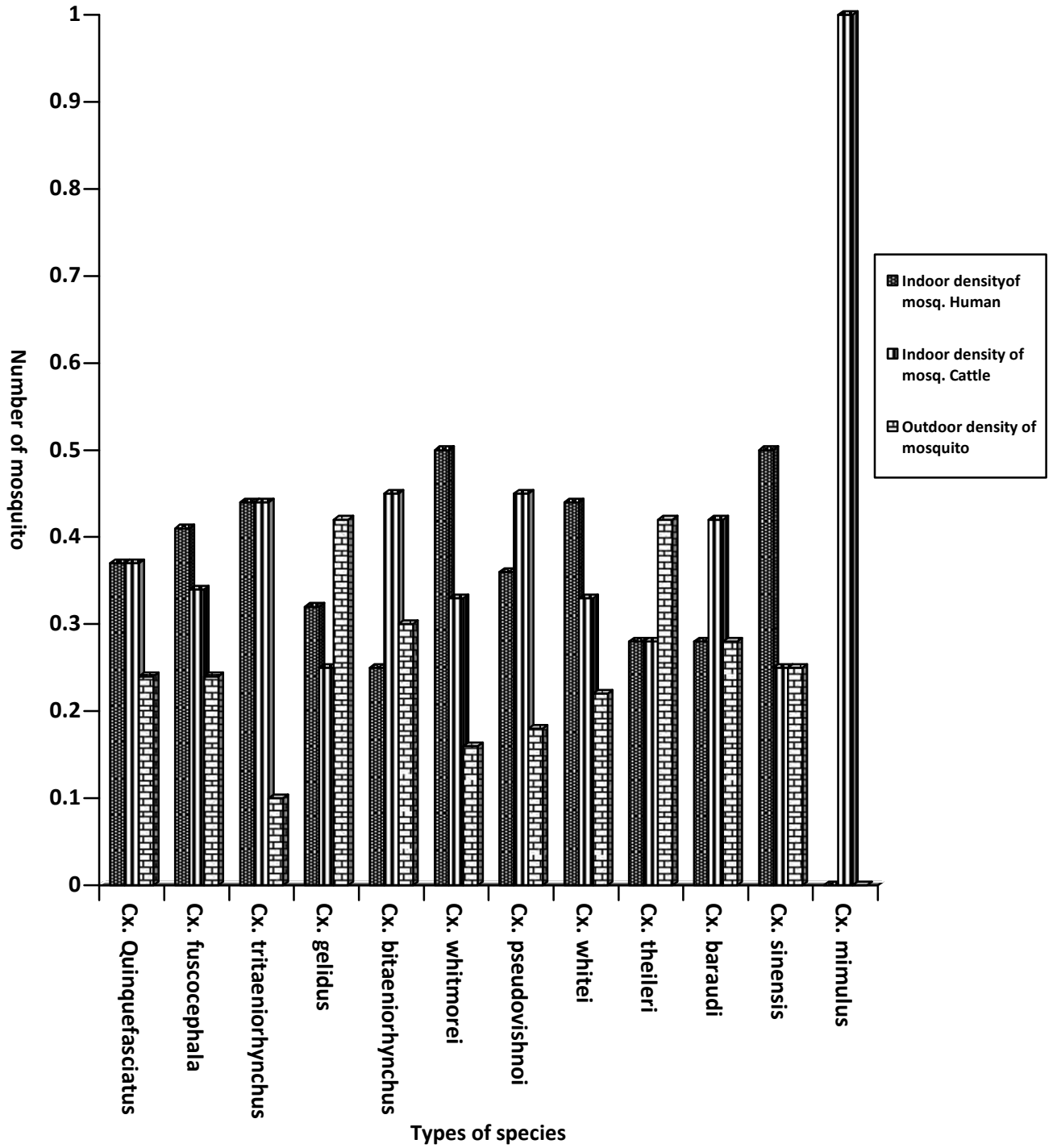


Figure 3
Density of mosquito
indoor and outdoor post monsoon



5.1.2 Pre-monsoon survey

The total of 479 *Culex* mosquitoes belong to five species (five species in indoor and four species in outdoor collection), were recorded during ten days period in March and April. *Culex quinquefasciatus* was the predominant species about 85,106 and 33 samples were recorded from human dwelling, Cattleshed and outdoor collection respectively. Most of the samples (n=226) were recorded from Cattleshed. *Culex bitaeniorhynchus* was least dominant species. *Cx. whitmorei*, *Cx. pseudovishnui*, *Cx. whitei*, *Cx. theileri*, *Cx. barraudi*, *Cx. sinensis*, *Cx. mimulus* were absent. Density of *Cx. tritaeniorhynchus* (0.42 in human dwelling, 0.57 in cattleshed) was highest. In total highest density 2.38 was recorded in Cattleshed. See table 2.

Table 2
Number and Density of Mosquitoes

| Name of species | IN DOOR | | | | OUT DOOR | | TOTAL | |
|------------------------------|----------------------|--------|-----------------------|--------|----------------------|-----------------------|----------------------|-----------------------|
| | Number of Mosquitoes | | Density of Mosquitoes | | Number of Mosquitoes | Density of Mosquitoes | Number of Mosquitoes | Density of Mosquitoes |
| | Human | Cattle | Human | Cattle | | | | |
| <i>Cx. quinquefasciatus</i> | 85 | 106 | 0.37 | 0.47 | 33 | 0.14 | 224 | 0.98 |
| <i>Cx. fuscocephala</i> | 80 | 92 | 0.40 | 0.46 | 26 | 0.13 | 198 | 0.99 |
| <i>Cx. tritaeniorhynchus</i> | 9 | 12 | 0.42 | 0.57 | 0 | 0 | 21 | 0.99 |
| <i>Cx. gelidus</i> | 6 | 7 | 0.33 | 0.38 | 5 | 0.27 | 18 | 0.98 |
| <i>Cx. bitaeniorhynchus</i> | 6 | 9 | 0.33 | 0.5 | 3 | 0.16 | 18 | 0.99 |
| Grand Total | 186 | 226 | 1.85 | 2.38 | 67 | 0.7 | 479 | 4.93 |

Figure 4
Pre-monsoon number of mosquitoes

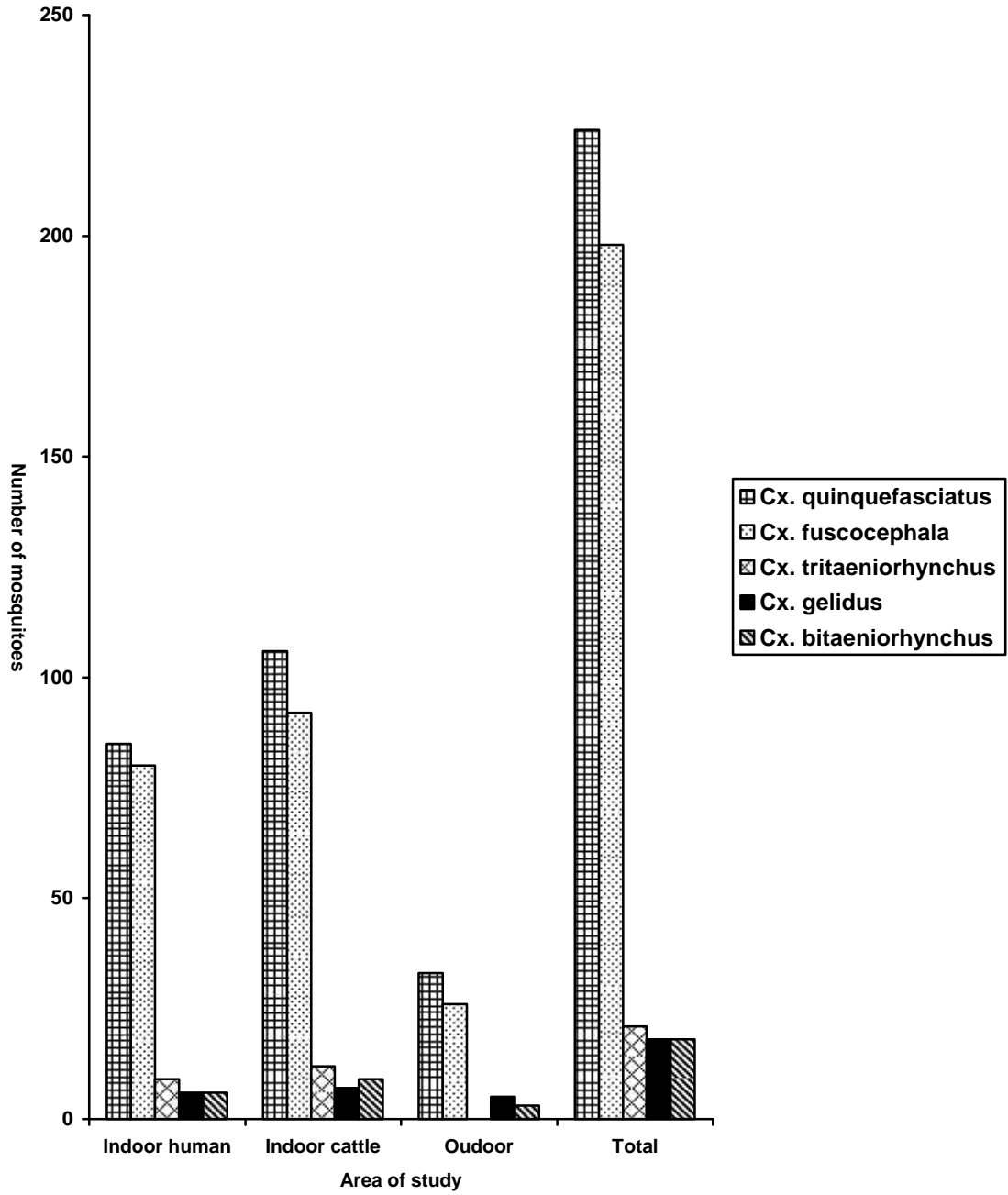
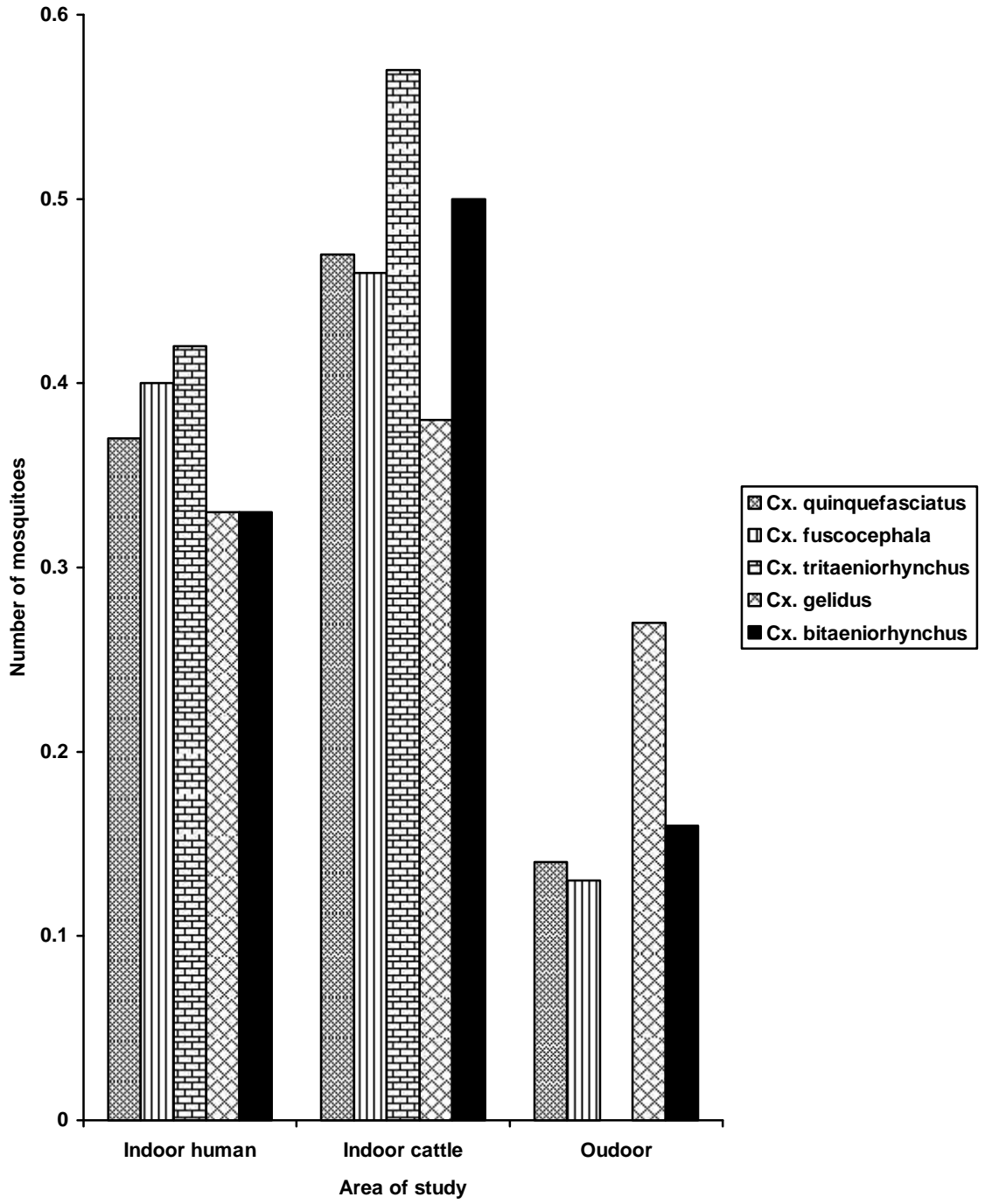


Figure 5
Pre-monsoon density of mosquitoes



5.2 Monthly variations of density of mosquitoes by indoor hand collection and outdoor collections

During the four months, the mosquito samples were highest in September. About 12 species were recorded in September from Bramhapuri village but only two species were recorded in April in same village. *Culex quinquefasciatus* and *Cx. fuscocephala* were widely distributed, they were found in four months of collecting period in both indoor and outdoor hand collection.

Table 3
Density of mosquitoes with Monthly Variation

| | | | IN DOOR | | | | OUT DOOR | |
|------------------------------|--------|--------------|---------|--------|-------|--------|----------------------|-----------------------|
| | | | | | | | Number of Mosquitoes | Density of Mosquitoes |
| Name of Species | Months | Villages | Human | Cattle | Human | Cattle | | |
| <i>Cx. quinquefasciatus</i> | August | Gitanagar | 11 | 17 | 0.15 | 0.23 | 9 | 0.12 |
| <i>Cx. fuscocephala</i> | | | 14 | 9 | 0.19 | 0.19 | 12 | 0.16 |
| <i>Cx. quinquefasciatus</i> | August | Kesharbag | 23 | 19 | 0.28 | 0.23 | 12 | 0.14 |
| <i>Cx. fuscocephala</i> | | | 10 | 6 | 0.12 | 0.07 | 5 | 0.06 |
| <i>Cx. tritaeniorhynchus</i> | | | 3 | 1 | 0.03 | 0.01 | 0 | 0 |
| <i>Cx. bitaeniorhynchus</i> | | | 0 | 2 | 0 | 0.02 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | August | Parashnagar | 12 | 23 | 0.16 | 0.31 | 6 | 0.08 |
| <i>Cx. fuscocephala</i> | | | 18 | 11 | 0.24 | 0.15 | 3 | 0.04 |
| <i>Cx. quinquefasciatus</i> | August | Suryapur | 9 | 10 | 0.24 | 0.27 | 11 | 0.29 |
| <i>Cx. fuscocephala</i> | | | 9 | 7 | 0.24 | 0.18 | 6 | 0.16 |
| <i>Cx. quinquefasciatus</i> | August | Khole- simal | 6 | 7 | 0.16 | 0.18 | 4 | 0.1 |
| <i>Cx. fuscocephala</i> | | | 8 | 5 | 0.21 | 0.13 | 7 | 0.18 |

| | | | | | | | | |
|------------------------------|-------|-------------|----|----|------|------|----|-------|
| <i>Cx. quinquefasciatus</i> | Sept. | Dobato | 12 | 5 | 0.24 | 0.1 | 7 | 0.14 |
| <i>Cx. fuscocephala</i> | | | 7 | 7 | 0.14 | 0.14 | 4 | 0.08 |
| <i>Cx. tritaeniorhynchus</i> | | | 2 | 2 | 0.04 | 0.04 | 1 | 0.02 |
| <i>Cx. bitaeniorhynchus</i> | | | 0 | 2 | 0 | 0.04 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | Sept. | Champanagar | 26 | 22 | 0.22 | 0.18 | 10 | 0.08 |
| <i>Cx. fuscocephala</i> | | | 22 | 17 | 0.18 | 0.14 | 4 | 0.03 |
| <i>Cx. tritaeniorhynchus</i> | | | 5 | 6 | 0.04 | 0.05 | 0 | 0 |
| <i>Cx. gelidus</i> | | | 3 | 0 | 0.02 | 0 | 1 | 0.008 |
| <i>Cx. quinquefasciatus</i> | Sept. | Vijayanagar | 7 | 4 | 0.12 | 0.06 | 6 | 0.1 |
| <i>Cx. fuscocephala</i> | | | 6 | 4 | 0.1 | 0.06 | 4 | 0.06 |
| <i>Cx. tritaeniorhynchus</i> | | | 4 | 2 | 0.06 | 0.03 | 2 | 0.03 |
| <i>Cx. gelidus</i> | | | 1 | 1 | 0.01 | 0.01 | 3 | 0.05 |
| <i>Cx. bitaeniorhynchus</i> | | | 1 | 1 | 0.01 | 0.01 | 2 | 0.03 |
| <i>Cx. whitmorei</i> | | | 1 | 2 | 0.01 | 0.03 | 1 | 0.01 |
| <i>Cx. whitei</i> | | | 1 | 1 | 0.01 | 0.01 | 1 | 0.01 |
| <i>Cx. pseudovishnui</i> | | | 0 | 1 | 0 | 0.01 | 1 | 0.01 |
| <i>Cx. theileri</i> | | | 0 | 0 | 0 | 0 | 1 | 0.01 |
| <i>Cx. quinquefasciatus</i> | Sept. | Rampur | 6 | 13 | 0.09 | 0.2 | 7 | 0.1 |
| <i>Cx. fuscocephala</i> | | | 5 | 8 | 0.07 | 0.12 | 9 | 0.14 |
| <i>Cx. tritaeniorhynchus</i> | | | 1 | 2 | 0.01 | 0.03 | 0 | 0 |
| <i>Cx. gelidus</i> | | | 2 | 2 | 0.03 | 0.03 | 4 | 0.06 |
| <i>Cx. bitaeniorhynchus</i> | | | 1 | 1 | 0.01 | 0.01 | 0 | 0 |
| <i>Cx. pseudovishnui</i> | | | 2 | 1 | 0.03 | 0.01 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | Sept. | Bramhapuri | 6 | 9 | 0.05 | 0.07 | 5 | 0.04 |
| <i>Cx. fuscocephala</i> | | | 7 | 5 | 0.06 | 0.04 | 9 | 0.07 |
| <i>Cx. tritaeniorhynchus</i> | | | 6 | 8 | 0.05 | 0.06 | 2 | 0.01 |
| <i>Cx. gelidus</i> | | | 3 | 4 | 0.02 | 0.03 | 4 | 0.03 |
| <i>Cx. bitaeniorhynchus</i> | | | 3 | 3 | 0.02 | 0.02 | 4 | 0.03 |
| <i>Cx. whitmorei</i> | | | 5 | 2 | 0.04 | 0.01 | 1 | 0.008 |

| | | | | | | | | |
|------------------------------|-------|-------------|----|----|------|-------|---|-------|
| <i>Cx. whitei</i> | | | 3 | 2 | 0.02 | 0.01 | 1 | 0.008 |
| <i>Cx. pseudovishnui</i> | | | 2 | 3 | 0.01 | 0.02 | 1 | 0.008 |
| <i>Cx. barraudi</i> | | | 2 | 3 | 0.01 | 0.02 | 2 | 0.01 |
| <i>Cx. theileri</i> | | | 2 | 2 | 0.01 | 0.01 | 2 | 0.01 |
| <i>Cx. sinensis</i> | | | 2 | 1 | 0.01 | 0.008 | 1 | 0.008 |
| <i>Cx. mimulus</i> | | | 0 | 1 | 0 | 0.008 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | March | Gitanagar | 14 | 8 | 0.25 | 0.14 | 6 | 0.1 |
| <i>Cx. fuscocephala</i> | | | 11 | 9 | 0.19 | 0.16 | 3 | 0.05 |
| <i>Cx. tritaeniorhynchus</i> | | | 1 | 2 | 0.01 | 0.03 | 0 | 0 |
| <i>Cx. bitaeniorhynchus</i> | | | 1 | 1 | 0.01 | 0.01 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | March | Champanagar | 2 | 4 | 0.07 | 0.15 | 0 | 0 |
| <i>Cx. fuscocephala</i> | | | 5 | 9 | 0.19 | 0.34 | 0 | 0 |
| <i>Cx. tritaeniorhynchus</i> | | | 1 | 2 | 0.03 | 0.07 | 0 | 0 |
| <i>Cx. bitaeniorhynchus</i> | | | 0 | 3 | 0 | 0.11 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | March | Parasnagar | 4 | 9 | 0.15 | 0.34 | 2 | 0.07 |
| <i>Cx. fuscocephala</i> | | | 4 | 7 | 0.15 | 0.26 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | March | Kesharbag | 6 | 8 | 0.14 | 0.19 | 1 | 0.02 |
| <i>Cx. fuscocephala</i> | | | 14 | 11 | 0.33 | 0.26 | 0 | 0 |
| <i>Cx. tritaeniorhynchus</i> | | | 2 | 0 | 0.04 | 0 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | April | Vijayanagar | 2 | 11 | 0.03 | 0.19 | 6 | 0.1 |
| <i>Cx. fuscocephala</i> | | | 5 | 12 | 0.08 | 0.21 | 2 | 0.03 |
| <i>Cx. gelidus</i> | | | 5 | 7 | 0.08 | 0.12 | 2 | 0.03 |
| <i>Cx. bitaeniorhynchus</i> | | | 0 | 3 | 0 | 0.05 | 1 | 0.01 |
| <i>Cx.</i> | April | Dobato | 12 | 17 | 0.19 | 0.27 | 4 | 0.06 |

| | | | | | | | | |
|-------------------------------|-------|-------------|----|----|------|------|---|------|
| <i>quinquefasciatus</i> | | | | | | | | |
| <i>Cx. fuscocephala</i> | | | 8 | 3 | 0.13 | 0.04 | 6 | 0.09 |
| <i>Cx. tritaeniorhynchus.</i> | | | 5 | 3 | 0.08 | 0.04 | 0 | 0 |
| <i>Cx. bitaeniorhynchus</i> | | | 2 | 1 | 0.03 | 0.01 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | April | Rampur | 10 | 6 | 0.23 | 0.14 | 2 | 0.04 |
| <i>Cx. fuscocephal</i> | | | 6 | 9 | 0.14 | 0.21 | 0 | 0 |
| <i>Cx. gelidus</i> | | | 3 | 1 | 0.07 | 0.02 | 0 | 0 |
| <i>Cx. bitaeniorhynchus</i> | | | 3 | 0 | 0.07 | 0 | 2 | 0.04 |
| <i>Cx. quinquefasciatus</i> | April | Suryapur | 15 | 27 | 0.19 | 0.34 | 2 | 0.02 |
| <i>Cx. fuscocephala</i> | | | 10 | 17 | 0.12 | 0.21 | 1 | 0.01 |
| <i>Cx. tritaeniorhynchus</i> | | | 0 | 5 | 0 | 0.06 | 0 | 0 |
| <i>Cx. bitaeniorhynchus</i> | | | 0 | 1 | 0 | 0.01 | 0 | 0 |
| <i>Cx. quinquefasciatus</i> | April | Khole-simal | 11 | 9 | 0.22 | 0.18 | 6 | 0.12 |
| <i>Cx. fuscocephala</i> | | | 6 | 7 | 0.12 | 0.14 | 9 | 0.18 |
| <i>Cx. quinquefasciatus</i> | April | Bramhapuri | 9 | 7 | 0.2 | 0.15 | 4 | 0.09 |
| <i>Cx. fuscocephala</i> | | | 11 | 8 | 0.26 | 0.18 | 5 | 0.11 |

5.3 Vector Abundance

5.3.1 Species composition: Percent abundance

The percent abundance of Mosquitoes is calculated as,

Percent Abundance: $\frac{\text{Number of Collected Mosquitoes (SPP)}}{\text{Total Number of Mosquitoes}} \times 100\%$

Total Number of Mosquitoes

(For Vector abundance and species composition)

Culex quinquefasciatus is the most abundant in post and pre monsoon, about 322 i.e, 26.90 percent found in post monsoon and about 224 i.e., 18.71 percent found in pre monsoon. *Cx. mimulus* is the least abundant in both

season about one i.e, 0.08 percent found in post monsoon and zero in pre monsoon.

The percent abundance of six species were found less than one percent in post-monsoon and the abundance of seven species is zero in pre monsoon.

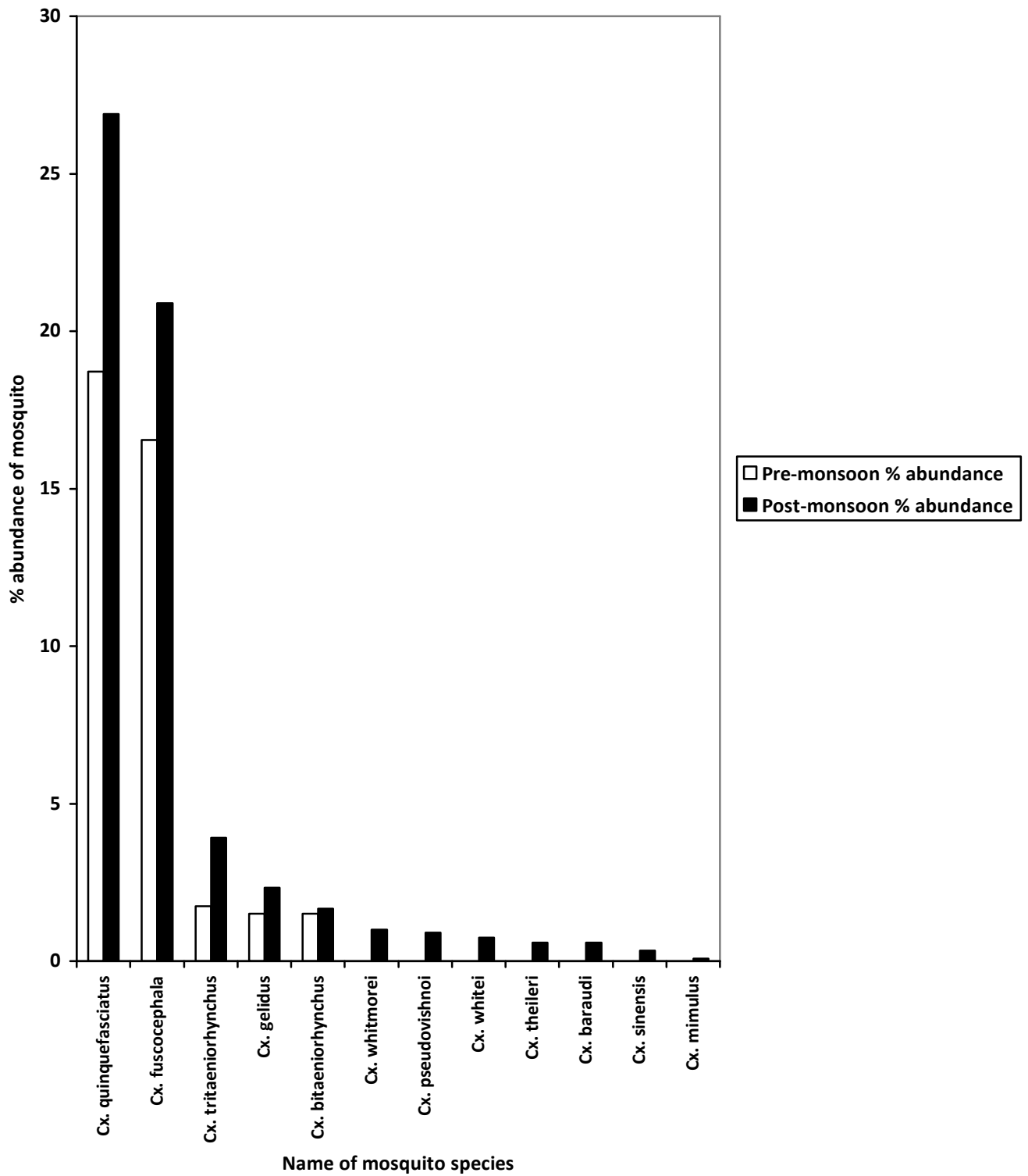
The percent abundance of *Cx. tritaeniorhynchus* in post-monsoon and pre monsoon is 3.92 percent and 1.75 percent respectively which is potential vector of JE in Nepal. The percent abundance of *Cx. fuscocephala* in post-monsoon and pre-monsoon is 20.88 percent and 16.54 percent respectively. See table 4.

Table 4
Percent Abundance of Mosquito species

| Name of Species | Post-Monsoon | | Pre-Monsoon | |
|------------------------------|--------------------------------|-------------------|--------------------------------|-------------------|
| | Number of Collected Mosquitoes | Percent abundance | Number of Collected Mosquitoes | Percent abundance |
| <i>Cx. quinquefasciatus</i> | 322 | 26.90 | 224 | 18.71 |
| <i>Cx. fuscocephala</i> | 250 | 20.88 | 198 | 16.54 |
| <i>Cx. tritaeniorhynchus</i> | 47 | 3.92 | 21 | 1.75 |
| <i>Cx. gelidus</i> | 28 | 2.33 | 18 | 1.50 |
| <i>Cx. bitaeniorhynchus</i> | 20 | 1.67 | 18 | 1.50 |
| <i>Cx. whitimorei</i> | 12 | 1.00 | 0 | 0 |
| <i>Cx. pseudovishnui</i> | 11 | 0.91 | 0 | 0 |
| <i>Cx. whitei</i> | 9 | 0.75 | 0 | 0 |
| <i>Cx. barraudi</i> | 7 | 0.58 | 0 | 0 |
| <i>Cx. theileri</i> | 7 | 0.58 | 0 | 0 |
| <i>Cx. sinensis</i> | 4 | 0.33 | 0 | 0 |
| <i>Cx. mimulus</i> | 1 | 0.08 | 0 | 0 |

Figure 6

Graphical presentation of Percent Abundance of Mosquito Species



5.3.2 Indoor resting Density (Human)

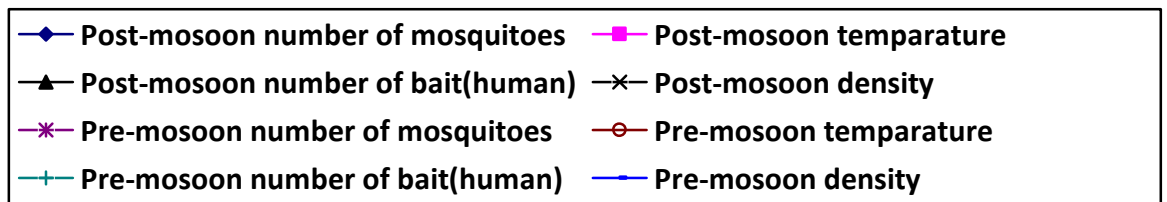
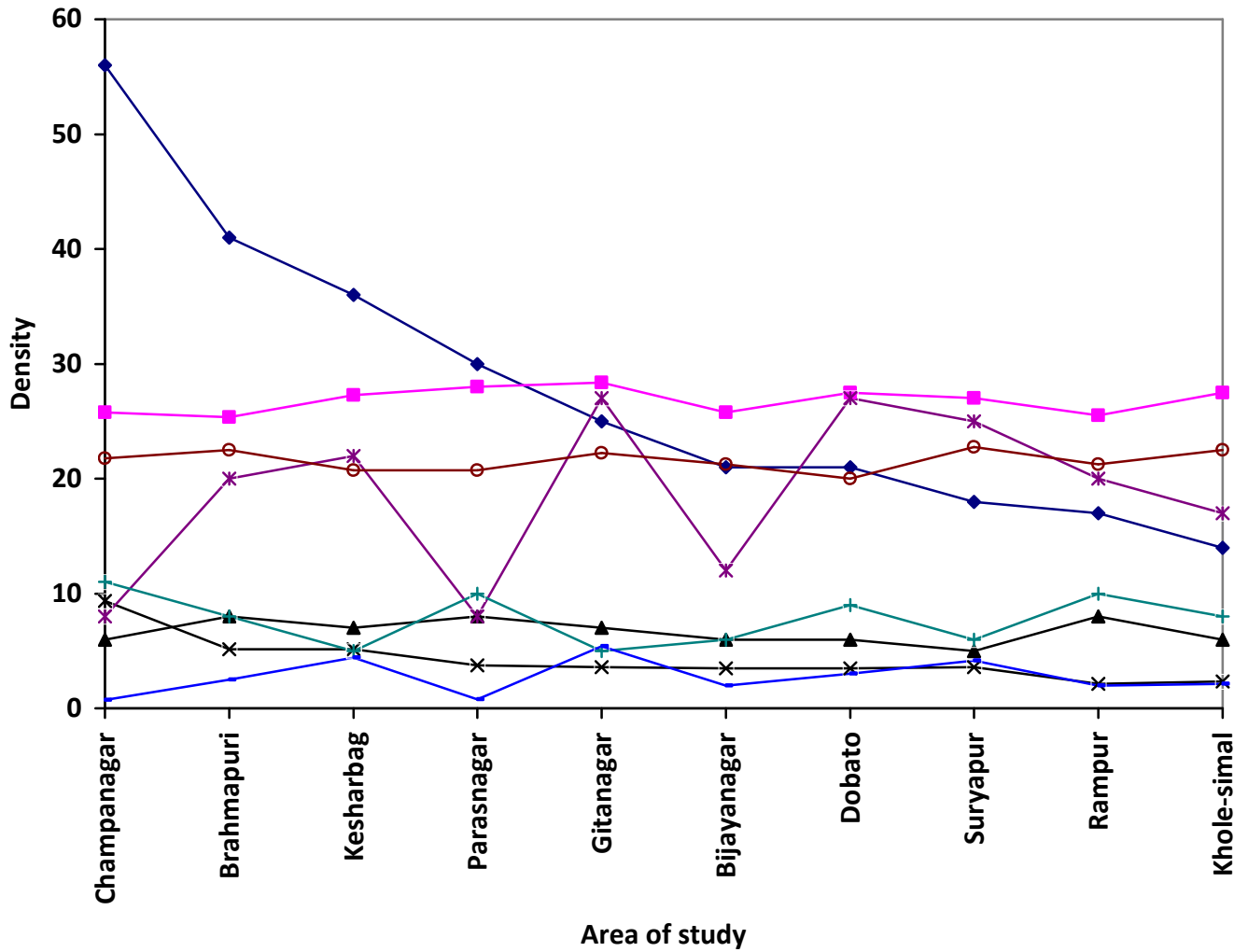
The indoor resting Density of Mosquitoes sample is calculated as,
 Indoor resting Density: $\frac{\text{Number of Mosquitoes}}{\text{Number of bait (Human)}}$

Highest indoor resting density was 9.33 at 25.75°C in post-monsoon and lowest indoor density 0.72 at 21.75°C in pre-monsoon reported in Champanagar village. See table 5.

Table 5
Indoor resting Density (Human)

| Name of village | Post- Monsoon | | | | Pre -Monsoon | | | |
|-----------------|----------------------|----------------------|------------------------|---------|----------------------|----------------------|------------------------|---------|
| | Number of Mosquitoes | Temp. ⁰ c | Number of bait (human) | Density | Number of Mosquitoes | Temp. ⁰ c | Number of bait (human) | Density |
| Champanagar | 56 | 25.75 | 6 | 9.33 | 8 | 21.75 | 11 | 0.72 |
| Brahmapuri | 41 | 25.37 | 8 | 5.12 | 20 | 22.5 | 8 | 2.5 |
| Kesharbag | 36 | 27.25 | 7 | 5.14 | 22 | 20.75 | 5 | 4.4 |
| Parashnagar | 30 | 28 | 8 | 3.75 | 8 | 20.75 | 10 | 0.8 |
| Gitanagar | 25 | 28.37 | 7 | 3.57 | 27 | 22.25 | 5 | 5.4 |
| Bijayanagar | 21 | 25.75 | 6 | 3.5 | 12 | 21.25 | 6 | 2 |
| Dobato | 21 | 27.5 | 6 | 3.5 | 27 | 20 | 9 | 3 |
| Suryapur | 18 | 27 | 5 | 3.6 | 25 | 22.75 | 6 | 4.16 |
| Rampur | 17 | 25.5 | 8 | 2.12 | 20 | 21.25 | 10 | 2 |
| Khole-simal | 14 | 27.5 | 6 | 2.33 | 17 | 22.5 | 8 | 2.12 |

Figure 7
Graphical presentation of Indoor Resting Density (Human)



5.3.3 Indoor resting Density (Cattle)

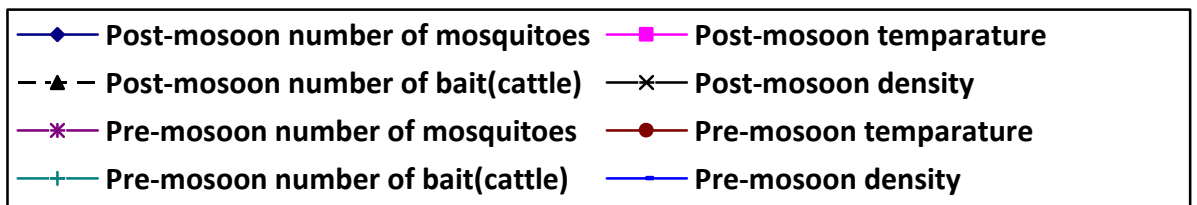
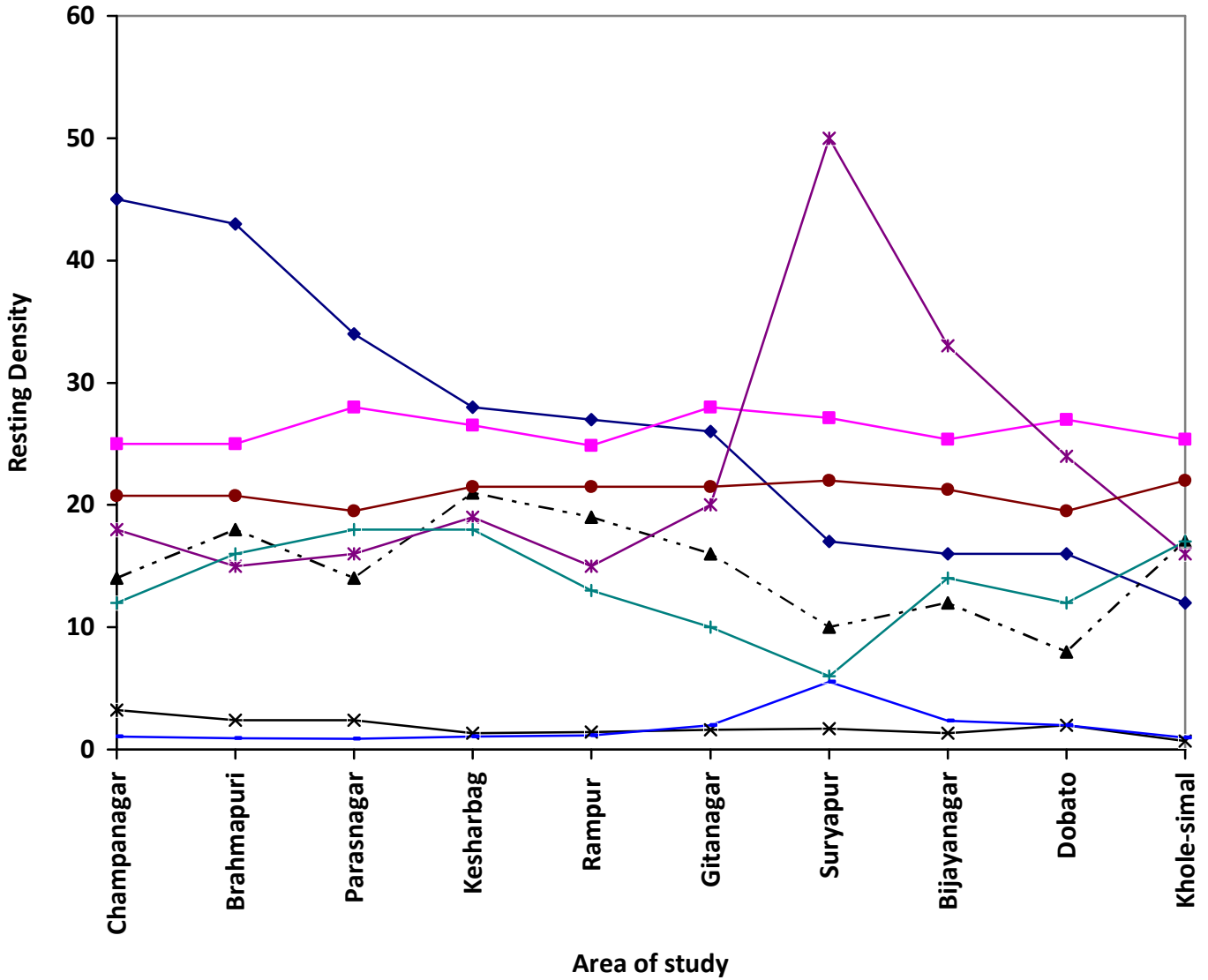
The indoor resting Density of Mosquitoes sample is calculated as,
 Indoor resting Density = $\frac{\text{Number of Mosquitoes}}{\text{Number of bait (cattle)}}$

Highest indoor resting density was 5.55 at 22°C in Suryapur in pre-monsoon and lowest indoor resting density was 0.70 at 25.37°C in Kholesimal in post-monsoon. See table 6.

Table 6
Indoor resting Density (Cattle)

| Name of village | Post -Monsoon | | | | Pre -Monsoon | | | |
|-----------------|-----------------------|----------------------|-------------------------|---------|----------------------|----------------------|-------------------------|---------|
| | Number. of Mosquitoes | Temp. ⁰ c | Number of bait (Cattle) | Density | Number of Mosquitoes | Temp. ⁰ c | Number of bait (Cattle) | Density |
| Champanagar | 45 | 25 | 14 | 3.21 | 18 | 20.75 | 12 | 105 |
| Brahmapuri | 43 | 25 | 18 | 2.38 | 15 | 20.75 | 16 | 0.93 |
| Parashnagar | 34 | 28 | 14 | 2.42 | 16 | 19.5 | 18 | 0.88 |
| Kesharbag | 28 | 26.5 | 21 | 1.33 | 19 | 21.5 | 18 | 1.05 |
| Rampurr | 27 | 24.87 | 19 | 1.42 | 15 | 21.5 | 13 | 1.15 |
| Gitanagar | 26 | 28 | 16 | 1.62 | 20 | 21.5 | 10 | 2 |
| Suryapur | 17 | 27.12 | 10 | 1.7 | 50 | 22 | 9 | 5.55 |
| Bijayanagar | 16 | 25.37 | 12 | 1.33 | 33 | 21.25 | 14 | 2.35 |
| Dobato | 16 | 27 | 8 | 2 | 24 | 19.5 | 12 | 2 |
| Khole-simal | 12 | 25.37 | 17 | 0.70 | 16 | 22 | 17 | 0.95 |

Figure 8
Graphical presentation of Indoor Resting Density (Cattle)



5.3.4 Outdoor Density

The Outdoor Density of Mosquitoes samples is calculated as,

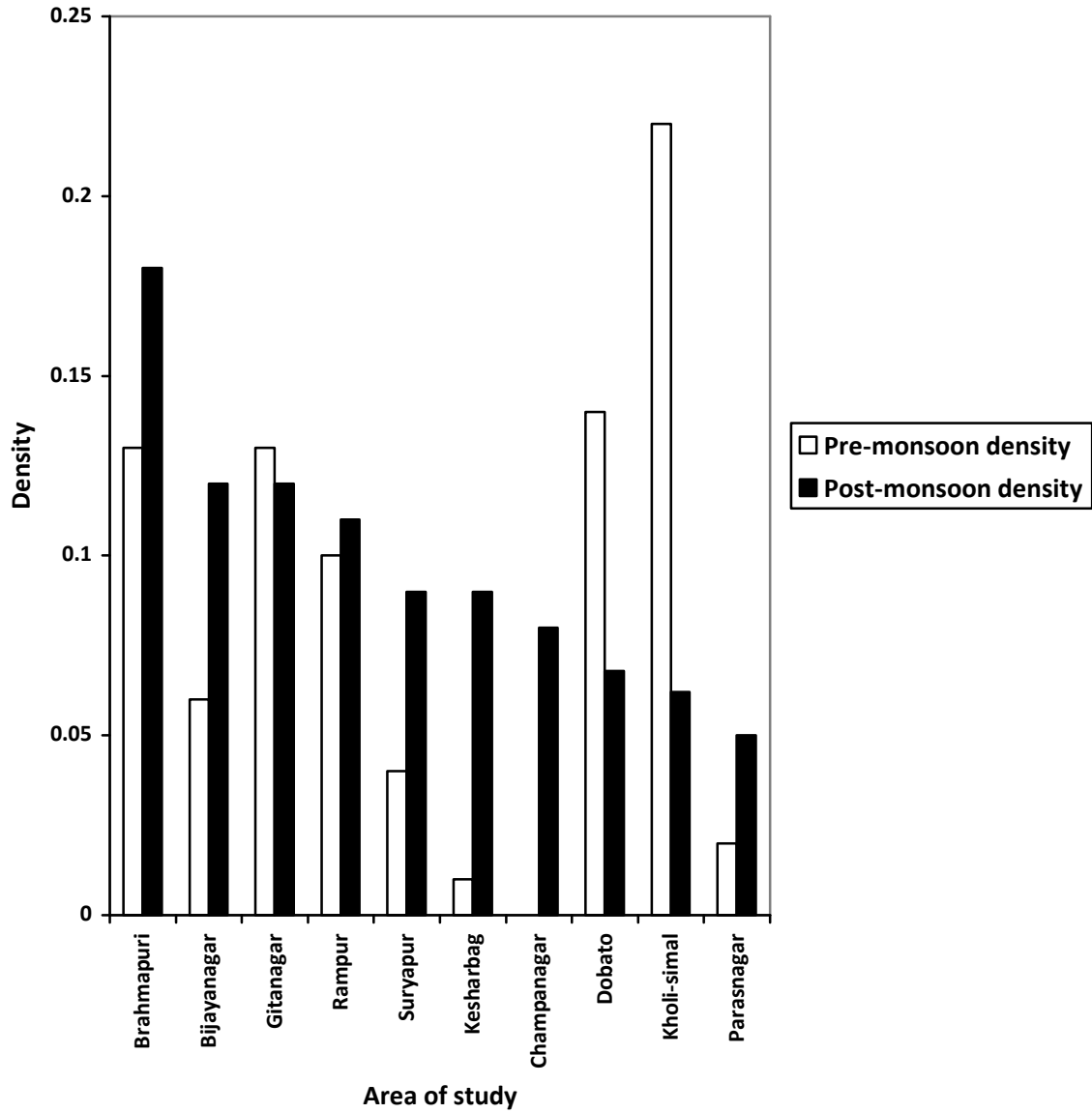
$$\text{Outdoor Density} = \frac{\text{Number of Mosquitoes}}{\text{Total number of Mosquitoes}}$$

Both Highest and Lowest outdoor density was found in pre-monsoon, highest 0.22 in Kholesimal at 22.5°C and lowest 0 at 21.5°C temperature in Champanagar village. See table 7.

Table 7
Outdoor Density of Mosquitoes

| Name of village | Post-Monsoon | | | Pre-Monsoon | | |
|-----------------|----------------------|----------|---------|----------------------|----------|---------|
| | Number of Mosquitoes | Temp. °C | Density | Number of Mosquitoes | Temp. °C | Density |
| Bramhapuri | 32 | 25.5 | 0.18 | 9 | 22 | 0.13 |
| Bijayanagar | 21 | 26.12 | 0.12 | 11 | 22.25 | 0.06 |
| Gitanagar | 21 | 29 | 0.12 | 9 | 22.25 | 0.13 |
| Rampur | 20 | 25.37 | 0.11 | 7 | 22.5 | 0.10 |
| Suryapur | 17 | 28.5 | 0.09 | 3 | 22.25 | 0.04 |
| Kesharbag | 17 | 26.75 | 0.09 | 1 | 22 | 0.01 |
| Champanagar | 15 | 25.62 | 0.08 | 0 | 21.5 | 0 |
| Dobato | 12 | 26.75 | 0.068 | 10 | 20.25 | 0.14 |
| Khole-simal | 11 | 26.12 | 0.062 | 15 | 22.5 | 0.22 |
| Parashnagar | 9 | 29 | 0.05 | 2 | 19.75 | 0.02 |
| Total | 175 | | | 67 | | |

Figure 9
Graphical Presentation of Outdoor Density of Mosquitoes



5.3.5 Resting Habit

The resting habit of Mosquitoes of each species is the ratio of number of mosquitoes of each species in indoor collection and number of mosquitoes of each species in outdoor collection.

Resting Habit: $\frac{\text{Number of Mosquitoes in indoor collection}}{\text{Number of Mosquitoes in outdoor collection}}$

Number of Mosquitoes in outdoor collection

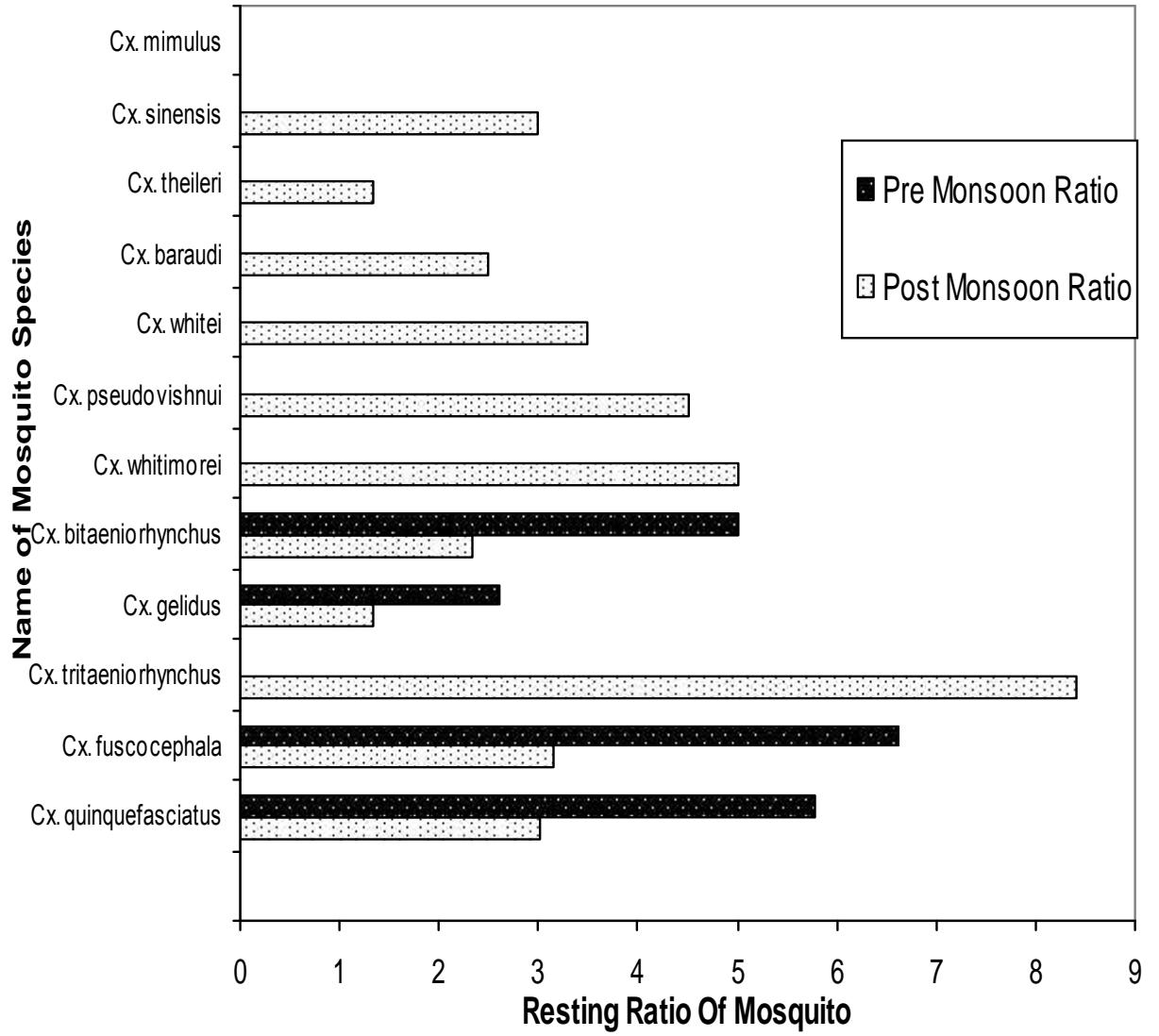
The resting habit of *Culex mimulus* and *Cx. tritaeniorhynchus* was infinite in post-monsoon and pre-monsoon respectively. The resting habit of *Cx. whitimorei*, *Cx. pseudovishnui*, *Cx. whitei*, *Cx. barraudi*, *Cx. theileri*, *Cx. sinensis* and *Cx. mimulus* is zero in pre-monsoon.

Table 8
Resting Habit of Mosquito Species

| Name of species | Post-Monsoon | | | Pre-Monsoon | | |
|------------------------------|----------------------|---------|---------------|----------------------|---------|---------------|
| | Number of Mosquitoes | | Resting Habit | Number of Mosquitoes | | Resting Habit |
| | Indoor | Outdoor | | Indoor | Outdoor | |
| <i>Cx. quinquefasciatus</i> | 242 | 80 | 3.01 | 191 | 33 | 5.78 |
| <i>Cx. fuscocephala</i> | 190 | 60 | 3.16 | 172 | 26 | 6.61 |
| <i>Cx. tritaeniorhynchus</i> | 42 | 5 | 8.4 | 21 | 0 | |
| <i>Cx. gelidus</i> | 16 | 12 | 1.33 | 13 | 5 | 2.6 |
| <i>Cx. bitaeniorhynchus</i> | 14 | 6 | 2.33 | 15 | 3 | 5 |
| <i>Cx. whitimorei</i> | 10 | 2 | 5 | 0 | 0 | 0 |
| <i>Cx. pseudovishnui</i> | 9 | 2 | 4.5 | 0 | 0 | 0 |
| <i>Cx. whitei</i> | 7 | 2 | 3.5 | 0 | 0 | 0 |
| <i>Cx. barraudi</i> | 5 | 2 | 2.5 | 0 | 0 | 0 |
| <i>Cx. theileri</i> | 4 | 3 | 1.33 | 0 | 0 | 0 |
| <i>Cx. sinensis</i> | 3 | 1 | 3 | 0 | 0 | 0 |
| <i>Cx. mimulus</i> | 1 | 0 | | 0 | 0 | 0 |

Figure 10

Graphical Presentation of Resting Habit of Mosquito Species



5.3.6 Room Density

The room density of mosquitoes is calculated as,
 Room Density = $\frac{\text{Number of Mosquitoes (each species)}}{\text{Number of Room}}$

The highest room density was (i.e. 6.5) of *Culex quinquefasciatus* in Champanagar village. In both season (pre-monsoon and post-monsoon), same (i.e.3.75) room density of *Cx. quinquefasciatus* was in Suryapur village. Room density of *Cx. bitaeniorhynchus* in Dobato and Gitanagar and *Cx. tritaeniorhynchus* in Gitanagar is zero in post-monsoon. In pre-monsoon room density of *Cx.tritaeniorhynchus*, *Cx. gelidus*, *Cx. bitaeniorhynchus*, *Cx. whitimorei*, *Cx. whitei*, *Cx. pseudovishnui*, *Cx. barraudi*, *Cx. theileri* and *Cx. sinensis* is zero. See table 9.

Table 9
Room Density of Mosquitoes Species

| Name of species | Name of village | Post-Monsoon | | Pre-Monsoon | |
|------------------------------|-----------------|----------------------|---------|----------------------|---------|
| | | Number of Mosquitoes | Density | Number of Mosquitoes | Density |
| <i>Cx. quinquefasciatus</i> | Gitanagar | 14 | 3.5 | 14 | 3.5 |
| <i>Cx. fuscocephala</i> | | 11 | 2.75 | 11 | 2.75 |
| <i>Cx. tritaeniorhynchus</i> | | - | - | 1 | 0.25 |
| <i>Cx. bitaeniorhynchus</i> | | - | - | 1 | 0.25 |
| <i>Cx. quinquefasciatus</i> | Kesharbag | 23 | 5.75 | 6 | 1.5 |
| <i>Cx. fuscocephala</i> | | 10 | 2.5 | 14 | 3.5 |
| <i>Cx. tritaeniorhynchus</i> | | 3 | 0.75 | 2 | 0.5 |

| | | | | | |
|------------------------------|-------------|----|------|----|------|
| <i>Cx. quinquefasciatus.</i> | Parashnagar | 12 | 3 | 2 | 0.5 |
| <i>Cx. fuscocephala</i> | | 18 | 4.5 | - | - |
| <i>Cx. quinquefasciatus.</i> | Suryapur | 9 | 2.25 | 15 | 3.75 |
| <i>Cx. fuscocephala</i> | | 9 | 2.25 | 10 | 2.5 |
| <i>Cx. quinquefasciatus.</i> | Khole-simal | 6 | 1.5 | 11 | 2.75 |
| <i>Cx. fuscocephala</i> | | 8 | 2 | 6 | 1.5 |
| <i>Cx. quinquefasciatus</i> | Dobato | 12 | 3 | 12 | 3 |
| <i>Cx. fuscocephala</i> | | 7 | 1.75 | 8 | 2 |
| <i>Cx. tritaeniorhynchus</i> | | 2 | 0.5 | 5 | 1.25 |
| <i>Cx. bitaeniorhynchus</i> | | - | - | 2 | 0.5 |
| <i>Cx. quinquefasciatus.</i> | Champanagar | 26 | 6.5 | 2 | 0.5 |
| <i>Cx. fuscocephala</i> | | 22 | 5.5 | 5 | 1.25 |
| <i>Cx. tritaeniorhynchus</i> | | 5 | 0.25 | 1 | 0.25 |
| <i>Cx. gelidus</i> | | 3 | 0.75 | - | - |
| <i>Cx. quinquefasciatus.</i> | Vijayanagar | 7 | 1.75 | 2 | 0.5 |
| <i>Cx. fuscocephala</i> | | 6 | 1.5 | 5 | 1.25 |
| <i>Cx. tritaeniorhynchus</i> | | 4 | 1 | - | - |
| <i>Cx. gelidus</i> | | 1 | 0.25 | 5 | 1.25 |
| <i>Cx. bitaeniorhynchus</i> | | 1 | 0.25 | - | - |
| <i>Cx. whitmorei</i> | | 1 | 0.25 | - | - |
| <i>Cx. whitei</i> | | 1 | 0.25 | - | - |
| <i>Cx. quinquefasciatus.</i> | Rampur | 6 | 1.5 | 2 | 0.5 |
| <i>Cx. fuscocephala</i> | | 5 | 1.25 | - | - |
| <i>Cx. tritaeniorhynchus</i> | | 1 | 0.25 | - | - |
| <i>Cx. gelidus</i> | | 2 | 0.5 | - | - |
| <i>Cx.</i> | | 1 | 0.25 | 2 | 0.5 |

| | | | | | |
|------------------------------|------------|---|------|----|------|
| <i>bitaeniorhynchus</i> | | | | | |
| <i>Cx.pseudovishnui</i> | | 2 | 0.5 | - | - |
| <i>Cx. quinquefasciatus</i> | Bramhapuri | 6 | 1.5 | 9 | 0.25 |
| <i>Cx. fuscocephala</i> | | 7 | 1.75 | 11 | 2.75 |
| <i>Cx. tritaeniorhynchus</i> | | 6 | 1.5 | - | - |
| <i>Cx. gelidus</i> | | 3 | 0.75 | - | - |
| <i>Cx. bitaeniorhynchus</i> | | 3 | 0.75 | - | - |
| <i>Cx. whitmorei</i> | | 5 | 1.25 | - | - |
| <i>Cx. whitei</i> | | 3 | 0.75 | - | - |
| <i>Cx. pseudovishnui</i> | | 2 | 0.5 | - | - |
| <i>Cx. barraudi</i> | | 2 | 0.5 | - | - |
| <i>Cx. theileri</i> | | 2 | 0.5 | - | - |
| <i>Cx. sinensis</i> | | 2 | 0.5 | - | - |

5.4 Temperature versus Month

The total of 12 species were found at 25.26°C in September in Bramhapuri village and only two species (i.e. *Culex quinquefasciatus* and *Cx. fuscocephala*) were found at 28.33°C in August, at 20°C in March in Parasnagar village and at 26.33°C in August, at 22.33°C in April in Kholesimal village. See table 10.

Table 10
Number and Density of Mosquito with respect to Temperature

| Name of species | Name of village | Post-Monsoon | | | | Pre-Monsoon | | | |
|------------------------------|-----------------|--------------|-----------------|---------|----------|-------------|-----------------|---------|----------|
| | | Month | Number of Mosq. | Density | Temp. °c | Month | Number of Mosq. | Density | Temp. °c |
| <i>Cx. quinquefasciatus</i> | Gitanagar | August | 35 | 0.48 | 28.45 | March | 28 | 0.5 | 22 |
| <i>Cx. fuscocephala</i> | | | 37 | 0.51 | | | 23 | 0.41 | |
| <i>Cx. tritaeniorhynchus</i> | | | - | - | | | 3 | 0.05 | |
| <i>Cx. bitaeniorhynchus</i> | | | - | - | | | 2 | 0.03 | |
| <i>Cx. quinquefasciatus</i> | Kesharbag | August | 54 | 0.66 | 26.83 | | 15 | 0.35 | 21.41 |
| <i>Cx. fuscocephala</i> | | | 21 | 0.25 | | March | 25 | 0.83 | |
| <i>Cx. tritaeniorhynchus</i> | | | 4 | 0.04 | | | 2 | 0.04 | |
| <i>Cx. bitaeniorhynchus</i> | | | 2 | 0.02 | | | - | - | |
| <i>Cx. quinquefasciatus</i> | Parashnagar | August | 41 | 0.65 | 28.39 | March | 15 | 0.57 | 20 |
| <i>Cx. fuscocephala</i> | | | 32 | 0.43 | | | 11 | 0.42 | |
| <i>Cx. quinquefasciatus</i> | Suryapur | August | 30 | 0.57 | 27.54 | April | 44 | 0.56 | 22.33 |
| <i>Cx. fuscocephala</i> | | | 22 | 0.42 | | | 28 | 0.35 | |
| <i>Cx. tritaeniorhynchus</i> | | | - | - | | | 5 | 0.06 | |
| <i>Cx. bitaeniorhynchus</i> | | | - | - | | | 1 | 0.01 | |
| <i>Cx. quinquefasciatus</i> | Khole-simal | August | 17 | 0.45 | 26.33 | April | 26 | 0.54 | 22.33 |
| <i>Cx. fuscocephala</i> | | | 20 | 0.54 | | | 22 | 0.45 | |
| <i>Cx. quinquefasciatus</i> | Dobato | Sept. | 24 | 0.48 | 27.08 | April | 33 | 0.54 | 19.91 |
| <i>Cx. fuscocephala</i> | | | 18 | 0.36 | | | 17 | 0.27 | |
| <i>Cx.</i> | | | 5 | 0.10 | | | 8 | 0.13 | |

| | | | | | | | | | |
|--------------------------------|-------------|-------|----|------|-------|-------|----|------|-------|
| <i>tritaeniorhynchus</i> | | | | | | | | | |
| <i>Cx. bitaeniorhynchus</i> | | | 2 | 0.04 | | | 3 | 0.04 | |
| <i>Cx. quinquefasciatus</i> | Champanagar | Sept. | 58 | 0.5 | 25.42 | March | 6 | 0.23 | 21.33 |
| <i>Cx. fuscocephala</i> | | | 43 | 0.37 | | | 14 | 0.53 | |
| <i>Cx. tritaeniorhynchus</i> | | | 11 | 0.09 | | | 3 | 0.11 | |
| <i>Cx. gelidus</i> | | | 4 | 0.03 | | | 3 | 0.11 | |
| <i>Cx. quinquefasciatus</i> | Vijayanagar | Sept. | 17 | 0.29 | 25.74 | April | 19 | 0.33 | 21.58 |
| <i>Cx. fuscocephala</i> | | | 14 | 0.24 | | | 19 | 0.33 | |
| <i>Cx. tritaeniorhynchus</i> | | | 8 | 0.13 | | | - | - | |
| <i>Cx. gelidus</i> | | | 5 | 0.08 | | | 14 | 0.25 | |
| <i>Cx. bitaeniorhynchus</i> | | | 4 | 0.06 | | | 4 | 0.07 | |
| <i>Cx. whitmorei</i> | | | 4 | 0.06 | | | - | - | |
| <i>Cx. whitei</i> | | | 3 | 0.05 | | | - | - | |
| <i>Cx. pseudovishnui</i> | | | 2 | 0.03 | | | - | - | |
| <i>Cx. theileri</i> | | | 1 | 0.01 | | | - | - | |
| <i>Cx. quinquefasciatus</i> | Rampur | Sept. | 26 | 0.40 | 25.24 | April | 18 | 0.42 | 21.75 |
| <i>Cx. fuscocephala</i> | | | 22 | 0.34 | | | 15 | 0.35 | |
| <i>Cx. tritaeniorhynchus h</i> | | | 3 | 0.04 | | | - | - | |
| <i>Cx. gelidus</i> | | | 8 | 0.12 | | | 4 | 0.09 | |
| <i>Cx. bitaeniorhynchus</i> | | | 2 | 0.03 | | | 5 | 0.11 | |
| <i>Cx. pseudovishnui</i> | | | 3 | 0.04 | | | - | - | |
| <i>Cx. quinquefasciatus</i> | Bramhapuri | Sept. | 20 | 0.17 | 25.26 | April | 20 | 0.45 | 21.75 |
| <i>Cx. fuscocephala</i> | | | 21 | 0.18 | | | 24 | 0.54 | |
| <i>Cx. tritaeniorhynchus</i> | | | 16 | 0.13 | | | - | - | |

| | | | | | | | | | |
|-----------------------------|--|--|----|-------|--|--|---|---|--|
| <i>Cx. gelidus</i> | | | 11 | 0.09 | | | - | - | |
| <i>Cx. bitaeniorhynchus</i> | | | 10 | 0.08 | | | - | - | |
| <i>Cx. whitmorei</i> | | | 8 | 0.06 | | | - | - | |
| <i>Cx. whitei</i> | | | 6 | 0.05 | | | - | - | |
| <i>Cx. pseudovishnui</i> | | | 6 | 0.06 | | | - | - | |
| <i>Cx. barraudi</i> | | | 7 | 0.07 | | | - | - | |
| <i>Cx. theileri</i> | | | 6 | 0.05 | | | - | - | |
| <i>Cx. sinensis</i> | | | 4 | 0.03 | | | - | - | |
| <i>Cx. mimulus</i> | | | 1 | 0.008 | | | - | - | |

CHAPTER VI

DISCUSSION AND COMMENTS

6.1 Discussion

The present study was conducted from August 2007 to September 2007 and March 2008 to April 2008 in Chitwan district of the Country. In this study, an attempt was made to determine the distribution of different *Culex* mosquito species on ten villages of Chitwan district, the species present in the study area and their relative density in indoor and outdoor, to assess seasonal changes, to determine vector abundance and species composition in the study area and to determine resting habit of mosquito species. The total of 1197 sample was collected during the study period and they were identified in the Lab. About 718 and 479 sample were recorded from post-monsoon and pre-monsoon collection respectively. In both post-monsoon and pre-monsoon *Culex quinquefasciatus* was abundant species. The total of twelve species was reported in post-monsoon and only five species in pre-monsoon. Five species of post-monsoon and four species of pre-monsoon were suspected vector of different mosquito born disease. Three suspected vector of JE i.e. *Culex tritaeniorhynchus*, *Cx. fuscocephala*, *Cx. gelidus*, and one principal vector of Filariasis i.e. *Cx. quinquefasciatus* and one suspected vector of West-Nile virus i.e., *Cx. theileri* were recorded.

In the overall present study about twelve *Culex* species were recorded in cattle shed; eleven species from human dwelling and outdoor collection in post monsoon and in pre monsoon about five species from indoor (Human and Cattle shed) and only four species from outdoor collection. The numbers of species were greater in Cattle shed; this may due to open type of Cattle sheds.

The Correlation Coefficient between number of mosquitoes (X) and Temperature (Y) is 0.0685 (See Appendix 1). The number of mosquitoes depends on temperature. The calculated value of χ^2 (40.781) is greater than tabulated value (19.68) at 5% level of significance and 11 degree of freedom.

So, null hypothesis is rejected. Hence, the number of mosquitoes collected in Post Monsoon Survey and Pre Monsoon Survey is dependent.

In the present study *Culex quinquefasciatus* is most abundant species followed by *Cx. fuscocephala*, *Cx. tritaeniorhynchus*, *Cx. gelidus*, *Cx. bitaeniorhynchus*, *Cx. whitmorei*, *Cx. pseudovishnui*, *Cx. whitei*, *Cx. theileri*, *Cx. barraudi*, *Cx. sinensis* and *Cx. mimulus*. *Cx. tritaeniorhynchus* was the third most abundant species in indoor collection. The numbers of *Cx. tritaeniorhynchus* in indoor and outdoor collection were 42 and five in post-monsoon season and 21 and zero in pre-monsoon season respectively.

Kanojia (2006) reported *Culex quinquefasciatus* is the fourth most abundant species, one isolate of JE virus has been obtained from this species. *Cx. tritaeniorhynchus*, *Cx. quinquefasciatus*, *Anopheles subpictus* and *An. peditaeniatus* were collected throughout the year. *Cx. tritaeniorhynchus* was the most abundant species showed two density peaks, first in February and second in October. Its population dynamics is closely associated with paddy cultivation in Bellary district, Karnataka.

Das et al., (2004) reported *Culex tritaeniorhynchus* is pre dominant in outdoors and playing a main role in JE transmission in Warangal and Karim Nagar district of Andhra Pradesh.

Arunachalam et al., (2004) reported *Culex tritaeniorhynchus* (66.7%) was most abundant species in Kerala, south India with increase in number associated with rice cultivation.

Kumar et al; (2004) recorded a total of 60 species belonging to 10 genera. Genus *Culex* was predominant in Rajiv Gandhi National Park (Nagarahole), Karnataka state, India.

Minakawa et al., (2002) reported *Anopheles gambiae* was the pre dominant species in both larval and adult samples. Livestock and human host availability affect the relative abundance of *An.gambiae* larvae in aquatic habitats, but the distribution of Anopheline adults in houses is determined by the distance from houses to larval habitats.

In the present study, *Culex quinquefasciatus* and *Cx. fuscocephala* were reported in 4 months (August, September, March and April). A total of 12 species were reported in September from Bramhapuri village. *Culex whitmorei*, *Cx. pseudovishnui*, *Cx. whitei*, *Cx. theileri*, *Cx. barraudi*, *Cx.*

sinensis and *Cx. mimulus* were reported in August, March and April. Mosquito's population was peak in August-September (post-monsoon).

Geevaryhes et al., (1994) reported that mosquito's population was peak during March-April and September. Very high incidence of JE cases were in extensively irrigated areas and a low incidence in some of taluks with less or no irrigation system.

Muturi et al., (2006) reported *Anopheles arabiensis*, *Culex quinquefasciatus*, and *Anopheles pharoensis* were more abundant in rice agro ecosystem than in the non-irrigated agro ecosystem and in planned than in the unplanned rice agro ecosystems. But *An. funestus* was more abundant in the non-irrigated agro ecosystem.

In the present study, among 1197 *Culex* sample, 12 species were recorded. *Culex quinquefasciatus* was the most abundant species in both post-monsoon and pre-monsoon season. *Culex mimulus* was the least abundant species. Vector abundance of *Cx. quinquefasciatus* was 26.90% in August-September (post-monsoon) and 18.71% in March-April (pre-monsoon). The vector abundance of *Cx. tritaeniorhynchus* was 3.92% in August-September (post-monsoon) paddy cultivation period and 1.75% in March-April (pre-monsoon) which is the third most abundant species. The percentage abundance of *Culex whitmorei*, *Cx. pseudovishnui*, *Cx. whitei*, *Cx. barraudi*, *Cx. theileri*, *Cx. sinensis*, and *Cx. mimulus* was least in post monsoon and zero in pre-monsoon. All 12 species were more abundant in August-September (post-monsoon).

The vector abundance was least in pre-monsoon season i.e. hot and dry season (March-April) and highest in post-monsoon season i.e. wet or rainy season (August-September), paddy cultivation period.

Gingrich et al; (1992) state that, vector abundance was high in monsoon (May-October), moderate in transition (March-April and November-December) and low in dry (January-February) seasons in Bangkok.

Same to present study, Gajanana et al; (1997) reported vector abundance was lowest in the hot and dry season (April-June) and highest in the cool and wet season (October-December) in the south Arwt district in Tamil Nadu.

Pramanik and Raut (2002) reported *Culex quinquefasciatus* was dominant (62.96%) on average of 492 mosquitoes in Kolkata.

Barboso et al., (2003) reported 15 species among the collected 312 Culicine specimens in a restrict forest insi Curitiba urban area (Parana, Brazil).

Rajendran et al., (2003) reported average vector abundance per man hour for *Culex tritaeniorhynchus* was 324.5 per month for the period June 1998-May 2000 in south Indian villages.

Kanojia et al., (2003) states that *Culex tritaeniorhynchus* the most likely vector of JE together with other known vector species remains more active during the period of paddy cultivation in Gorakhpur district.

Anopheles sinensis was most abundant (53.4%) in species ratio, followed by *Cx. tritaeniorhynchus* (43.0%), *Cx. inatomii* (1.6%), *Ochleratatus dorsalis* (1.3%) and *Cx. pipiens pallens* (0.5%) in a study carried out by Jeong et al., (2003).

Kanojia et al., (2004) reported, *Culex tritaeniorhynchus*, the primary vector of JE in Bellary district, India. More specimens (n=20,966) were found resting in indoor habitats than in outdoor vegetation (n=383), despite the availability of outdoor resting sites. Indoor residual insecticide may provide an effective control method in this area.

Mwangangi et al., (2006) reported that *Culex quinquefasciatus* (65.7%) was the non- anopheline species. Culicine and Aedine species densities were significantly higher during the Post-harvesting period. Transplanting stage is favourable for the growth of immature stages of *Anopheles arabiensis* and provides a narrow window for targeted larval intervention in rice.

Yaw et al., (2006) state that Mosquito survivorship and fecundity may be affected by environmental factors such as temperature and humidity. A Chigher smbient temperature may be facilitate blood meal digestion, reduce the length of the gonotropic cycle, and change the lifetime fecundity of a mosquito.

In present study In Suryapur village rooms of only one house was spread by GHAMBHIB Chloropyriphos 50%+ Cypermethrin 5% EC and in the room of first house of Kesharbag, DHOOM All Night (Alethrin Mat) was used in post monsoon. More specimens were found resting in indoor habitats

(n=242). Indoor resting specimens in Post monsoon (n=543) and in pre monsoon (n=412). The specimens in outdoor vegetation in Post monsoon (n=175) and in pre monsoon (n=67).

Odiere et al; (2007) recorded 63% *Anopheles gambiae* S.S. (46% female) and 37% were *An. arabiensis* (66% female) out of total 10,517 mosquitoes samples in outdoor resting. Additionally 617 *An. funestus* (58%female) and 5,232 *Culex spp.* (males and females together) were collected in Kenya.

6.2 Comments

The major Comments of the study are given below:

1. A total of 1197 Culex Mosquitoes (718 in post-monsoon survey and 479 in pre-monsoon survey) were recorded.
2. About 12 species were recorded (12 species in cattle shed, 11 in human dwelling and outdoor collection) in post-monsoon survey and 5 species (5 in indoor-cattle shed and human dwelling, 4 in outdoor collection) in pre-monsoon survey.
3. Highest number of Mosquitoes sample (n=403) was collected in September.
4. *Culex quinquefasciatus* is the most abundant species in both season (26.90 percent in post monsoon survey and 18.71percent in pre-monsoon survey).
5. Indoor resting density (human) 9.33 was highest in post-monsoon survey at 25.75°C in Champanagar village.
6. Indoor resting density (cattle) 5.55 was highest in pre-monsoon survey at 22°C in Suryapur village.
7. Outdoor density, 0.22 was highest in pre-monsoon survey at 22.5°C in Kholesimal village.
8. The resting habit of *Culex mimulus* and *Cx. tritaeniorhynchus* was infinitive in post-monsoon and pre-monsoon survey respectively.
9. The room density of *Culex quinquefasciatus* (6.5) was highest in Champanagar village.
10. About 12 species of Culex Mosquitoes were found at 25.26°C in September in Bramhapuri village.
11. The number of mosquitoes is positively correlated with temperature. The number of mosquitoes is dependent on temperature.
12. The number of mosquitoes collected in post-monsoon survey and pre-monsoon survey is dependent.

CHAPTER VII

RECOMMENDATION

1. To reduce the vector contact, use of insecticide impregnated mosquito (bed) nets is strongly suggested. Moreover reduction of outdoor activities, discouraging to sleep outside the house during summer and rainy season, and un-exposing body parts by wearing long sleeves clothes should be highlighted to protect the people from mosquito bites.
2. Intermittent irrigation and periodic flushing of rice fields during rice cultivation should be followed.
3. Use of biological control methods viz., microbial agents like *Bacillus thuringiensis* and *B. sphaericus*, and introduction of carnivorous fishes like *Gambusia affinalis* etc. are recommended to control JE & Filaria vectors rather than chemical ones.
4. There is lack of general knowledge about Mosquitoes disease (JE & Filaria) in the district. So considerable mass awareness and public health education campaigns are thus conducted to change public perception of communities and VDCs level about the disease problem, its emergence and for its prevention and control.
5. Environmental and entomological studies in relation to the endemicity of JE particularly with respect to vector, larval habitat, temperature, humidity etc. as well as amplifying hosts should be carried out.
6. Public Awareness Program should be applied from the government level (Health Ministry) and related other government and private sector to aware the villagers about the *Culex* mosquitoes (Character, breeding and biting habit) their diseases and prevention from the bite and control these mosquitoes.

ANNEXES

A. Hypothesis Testing

The aim of the present research study is also testing the hypotheses which are mentioned in **CHAPTER I**. The methodology has been also mentioned in **CHAPTER III**. The following principles have been adopted for testing the hypothesis.

Annex - 1

First Hypothesis Testing

The First Hypothesis is “The number of mosquitoes depends on temperature i.e. there is positive relationship between the number of mosquitoes and temperature.”

Procedures

At First, Simple correlation coefficient of number of mosquitoes on temperature has to be found out.

Correlation Analysis

For Calculating the correlation coefficient of number of mosquitoes and temperature, let us denote the variables symbolically, i.e.

Number of Mosquitoes =X

Temperature (°c) =Y

Number of observation = n

The variable X denotes the number of mosquitoes of different villages. The variable Y represents the field temperature.

Table 1
Computation of Correlation Coefficient

| Serial Number | X | Y | X ² | Y ² | XY |
|---------------|---------------|-----------------|----------------------------|-------------------------------|--------------------|
| 1 | 116 | 25.74 | 13456 | 662.547 | 2985.84 |
| 2 | 116 | 25.42 | 13456 | 646.176 | 2948.72 |
| 3 | 81 | 26.83 | 6561 | 719.848 | 2173.23 |
| 4 | 78 | 22.33 | 6084 | 498.628 | 1741.74 |
| 5 | 73 | 28.33 | 5329 | 802.588 | 2068.09 |
| 6 | 72 | 28.45 | 5184 | 809.402 | 2048.4 |
| 7 | 64 | 25.24 | 4096 | 637.057 | 1615.36 |
| 8 | 61 | 19.91 | 3721 | 396.408 | 1214.51 |
| 9 | 58 | 25.74 | 3364 | 662.547 | 1492.92 |
| 10 | 56 | 22 | 3136 | 484 | 1232 |
| 11 | 56 | 21.58 | 3136 | 465.696 | 1208.48 |
| 12 | 52 | 27.54 | 2704 | 758.451 | 1432.08 |
| 13 | 49 | 27.08 | 2401 | 733.326 | 1326.92 |
| 14 | 48 | 22.33 | 2304 | 498.628 | 1071.84 |
| 15 | 44 | 21.75 | 1936 | 473.062 | 957 |
| 16 | 42 | 21.75 | 1764 | 473.062 | 913.5 |
| 17 | 42 | 21.41 | 1764 | 458.388 | 899.22 |
| 18 | 37 | 26.33 | 1369 | 693.268 | 974.21 |
| 19 | 26 | 21.33 | 676 | 454.968 | 554.58 |
| 20 | 26 | 20 | 676 | 400 | 26.2 |
| Total | X=1197 | Y=481.09 | X²=83117 | Y²=11728.05 | XY=28884.84 |

$$\text{Correlation Coefficient}(r) = \frac{n \sum XY - \sum X \sum Y}{\sqrt{[n \sum X^2 - (\sum X)^2][n \sum Y^2 - (\sum Y)^2]}}$$

$$r = \frac{20 \times 28884.84 - 1197 \times 481.09}{\sqrt{[20 \times 83117 - 1197^2][20 \times 11728.05 - 481.09^2]}}$$

$$= \frac{577696.8 - 575864.43}{\sqrt{[1662340 - 1432809][234561 - 231447.588]}}$$

$$\begin{aligned}
&= \frac{1832.07}{\sqrt{229531 - 3113.412}} \\
&= \frac{1832.07}{26732.462} \\
&= 0.0685
\end{aligned}$$

Hence, there is positive relationship between number of mosquitoes (X) and Temperature (Y). In other words, they are dependent on each other.

Annex - 2

Second hypothesis Testing:

The second hypothesis is “The number of mosquitoes collected in post monsoon survey and pre monsoon survey is independent.”

For the testing of this hypothesis, Chi –Square (t^2) test has been used.

Procedures

1. Setting of Hypothesis:

- i. Null Hypothesis (H_0): The number of Mosquitoes collected in post monsoon survey and pre monsoon survey is independent.
- ii. Alternative Hypothesis (H_1): The number of Mosquitoes collected in post monsoon survey and pre monsoon survey is dependent.

2. Level of significance: 5 percent

3. Test Statistics: t^2 - test

4. Computation: $t^2 = \sum \frac{(O - E)^2}{E}$

Table 2
Observed Frequency Table

| Name of Species | Post-Monsoon | Pre-Monsoon | Total |
|------------------------------|---------------------|--------------------|--------------|
| <i>Cx. quinquefasciatus</i> | 322 | 224 | 546 |
| <i>Cx. fuscocephala</i> | 250 | 198 | 448 |
| <i>Cx. tritaeniorhynchus</i> | 47 | 21 | 68 |
| <i>Cx. gelidus</i> | 28 | 18 | 46 |
| <i>Cx. bitaeniorhynchus</i> | 20 | 18 | 38 |
| <i>Cx. whitmorei</i> | 12 | 0 | 12 |
| <i>Cx. pseudovishnui</i> | 11 | 0 | 11 |
| <i>Cx. whitei</i> | 9 | 0 | 9 |
| <i>Cx. barraudi</i> | 7 | 0 | 7 |
| <i>Cx. theileri</i> | 7 | 0 | 7 |
| <i>Cx. sinensis</i> | 4 | 0 | 4 |
| <i>Cx. mimulus</i> | 1 | 0 | 1 |
| Total | 718 | 479 | 1197 |

Table 3
Calculation of Estimated Frequency Table

| Name of Species | Post- Monsoon | Pre- Monsoon | Grand Total |
|------------------------------|---|---|--------------------|
| <i>Cx. quinquefasciatus</i> | $\frac{546 \times 718}{1197} = 327.50$ | $\frac{546 \times 479}{1197} = 218.491$ | 546 |
| <i>Cx. fuscocephala</i> | $\frac{448 \times 718}{1197} = 268.725$ | $\frac{448 \times 479}{1197} = 179.27$ | 448 |
| <i>Cx. tritaeniorhynchus</i> | $\frac{68 \times 718}{1197} = 40.788$ | $\frac{68 \times 479}{1197} = 27.211$ | 68 |
| <i>Cx. gelidus</i> | $\frac{46 \times 718}{1197} = 27.594$ | $\frac{46 \times 479}{1197} = 18.407$ | 46 |
| <i>Cx. bitaeniorhynchus</i> | $\frac{38 \times 718}{1197} = 22.793$ | $\frac{38 \times 479}{1197} = 15.206$ | 38 |
| <i>Cx. whitimorei</i> | $\frac{12 \times 718}{1197} = 7.197$ | $\frac{12 \times 479}{1197} = 4.802$ | 12 |
| <i>Cx. pseudovishnui</i> | $\frac{11 \times 718}{1197} = 6.598$ | $\frac{12 \times 479}{1197} = 4.401$ | 11 |
| <i>Cx. whitei</i> | $\frac{9 \times 718}{1197} = 5.398$ | $\frac{9 \times 479}{1197} = 3.601$ | 9 |
| <i>Cx. baraudi</i> | $\frac{7 \times 718}{1197} = 4.198$ | $\frac{7 \times 479}{1197} = 2.801$ | 7 |
| <i>Cx. theileric</i> | $\frac{7 \times 718}{1197} = 4.198$ | $\frac{7 \times 479}{1197} = 2.801$ | 7 |
| <i>Cx. sinensis</i> | $\frac{4 \times 718}{1197} = 2.399$ | $\frac{4 \times 479}{1197} = 1.600$ | 4 |
| <i>Cx. mimulus</i> | $\frac{1 \times 718}{1197} = 0.599$ | $\frac{1 \times 479}{1197} = 0.4$ | 1 |
| Total | 718 | 479 | 1197 |

Table 4
Calculation of t^2

| Observed Frequency (O) | Expected Frequency (E) | $(O - E)$ | $(O - E)^2$ | $\frac{(O - E)^2}{E}$ |
|------------------------|------------------------|-----------|-------------|-------------------------------------|
| 322 | 327.50 | -5.5 | 30.25 | 0.092 |
| 250 | 268.725 | -18.725 | 350.62 | 1.304 |
| 47 | 40.788 | 6.212 | 38.588 | 0.946 |
| 28 | 27.59 | 0.41 | 0.168 | 0.006 |
| 20 | 22.793 | -2.793 | 7.8 | 0.375 |
| 12 | 7.197 | 4.803 | 23.068 | 3.205 |
| 11 | 6.598 | 4.402 | 19.377 | 2.936 |
| 9 | 5.398 | 3.602 | 12.974 | 2.403 |
| 7 | 4.198 | 2.802 | 7.851 | 1.87 |
| 7 | 4.198 | 2.802 | 7.851 | 1.87 |
| 4 | 2.399 | 1.601 | 2.563 | 1.068 |
| 1 | 0.599 | 0.401 | 0.160 | 0.268 |
| 224 | 218.491 | 5.509 | 30.349 | 0.138 |
| 198 | 179.27 | 18.73 | 350.81 | 1.956 |
| 21 | 27.211 | -6.211 | 38.576 | 1.417 |
| 18 | 18.407 | -0.407 | 0.165 | 0.008 |
| 18 | 15.206 | 2.794 | 7.806 | 0.513 |
| 0 | 4.802 | -4.802 | 23.059 | 4.802 |
| 0 | 4.401 | -4.401 | 19.368 | 4.401 |
| 0 | 3.601 | -3.601 | 12.967 | 3.601 |
| 0 | 2.801 | -2.801 | 7.845 | 2.801 |
| 0 | 2.801 | -2.801 | 7.845 | 2.801 |
| 0 | 1.600 | -1.600 | 2.56 | 1.600 |
| 0 | 0.4 | -0.4 | 0.16 | 0.4 |
| | | | | $\sum \frac{(O - E)^2}{E} = 40.781$ |

5. Degree of freedom (d. f.) = (r-1) (c-1)
= (2-1) (12-1)
=11

6. Tabulated Value of t^2 at 5 percent level of significance of 11 degree of freedom is 19.68.

7. **Decision:** Since the calculated value of t^2 is greater than tabulated value, null hypothesis is rejected. Hence the number of mosquitoes collected in post-monsoon survey and pre-monsoon survey is dependent.

B. Recorded Temperature in Post Monsoon and Pre Monsoon

Table 5
Temperature in pre and post monsoon

| Village | Number | Temperature in °c in Post monsoon | | | Temperature in °c in Pre monsoon | | |
|------------|--------|-----------------------------------|--------|---------|----------------------------------|--------|---------|
| | | Indoor | | Outdoor | Indoor | | Outdoor |
| | | Human | Cattle | | Human | Cattle | |
| Gitanagar | 1 | 27 | 27 | 28 | 18 | 17 | 14 |
| | 2 | 30 | 28 | 29 | 21 | 19 | 18 |
| | 3 | 27 | 28 | 30 | 23 | 24 | 28 |
| | 4 | 29 | 29 | 29 | 27 | 26 | 29 |
| Kesherbag | 1 | 27 | 24 | 25 | 17 | 17 | 16 |
| | 2 | 26 | 26 | 27 | 20 | 19 | 19 |
| | 3 | 28 | 27 | 27 | 22 | 24 | 25 |
| | 4 | 28 | 27 | 28 | 24 | 26 | 28 |
| Parasnagar | 1 | 27 | 27 | 26 | 17 | 17 | 16 |
| | 2 | 29 | 26 | 26 | 21 | 19 | 18 |
| | 3 | 28 | 29 | 28 | 22 | 20 | 21 |
| | 4 | 28 | 30 | 31 | 23 | 22 | 24 |
| Suryapur | 1 | 24 | 25.5 | 26 | 19 | 17 | 16 |
| | 2 | 27 | 26 | 28 | 23 | 21 | 20 |
| | 3 | 28 | 27 | 30 | 24 | 24 | 25 |
| | 4 | 29 | 30 | 30 | 25 | 26 | 28 |
| Kholesimal | 1 | 26 | 25 | 25.5 | 20 | 18 | 17 |
| | 2 | 28 | 25 | 26 | 22 | 21 | 19 |
| | 3 | 28 | 25.5 | 26 | 23 | 23 | 25 |
| | 4 | 28 | 26 | 27 | 25 | 26 | 29 |
| Dobato | 1 | 27 | 26 | 25 | 16 | 14 | 14 |
| | 2 | 27 | 26 | 26 | 18 | 16 | 16 |
| | 3 | 28 | 28 | 27 | 21 | 23 | 24 |

| | | | | | | | |
|-------------|---|------|------|------|----|----|----|
| | 4 | 28 | 28 | 29 | 25 | 25 | 27 |
| Champanagar | 1 | 25 | 25 | 24 | 19 | 18 | 17 |
| | 2 | 25 | 25 | 25.5 | 21 | 19 | 19 |
| | 3 | 25 | 25 | 26 | 22 | 22 | 23 |
| | 4 | 26 | 25.5 | 27 | 25 | 24 | 27 |
| Vijayanagar | 1 | 25 | 24 | 24 | 18 | 17 | 15 |
| | 2 | 26 | 25 | 25.5 | 19 | 19 | 18 |
| | 3 | 26 | 26 | 27 | 22 | 23 | 27 |
| | 4 | 26 | 26.5 | 28 | 26 | 27 | 19 |
| Rampur | 1 | 25 | 24 | 24 | 19 | 16 | 18 |
| | 2 | 25 | 24.5 | 24.5 | 20 | 20 | 19 |
| | 3 | 26 | 25 | 26 | 23 | 24 | 25 |
| | 4 | 26 | 26 | 27 | 24 | 26 | 28 |
| Bramhapuri | 1 | 25 | 24 | 24 | 18 | 16 | 16 |
| | 2 | 25 | 25 | 25 | 22 | 19 | 18 |
| | 3 | 25.5 | 25 | 26 | 24 | 23 | 26 |
| | 4 | 26 | 26 | 27 | 26 | 25 | 28 |

Identification key to the genera *Culex*

- Proboscis not only slightly curved, if at all; posterior border of wing evenly rounded or only slightly emarginated
- Scutellum trilobed, with setae in three distinct groups; maxillary palpi shorter than proboscis
- Proboscis only slightly swollen near tip, if at all; with neither flexible long setae apically
- Pre-spiracular area bare
- Mesopostnotum without setae; scutum with another type of scale
- Fore and mid tarsomeres one shorter than other four tarsomeres combined, tarsomeres 4 of fore and mid legs much longer than wide
- Post spiracular setae absent
- Alula fringed with narrow scales
- Pulvilli present; tarsal claws unusually small

Identification key to the *Culex fuscocephala*

- One or two mesepimeral setae present; proboscis without distinct pale-scaled band; tarsomeres without pale bands at joints.
- Anterior surface of midfemur without median longitudinal pale-scaled stripe
- Abdominal terga without basal transverse, pale-scaled band; pleuron with striking pattern of dark and pale integumental stripes

Identification key to the *Culex quinquefasciatus*

- One or two mesepimeral setae present; proboscis without distinct pale-scaled band; tarsomeres without pale bands at joints.
- Anterior surface of midfemur without median longitudinal pale-scaled stripe
- Abdominal terga without basal transverse, pale-scaled band; pleuron with striking pattern of dark and pale integumental stripes
- Integument of thoracic pleuron without dark stripe; scutal integument yellowish or pale brown

Identification key to the *Culex theileri*

- One or two lower mesepimeral setae present; proboscis without distinct pale-scaled band; tarsomeres without pale bands at joints
- Anterior surface of midfemur without median longitudinal pale-scaled stripe
- Post spiracular area and base of prealar knob with distinct pale-scaled patches

Identification key to the *Culex whitei*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands
- Wing without pattern of pale-scaled or streaks
- Abdominal terga dark-scaled, with or without pale-scaled bands
- Abdominal terga 2-4 entirely dark –scaled, without pale bands or apicolateral pale patches

Identification key to the *Culex sinensis*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands
- Wing without pattern of pale-scaled or streaks
- Abdominal terga 2-4 with bands or patches or pale scales
- Wing with dark scales on all veins; hind tarsomeres with pale bands at bases longer than those at apices

Identification key to the *Culex barraudi*

- Wing without pattern of pale-scaled or streaks

- Abdominal terga dark-scaled, with or without pale-scaled bands
- Abdominal terga 2-4 with bands or patches or pale scales
- Abdominal terga 2-4 with basal pale-scaled bands only
- Erect scales in center of vertex pale yellow, dingy white or all dark; anterior 0.7 of scutum covered with being yellow, golden or dark scales
- Midfemur with longitudinal stripe of pale scales on anterior surface; post spiracular area with small patch of semi-erect scales on lower anterior aspect
- Longitudinal pale-scaled stripe on anterior surface of midfemur broken into small spots at middle ; costal vein entirely dark –scales

Identification key to the *Culex mimulus*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands
- Wing without pattern of pale-scaled or streaks on at least 2 areas of costa and 1 area on other veins
- Basalmost pale scaled costal spot involves C, Sc, R, and sometimes R₅
- And Cu; basal pale bands of abdominal terga marrow, usually less than 0.25 length of segment

Identification key to the *Culex gelidus*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands
- Wing without pattern of pale-scaled spots or streaks

- Abdominal terga dark-scaled, with or without pale-scaled bands
- Abdominal terga 2-4 with bands or patches or pale scales
- Abdominal terga 2-4 with basal pale-scaled bands only
- Erect scales in center of vertex pale yellow, dingy whitish; anterior 0.7 of scutum covered with white scales
- Anterior surface of fore and midfemora without speckling of pale scales; white-scaled patch on scutum dense, extending to wing root, dark –scaled posterior to that; wing veins R₁, R₄₊₅ and Cu with narrow scales

Identification key to the *Culex whitmorei*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands
- Wing without pattern of pale-scaled spots or streaks
- Abdominal terga dark-scaled, with or without pale-scaled bands
- Abdominal terga 2-4 with bands or patches or pale scales
- Abdominal terga 2-4 with basal pale-scaled bands only
- Erect scales in center of vertex pale yellow, dingy whitish; anterior 0.7 of scutum covered with white scales
- Anterior surface of fore and midfemora extensively speckled with pale scales; pale-scaled patch on scutum thinner, extending posterior to wing root in 4 lines, wing veins R₁, R₄₊₅ and Cu with broad scales

Identification key to the *Culex bitaeniorhynchus*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands

- Wing without pattern of pale-scaled spots or streaks
- Abdominal terga dark-scaled, with or without pale-scaled bands
- Abdominal terga 2-4 with bands or patches or pale scales
- Abdominal terga 2-4 with apical or basal pale-scaled bands only
- Wing with mixed pale and dark scales; hind tarsomeres with apical and basal pale bands about same length
- Abdominal terga 2-4 with broad apical bands of pale scales; abdominal terga heavily speckled with pale scales

Identification key to the *Culex tritaeniorhynchus*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands
- Wing without pattern of pale-scaled spots or streaks
- Abdominal terga dark-scaled, with or without pale-scaled bands
- Abdominal terga 2-4 with bands or patches or pale scales
- Abdominal terga 2-4 with apical or basal pale-scaled bands only
- Erect scales in center of vertex pale yellow, dingy whitish; anterior 0.7 of scutum covered with beige, yellow, golden or dark scales
- Midfemur entirely dark-scaled or specking of pale scales not forming definite stripe; postspiracular area without scales on lower anterior aspect
- Anterior surface of fore and midfemora entirely dark scaled
- Erect scales on vertex mostly dark anterior surface of hindfemur pale-scaled with narrow black-scaled ring apically; scutum covered with dark coppery gold scales

Identification key to the *Culex pseudovishnui*

- Lower mesepimeral setae absent; proboscis with distinct pale-scaled band; tarsomeres with basal and apical pale bands
- Wing without pattern of pale-scaled spots or streaks
- Abdominal terga dark-scaled, with or without pale-scaled bands
- Abdominal terga 2-4 with bands or patches or pale scales
- Abdominal terga 2-4 with apical or basal pale-scaled bands only
- Erect scales in center of vertex pale yellow, dingy whitish; anterior 0.7 of scutum covered with beige, yellow, golden or dark scales
- Midfemur entirely dark-scaled or specking of pale scales not forming definite stripe; postspiracular area without scales on lower anterior aspect
- Erect scales on vertex pale yellow in center, dark-scaled posterolaterally; hind femur marked otherwise; scutum with scales paler
- Femora and proboscis never speckled with pale scales; scutum with yellow to silvery scales; hind femur with dark band apically contrasting with pale-scaled areas.

REFERENCES

- Afrane A Yaw, Goufa Zhou, Bernard W. Lawson, Andrewk Githeko, and Guiyun Yan (2006). Effect of Microclimatic Changes caused by Deforestation on the Survivorship And Reproductive Fitness of *Anopheles gambiae* in Western Kenya High lands.
- Akiba, T.K., Osaka, S.Tang, M.Nakayama, A. Yamamoto, I.Kurane et al; (2001). Analysis of Japanese encephalitis epidemic in Western Nepal in 1997. *Epidemiol. and infectmi.* **126** (1) 81-88.
- Annual Report (2001). Epidemiology and Disease Control Division (EDCD), Department of Health Services (DHS), Ministry of Health (MOH), HMG, Nepal 42-55.
- Apperson, Charles S. Bruce A. Harrison, Thomas R. Unnasch Hassan K. Hassan, William S. Irby, Harry M. Savage, Stephen E. Aspen, D wesley Wastson, Leopoldo M. Rueda, Barry R. Engber and Roger, S. Nasci (2002). Host feeding habits of *Culex* and other mosquitoes (Diptera: Culicidae) in the Brough of Queens in New York City, with characters and techniques for identification of *Culex* mosquitoes. *Journal of Medical Entomology.* **39** (5): 777-785
- Arunachalam N. P. Philip Samuel, J. Hiriyani, R.Rajendran and A.P. Dash (2005). Observation on the Multiple feeding Behavior of *Culex tritaeniorhynchus* (Diptera; Culicidae), The vector of Japanese Encephalitis in Eerala in Southern India. *Am. J. Trop. Med. Hyg.* **72** (2): 198-200.
- Arunachalam, Philip Samuel N, P., Hiriyani N, P., Thenmozhi V. and Gajanana A. (2004). Japanese encephalitis in Kerala, South India: Can *Mansonia* (Diptera: culicidae) play a supplemental role in transmission. *Journal of Medical Entomology.* **41** (3): 456-461.
- Banerjee, K., and Deshmukh, P.K. (1984) Transmission of JEV to chicks individual *Culx tritaeniorhynchus* mosquitoes. *Indian J.Med.Res.*, **86**. 726-27(Eng.).

- Barbosa, Andreia Aparecida*, Mario. Antonia Navarrosilva and Daniela Calado* (2003). Culicidae activity in a restrict forest insi Curitiba urban area (Parana, Brazil). *Revista Brasileira de Zoologia*. **20** (1): 59-63.
- Bhusal, K.P., Joshi, A.B.; Mishra, P.N. and Bhusal, K. (2002). Prevalence of wuchereria bancrofti infection in Tokha, Chandeswori VDC of Kathmandu. *Journal of the Institute of Medicine, Kathmandu*. **22**: 204-211.
- Brydon, H.W., Joshi G. and Pradhan S. (1961). *Anopheles* distribution and its relation to the Malaria eradication Program in Central Nepal. *Mosq. News* **21**:102-105.
- Buescher, E.L., Schere, W. R. (1959). Ecological studies of Japanese encephalitis virus in Japan. IX. Epidemiologic correlations and conclusions. *Am. J. Trop. Med. Hyg.* **8**: 719- 122.
- Buescher, E.L., Scherer W.F., Rosenberg M.Z., Greesser L. Hardy J.L. and Bullock H.R.. (1959). ecological studies of Japanese Encephalitis Virus in Japan II. Mosquito infections. *Am. J. Trop. Med. Hyg.* **8**: 651-677.
- Burgess, M. (1990). *Mosquitoes of Nepal*. *Mosq. Sys.* **21** (3): 55-79.
- Carey, D.E., Reuban, R. and Myers, R.M. (1968). Japanese encephalitis studies in Vallore, South India. Part IV. Search for virological and serological evidence of infection in animals other than man. *Ind. J. Med. Res.* **56**: 1340.
- Darsie, R.F. (1994). Zoogeography of the mosquitoes of Nepal. *J. Am. mosquito control Assoc.* **7**:28-33.
- Darsie, R.F. and Pradhan S.P. (1990). The mosquitoes of Nepal: Their identification, distribution and biology. *Mosquito syst.* **22** (2): 69-130.
- S.P. (Darsie, R.F. and Pradhan 1994). Notes of Anopheles mosquito of Nepal. *J. Nepal. Med. Assoc.* **8**: 89-97.
- Darsie, R.F. and Pradhan (1994). Keys to the Mosquitoes of Nepal.
- Das BP, Lal S, Saxena VK, (2004). Outdoor resting Preference of *Culex tritaeniorynchus*, the vector of Japanese encephalitis in Warangal and Karim Nagar districts, Andhra Pradesh. *Journal of vector Borne Disease.* **41** (1-2): 32-36.
- Das, Bina Pani*, Sharma S.K. and Datta* K.K. (2000). Prevalence of *Aedes aegypti* at the international port and airport, Kolkata (West Bengal). India, *Dengue Bulletin.* **24**:124-126.

- Detels, R., Cates, M.D., Cross, G.S., Warren R.H. (1970). Ecology of Japanese encephalitis virus in Taiwan in 1968. *Am. J. Trop. Med. Hyg.* **19**(4): 716 -723
- DoHS, Ministry of Health and Population, (2005/06). Lymphatic Filariasis. An Annual Report, Nepal, 139-141.
- Dos Santos, Roseli La Corte*, Oswaldo Paulo Forattini and Marcelo Nascimento Burattini (2004). *Anopheles albitarsis* 5% (Diptera: Culicidae) survivorship and density in a rice irrigation area of the state of Sao Paulo, Brazil. *Journal of Medical Entomology.* **41** (5): 997-1000.
- Dutta, P., Khan S. A, Sharma C. K., Hazarika* N.C. and Mahanta J. (2003). Survey of medically important mosquito fauna in Mizoram. *Entomon* **28** (3): 237-240.
- Elizondo-Quiroga, Armando, Adriana Flores-Suarez, Darwin Elizondo Quiroga, Gustavo Ponce-Garcia, Bradley J. Blitvich, Juvan Francisco Contreras-Cordero, Jose Ignacio Gonzales-Rojas, Roberto Mercado Hernandez Berry J. Beaty and Ildefonso Fernandez-Salas(2006). Host feeding preference of *Culex quinquefasciatus* in Monterrey, Northeastern Mexico. *Journal of the American Mosquito Control Association.* **22** (4): 654-661.
- Fakoorziba, M.R., and Vijayan V.A. (2003). Variation in morphological characters of *Culex tritaeniorhynchus* (Diptera: Culicidae), a Japanese encephalitis vector at Mysore India. *Journal of communicable Diseases.* **35** (3): 206-213.
- Feng Guang Zhu, Rong Xu (2004). Annual dynamic change of Mosquito density during 1999-2002 in Ningbo city of Zhejiang Province. *Chinese Journal of Vector Biology and Control.* **15** (3): 182-183.
- Gautam, Dhimal M. D, Shrestha S.R. and Tamrakar A.S (2009). First Record of *Aedes aegypti* (L.) Vectors of Dengue virus from Kathmandu, Nepal. *J. Nat. Hist. Mus.* **24**
- Geevarghese G, Mishra AC, Jacob PG, Bhat HR (1994). Studies on the mosquito vectors of Japanese encephalitis virus in Mandya District, Karnataka, India. *Southeast Asian J. Trop Med Public Health.* **25** (2): 378-382.
- Gingrich JB, Nisalak A, Latendresse JR, Sattabong Kot J, Hoke J, Hoke CH, Pomsdhit J, Chantalakana C, Satayaphanta Uechiewcharnkit K, Innis BL.

- (1992). Japanese Encephalitis virus in Bangkok: factors influencing vector infections in three suburban communities. *Journal of Medical Entomology*. **29** (3): 436-444.
- Gomes, A.C., Silva N.N., Marques G.R.A.M. and Brito M.(2003). Host-feeding patterns of Potential human disease vectors in the Paraiba Valley Region, state of Sao Paulo Brazil. *Journal of vector ecology*. **28** (1):74-78.
- Gould, D.J., Barnett H.C. and Suyemoto. W. (1962). Transmission of Japanese Encephalitis virus by *Culex gelidus* Theobald. *Trans. R. Soc. Trop. Med. Hyg.* **56**: 429-135.
- Gould, D.J., Edelman R., Grossman R.A., Nisalik A. and Sullivan M.F. (1974). Study of Japanese Encephalitis virus in Chiangmai Valley, Thailand. Iv. Vector studies. *Am. J. Epidemiol.* **100**: 49-56.
- Gould, D.J., Yuill T.M., Moussa M.A., Simasathien P. and Rutledge L.C 1968). An insular outbreak of dengue hemorrhagic fever III. Identification of vectors and observations on vector ecology. *Am. J. Trop. Med. Hyg.* **17**:609-618.
- Guimaraes, Erico Anthony, Gentile Carla, Lopes Macedo and Hello Rubens Pinto (2000).Ecology of mosquitoes (Diptera:Culicidae) in areas of Serra do Mar State Park State of Sao Paulo, Brazil.II:Habitat distribution, *Memorias do Instituto Oswaldo Cruz* **95**(1):17-28.
- Gurung, C.K. and. Singh R.B (2003). Factors associated with Japanese encephalitis in Nepal. *J. Assoc. Med. Lab. Sci* **5** (5): 41-45.
- Hale, J.H., Colless D.H. and Kim. K.A. (1957). Investigation of the Malaysian form of *Culex tritaeniorynchus* as a potential vector of Japanese B Encephalitis virus on Singapore Island. *Ann. Trop. Med. Parasitol.* **51**: 17-25.
- Hammon, W. Mc D., rees, D.M., Casals, J. and Meiklejohn, G. (1949). Experimental transmission of japanese encephalitis virus by *Cx.tritaeniorhynchus* and *Cx. pipiens pallens*, suspected natural vectors. *Am.J. Hyg.* **50**; 46-50.
- Hammon, W.M., Tigertt W.D, Salter G. and Schenker H. (1949). Isolation of Japanese B Encephalitis virus from naturally infected *Culex tritaeniorynchus* collected in Japan. *Am. J. Hyg.* **50**:51-56.
- Hammon, W.H., Tigertt, W.D., and Sather, G.E. (1958). Epodemiological studies of concurrent "virgin" epidemics of japanese B. encephalitis and

mumps on Guam, 1947-48; with subsequent observations including dengue, through 1957. *Am.J.Med.Hyg.* **7**(4): 441-467.

Hanna, JN, Ritchie SA, Phillips DA, Lee JM, Hills SL, Vanden Hurk AF, Pyke AT, Johansen CA, Mackenzie JS (1999). Japanese encephalitis in north Queensland, Australia. *Medical Journal of Australia.* **170** (11): 533-536.

Harbison, J.E., Mathenge E.M., Misiani G.O., Mukabana W.R. and. Day J.F . (2006). A simple method for sampling indoor resting malaria mosquitoes *Anopheles gambiae* and *Anopheles funestus* (Diptera: Culicidae) in Africa. *Journal of Medical Entomology.* **43** (3): 473-479.

Henderson, A., Leake and D.S. Burke, 1983. Japanese Encephalitis in Nepal. *Lancet* **8363**: 1359-1360.

Howell, Paul I, and Dave D. chadee (2007). The influence of house construction on the indoor abundance of mosquito. *Journal of vector Ecology* **32** (1): 69-74.

Huang, Y.M., (1972). Contributions to the mosquito fauna of Southeast Asia XIV. The subgenus *Stegomyia* of *Aedes* in Southeast Asia. I-the scutellaris group of species contrib. *Am. Entomol. Inst. (Ann Arbor).* **9** (1): 1-109.

Jeong, Young-Seok and Dong-Kyu lee (2003). Prevalence and seasonal abundance of the dominant mosquito species in a large Marsh near coast of Ulsan. *Journal of Applied Entomology.* **42** (2): 125-132.

Joshi, D.D., wald A. and Joshi A.B. (1995). Japanese encephalitis outbreak during 2046-2049 B.S. (1989/1990-1992/1993) in Nepal. *J. Inst. Med.* **17** (1): 1-11.

Joshi, D.D. (2000). Epidemiological profile of Japanese encephalitis in Nepal, 1966-1997. *J. Inst. Med.* **22**. 23 -31.

Joshi, G., Pradhan S. and Darsie R.F. (1965). Culicine sabethine and taxorhynchitine mosquitoes of Nepal including new country records. *Proc. Entoml. Soc. Wash.* **67**: 137-146.

Johnson, D.R. 1966. Progress toward Malaria eradication in Asia. *Mosq. News* **26**: 478-482.

Jung, R.K. 1973. A brief study on the epidemiology of Filariasis in Nepal. *J. Nepal Med. Assoc.* **11**: 155-168

- Kanojia PC.2006. Rain fall, mosquito density and the transmission of Ross River Virus: A time-series forecasting model in Bellary district, Karnataka. *Ecological Modelling*. **196** (3-4):505-514.
- Kanojia PC 2007. Ecological study on mosquito vectors of Japanese encephalitis virus in Bellary district, Karnataka. *Indian J.Med. Res.* **126** (2): 142-157.
- Kanojia, P.C. and Geevarghese G.(2004). First report on high degree endophilism in *Culex tritaeniorynchus* (Diptera: culicidae) in an area endemic for Japanese encephalitis. *Journal of medical entomology*. **41**(5): 994-996.
- Kanojia PC, Shetty PS, Geevarghese G. (2003). A long-term study on vector abundance and seasonal prevalence in relation to the occurrence of Japanese encephalitis in Gorakhpur district, Uttar Pradesh. *Indian J Med Res.* **117**: 104-110.
- Keating Joseph, Charles M Mbogo, Joseph Mwangangi, Joseph G Nzovu, Vweidong Gu, James L Regens, Guiyun Yan, John I Githure, John C Beier (2005).Anopheles gambiae s.l. and Anopheles funestus mosquito distributions at 30 villages along the Kenyan Coast. *Journal of Medical Entomology*. **42**(3)May:241-246.
- Kent, Rebekah J., Philip E. Thuma, Sungano Maharakurwa and Douglas E. Norris.I. (2007). Seasonality, blood feeding behaviour, and transmission of Plasmodium falciparum by *Anopheles arabiensis* after an extended drought in Southern Zambia. *American Journal of Tropical Medicine and Hygiene*. **76** (2): 267-274.
- Kessel, J.F. 1966. Filariasis as a world problem. *Mosq. News* 26: 192-196.
- Khatri, I.B., D.D. Joshi, T.M.S. Pradhan and S. Pradhan. (1989). Status of viral Encephalitis (Japanese Encephalitis) in Nepal. *J. Nepal Med. Assoc.* **21**: 97-110.
- Kumar Kaushal, Rakesh Katyal and Kuldip Singh Gill (2002). Feeding pattern of Anopheline and Culicine mosquitoes in relation to biotypes and seasons in Delhi and environs. *Journal of communicable Disease*. **34** (1): 59-64.
- Kumar, Yadurappa satish, Nagabhushanrao Ganesh and Achuthan Vijayan (2004). Mosquito diversity in Rajiv Gandhi National Park (Nagarahole), Karnataka state, India. *Journal of the Entomological Research Society*. **6** (2): 1-13.

- Leake, C.J., Ussery M.A, Nisalik ., A., Hoke C.H., Andre R.G. and Burke D.S.. (1986). Virus Isolation from mosquitoes collected during the 1982 Japanese Encephalitis epidemic in Northern Thailand. *Trans. Soc. Trop. Med. Hyg.* **80**: 831-837.R
- Maier, watter A (2002). Effects of environmental changes on vectors of disease in Germany and neighboring countries with special reference to mosquitoes. *Denisia* (6): 535-547.
- Mcintosh, B. M., Jupp P.G., Dickinson D.B., Gillivray G.M. Mc and Sweetman J. (1967). Ecological studies on sindbis and west Nile Viruses in South Africa. I. Viral activity as revealed by infection of mosquitoes and sentinel fowls. *S. Afr. J. Med. Sci.* **32**: 1-14.
- Merdic, Enrih and Ivana Boca (2004). Seasonal dynamics of the *Anopheles maculipennis* complex in Osijek, Croatia. *Journal of vector ecology.* **29** (2): 257-263.
- Mitamure, et.,al. (1938). Isolation of virus of Japanese encephalitis from mosquitoes caught in nature. Report of ninth meeting of the committee on encephalitis, Tokyo. *Iji Shusi* **62**: 820-824.
- Mitchell CJ, Savage HM, Smith GC, Flood SP, Castro LT, Roppul M. (1993). Japanese encephalitis on Saipan: a survey of suspected mosquito vectors. *Am J Trop Med Hyg.* **48** (4): 585-590.
- Molaes, Goudarz, Theodore G. Andreadis, Philip M. Armstrong, Rudy Jr. Bueno, James A. Dennett, Susun V.Real, Chris Sargent, Adilelkhidir Bala, Yvonne Randle, Hilda Guzman, Amelia Travaassos da Rosa, Taweesak wuithiranyagool and Rober B. Teshl (2007). Host feeding pattern of *Culex quinquefasciatus* (Diptera:Culicidae) and its role in transmission of west Nile virus in Harris country, Texas. *American Journal of Tropical Medicine and Hygiene.* **77** (1): 73.81
- Muturi J Ephantus, Josephat Shililu, Benjamin Jacob,Weidong Gu,John Githure, Robert Novak(2006). Mosquito species diversity and abundance in relation to land use in a rice land agro ecosystem in Mwea, Kenya. *J Vector Ecol.* **31** (1) Jun: 129-137.
- Mwandawiro, Charles, Nobuko Tuno, Wannapa Suwonkerd, Yoshio Tsuda, Tetsuo Yanagi and Masahiro Takagi (1999). Host Preference of Japanese

encephalitis vectors in Chiangmai, Northern Thailand. *Medical Entomology and Zoology*. **50** (4): 323-333.

Mwangangi Joseph, Josephat Shiliu, Ephantus Muturi, Weidong Gu, Charles Mbogo, Ephantus Kabaru, Benjamin Jawb, John Githure, Robert Novak (2006). Dynamic of immature stages of *Anopheles arabiensis* and other mosquito species (Diptera: Culicidae) in relation to rice cropping in a rice agro-ecosystem in Kenya. *J. Vector Ecol.* **31** (2): 245-251.

Myrty US, Satyakumar DV, Sriram K, Rao KM, Singh TG, Arunachalam N, Samuel PP. 2002. Seasonal Prevalence of *Culex vishnui* subgroup, the major vector of Japanese encephalitis virus in an endemic district of Andhra Pradesh, India. *J Am Mosq Control Assoc.* **18** (4): 290-293.

Parajuli, M.B., Shrestha S.L., Vaidya R.G. and White G.B. (1981). Nationwide disappearance of *Anopheles minimus Theobald* (1901), previously the principal malaria vector of Nepal. *Trans. R. Soc. Trop. Med. Hyg.* **75**: 603.

Parajuli, M.B., Joshi D.D., Pradhan S.P., Chamling M. and Joshi A.B. (1992). Incidence of Japanese encephalitis during 1989 in Nepal. *J. Nepal Med. Assoc.* **30** (101). 7-14.

Parida, S.K., Hazra R.K., Morai N., Tripathy H.K. and Mahapatra N. (2006). Host feeding patterns of malaria vectors of Orissa, India. *Journal of American Mosquito Control Association.* **22** (4): 629-634.

Peters, W., S.C. Dewar, B.D. Bhalla and T.L. Manandhar. 1955. A preliminary note on the Anophelini of the Rapti Valley area of the Nepal Terai. *Indian J. Malariol.* **9**: 207-212.

Peters, W. and Dewar S. (1956). A preliminary record of the megarrhine and Culicine mosquitoes of Nepal with notes on their taxonomy (Diptera: Culicidae). *Indian J. Malariol.* **10**: 37-51.

Pradhan, J.M., Shrestha S.L. and Vaidya R.G. (1970). Malaria transmission in high mountain valleys of West Nepal including first record of *Anopheles masculatus willmori* (James) as a third vector of malaria. *J. Nepal. Med. Assoc.* **8**: 89-97.

Pradhan, S.P. 1981. Role of mosquitoes in the transmission of Japanese Encephalitis Virus. Siddhartha Jaycees Souvenir 23 Jestha, 2038, pp.6-8.

Pradhan, S.P., 1988. Notes on the mosquitoes of Nepal. *J. Am Mosquito Control. Assoc.* **3**: 21-25.

- Pradhan, S.P., Parajuli, M.B. and Joshi, D.D. (1991). Review of Japanese Encephalitis in Nepal. *J Inst Med.* **13**(3):271-286.
- Pradhan, S.P.; Shrestha, I. Palikhey, N. And Uprety, R.B. (1997). Epidemiological study of lymphatic filariasis in Gokarna VDC of Kathmandu during August and September. *Journal of Nepal Health Research Council, Kathmandu.* **2**:13.
- Prakash, A., Bhattacharya D.R., Mohapatra P.K. and Mahanta J.(1997). Indoor biting behaviour of *Anopheles dirus* Peyton and Harrison, 1979 in upper Assam, India. *Mosquito Borne Diseases Bulletin.* **14** (1-2): 31-37.
- Pramanik, M.K. and Raut S.K.(2002). Human dwelling and cattle shed mosquitoes of Kolkata: Past and present. *Journal of National Taiwan Museum.* **55** (1): 1-16.
- Rajendran, R., Thenmozhi V., Tewari, S.C., Balasubramanian, A. Ayanar, K., Manavalan, R., Gajanana, A. , Kabilan, L., Thakore, J.P. and Satyanarayana, K. (2003). Longitudinal studies in South Indian villages of Japanese encephalitis virus infection in mosquitoes and seroconversion in goats. *Tropical Medicine and International Health.* **8** (2): 174-181.
- Reuben, R. 1971 a. Studies on the mosquitoes of North Arcot District, Madras State, India. Part 5 Breeding places of the *Culex vishnui* group of species. *J. Med. Entomol.* **8**: 363-366.
- Reuben, R. (1971 b). Studies on the mosquitoes of North Arcot District, Madras State, India. Part 6. Seasonal prevalence of the *Culex vishnui* group of species. *J. Med. Entomol.* **8**: 367-371.
- Richard, F. Darsie, Jr. and Pradhan Shreedhar P. (1990). the mosquitoes of Nepal : Their Identification, Distribution and Biology. **22** (2).
- Richards, Stephanie L., Loganathan Ponnusamy, Thomas R. Unnasch, K. Hassan hassan and Charles S. Apperson 2006.Host feeding patterns of *Aedes albopictus* (Diptera: culicidae) in relation to availability of human and domestical animals in suburban landscapes of Central North Carolina. *Journal of Medical Entomology.* **43** (3): 543-551.
- Russel, C.R. 2005. Species diversity of mosquito in Florida. *J. Florida. Entomol. SOC.* **66**:4-31

Sharma, R.S., Kaul S.M. and Sokhay Jotna. (2005). Seasonal fluctuations of dengue fever vector, *Aedes aegypti* (Diptera:Culicidae) in Delhi, India. *South East Asian Journal of Tropical Medicine and Public Health*. **36** (1): 186-190.

Sherchand, J.B., Obsomer, V., Thakur, G.B. and Hommel, M. (2003). Mapping of lymphatic filariasis in Nepal. *Journal of Filaria, Nepal*, **2**(7): 16-26.

Shirasaka A., Sasa, M., Wada Y. (1969). Notes on laboratory colonization of *Cx. tritaenirhynchus summorosus*, the principal vector of Japanese encephalitis in Japan. *Jap.J.Exp.Med.* **69**(6): 423-436.

Shrestha, S.L. 1966. Mosquitoes. Pp.51-66. In: Wild is Beautiful.

Shrestha, S.L. and M.B. Parajuli, 1980. Reappearance of malaria in terai areas of Nepal and incrimination of *An. Annularis van der wulp*. *J. Nepal Med. Assoc.* **18**: 11-18.

Shrestha, S.L., Pradhan, S., Shrestha, J.P.B. , Shrestha, J.D. , Rajbhandari, Y. , Shrestha, G.L. , Swar, T.B. , Nusin, M.K. and Reisen. W.K. (1988). Observation on Anopheline and malaria ecology in the far western region of Nepal, 1986. *Bull. Soc. Vector Ecol.* **13**: 332-342.

Simpson, D.I.H., Bowen, E.J.W., Way, H.J. et al. (1874). Arbovirus infection in Sarawak, October, 1968- Feb. 1970: Japanese encephalitis virus isolation from mosquitoes. *Annals. Trop.Med. Parasit.* **68**: 393-404.

Smallegange, Renate, Yu Tong Qiu, Agnes M.S. Galimard, Maarten A. Posthumus, Teris A. Van Beek, Joop J.A. Van Loon and Willem Takken (2003). Why humans are attractive to malaria mosquitoes. *Entomologische Berichten (Amsterdam)* **63** (3):50-53.

Takashima I, Hashimoto N, Watanabe T, Rosen L. (1989). Mosquito collection in endemic areas of Japanese encephalitis in Hokkaido Japan. *Nippon Juigaku Zasshi.* **51** (5): 947-953.

Tandon Neelam and Ray Sudipta (2000). Host feeding pattern of *Aedes aegypti* and *Aedes albopictus* in Kolkata India. *Dengue Bulletin* (24): 117-120.

Ulloa Armandu, Gonzalezceron Lilia and Rodriguez. Mario H. (2006). Host selection and gonotrophic cycle length of *Anopheles punctimacula* in southern Mexico. *Journal Mosquito of the American Control Association.* **22** (4):648-653.

Wang, S.p., Grayston, J.T., Hu, S.M.K. (1962) Encephalitis on Taiwan. III. Virus isolation from mosquitoes. *Am. J. Trop. Med. Hyg.* **11**: 141- 148.

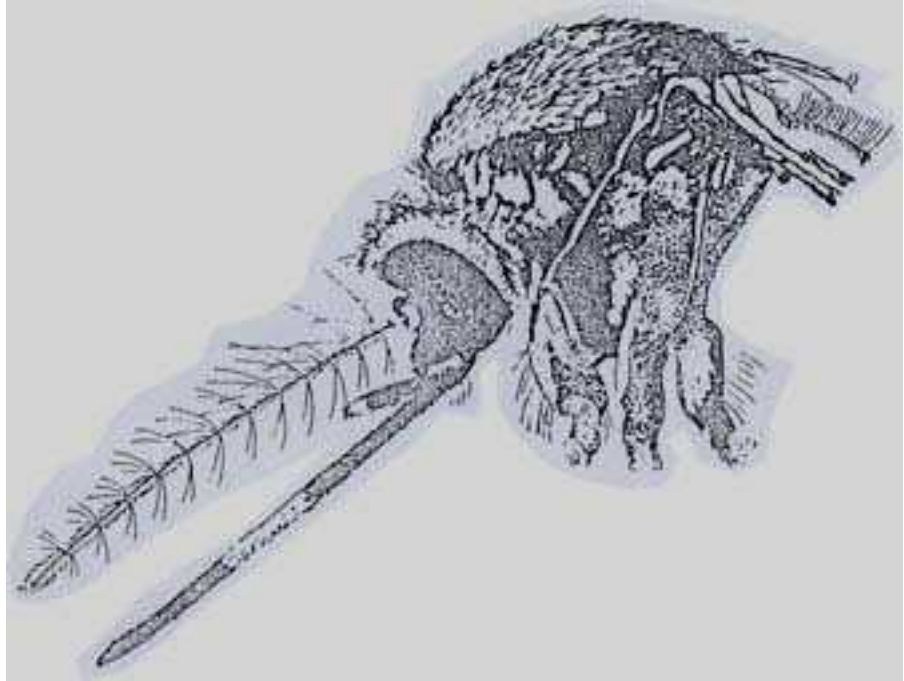
Zhou Guofa, Minakawa Noboru, Githeko Andrew and Yan Guiyun (2004). Spatial distribution patterns of malaria vectors and sample size determination in spatially heterogeneous environments. *Journal of Medical Entomology*. 41 (6): 1001-1009.

Zimmerman, M.D., Scott, R.M., Vaughn, D.W., Raj Bhandari, S. , Nisalak, A. and Shrestha M.P. (1997). Short report: An outbreak of Japanese encephalitis in Kathmandu, Nepal. *Am.J.Trop. Med. Hyg.* **57** (3): 283-284.

Pictorial of *Culex* (Female)

Figure a

Pleuron with distinct scale patches at least on upper and lower mesokatepisternum and anterior mesanepimeron



Pictorials of the vector i.e. *Culex fuscocephala*

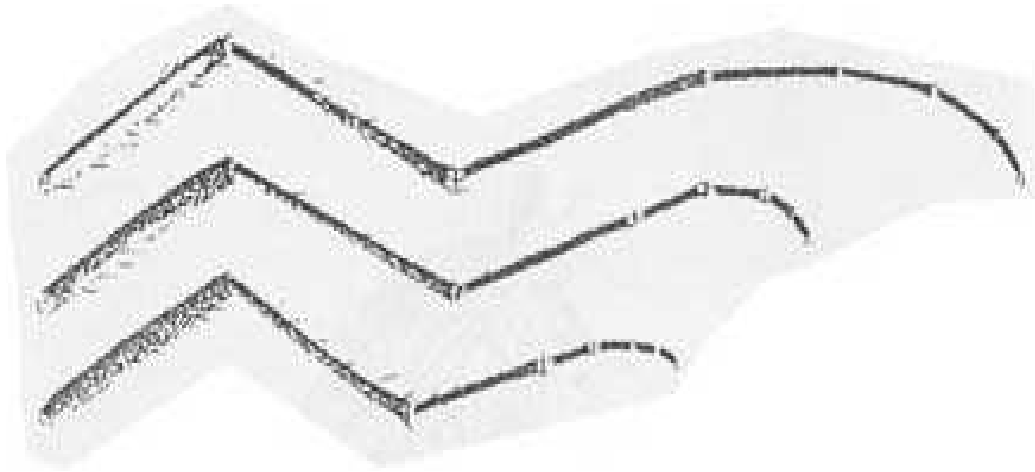


Figure b: Anterior surface of mid-femur without median longitudinal pale-scaled stripe



Figure c: Abdominal terga without basal transverse, pale-scaled bands

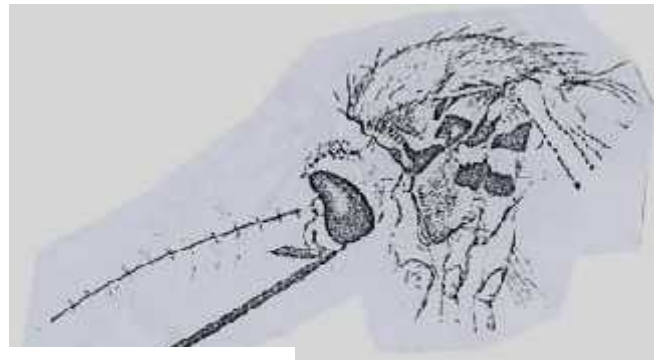


Figure d: Pleura of dark and pale

Pictorials of *Culex whitei*

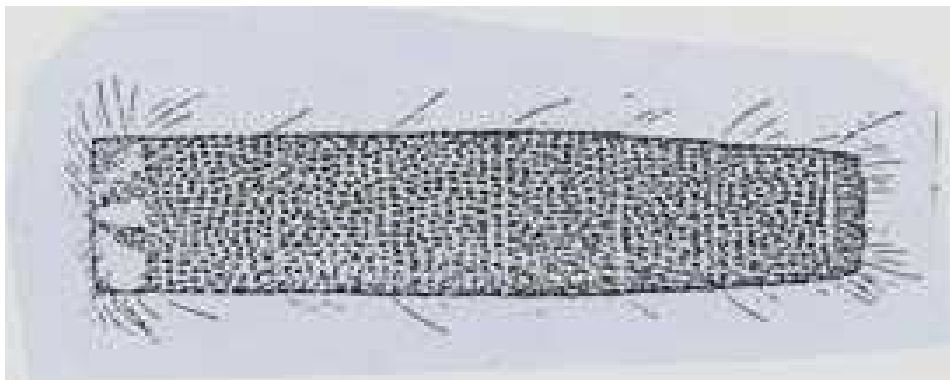


Figure e: Abdominal terga II-VI entirely dark scaled, without pale bands and epicolateral pale patches

Pictorials of *Culex sinensis*

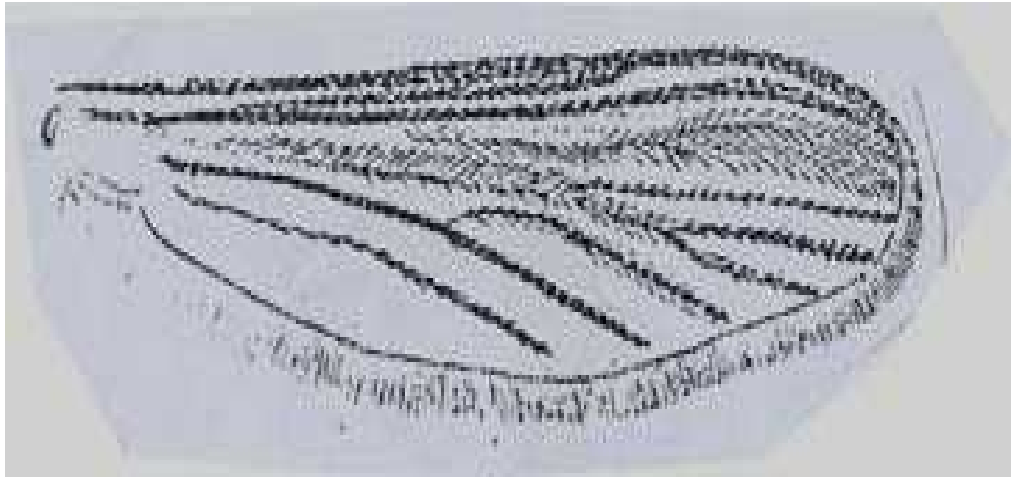


Figure f: wing with dark scales on all veins

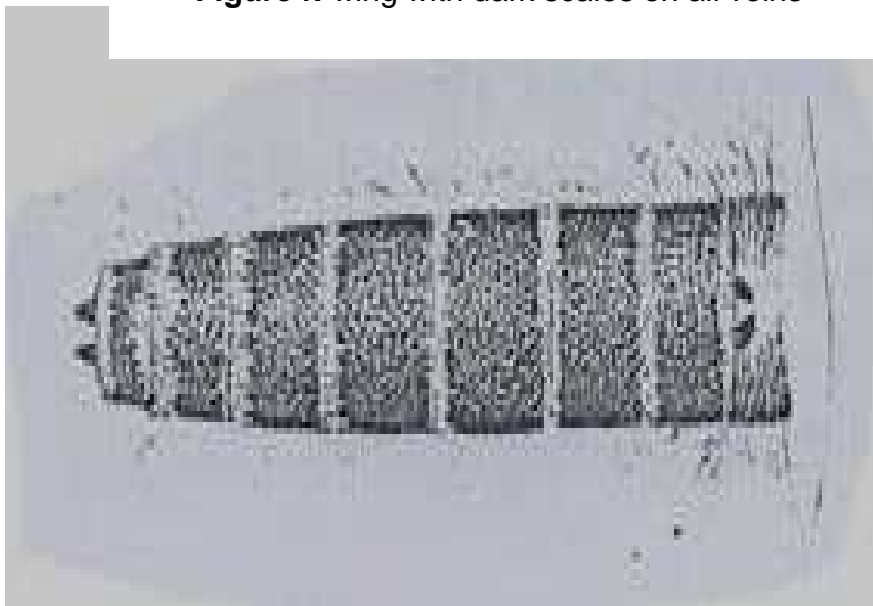


Figure g: Abdominal terga II-VI with dark areas not sprinkled with pale scaled

Pictorial of *Culex tritaeniorhynchus*

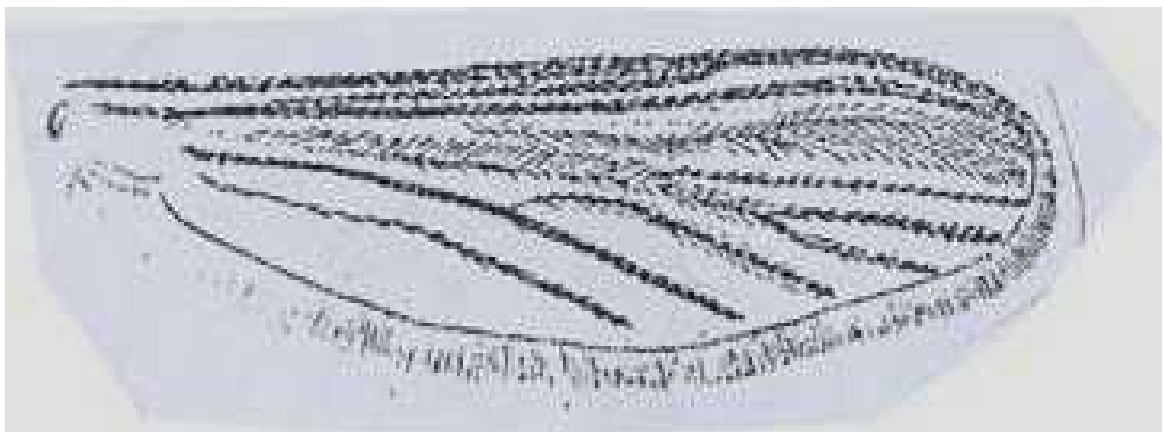


Figure h: wing with dark areas sprinkled with pale scales

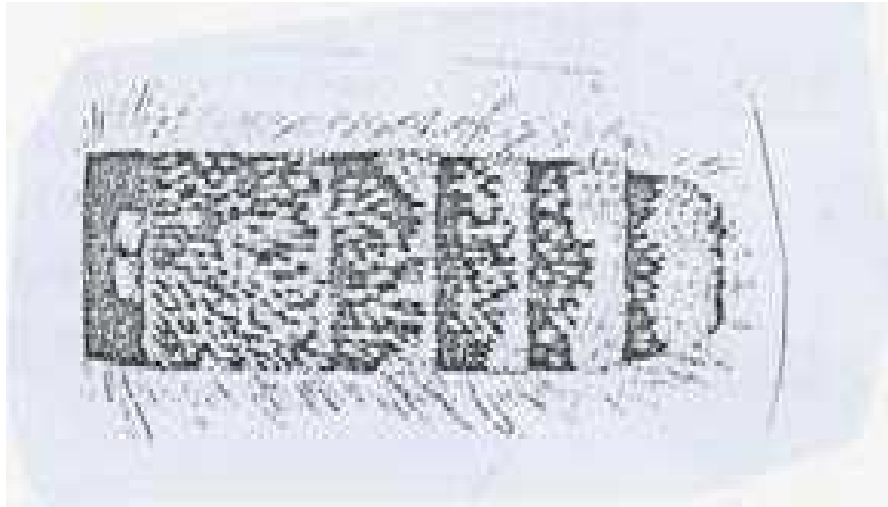


Figure i: Abdominal terga II-VI with dark areas sprinkled pale scaled

Pictorials of the vector i.e. *Culex gelidus*

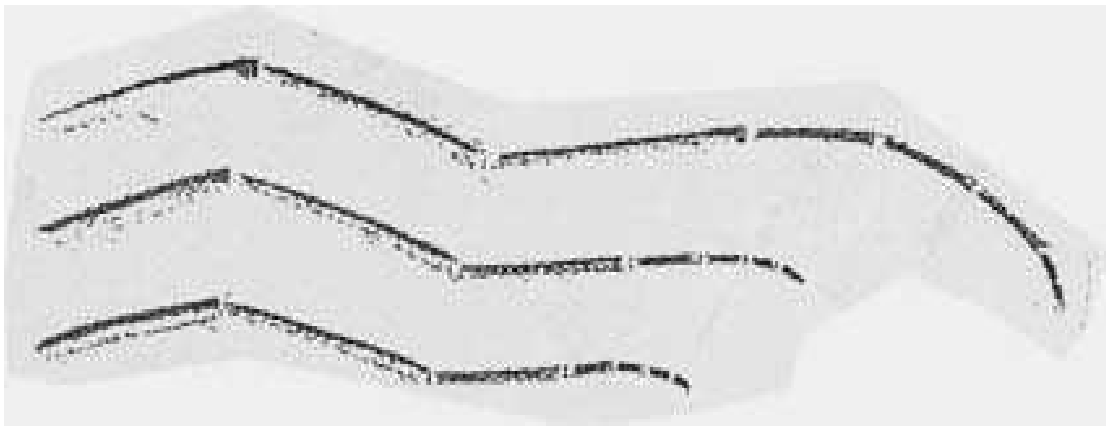


Figure j: Anterior surface of fore and mid femora without speckling of pale scales

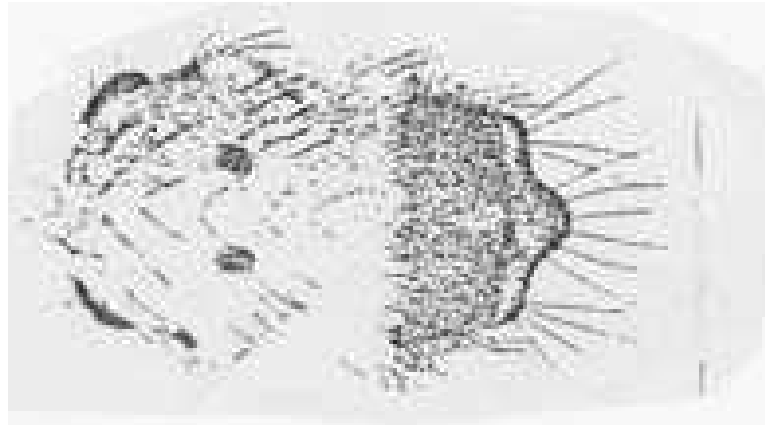


Figure k: Scales on lateral scutal area behind wing base, prescutellar area and on scutellum entirely

Pictorials of *Culex whitmorie*

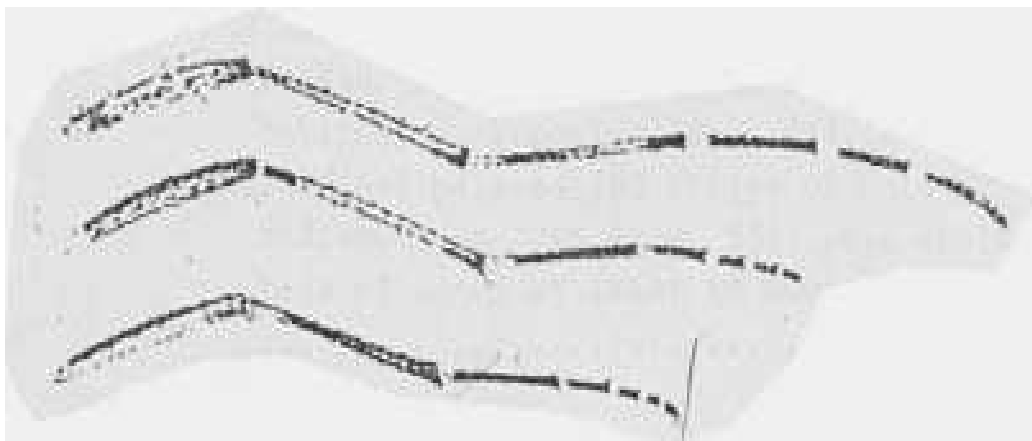
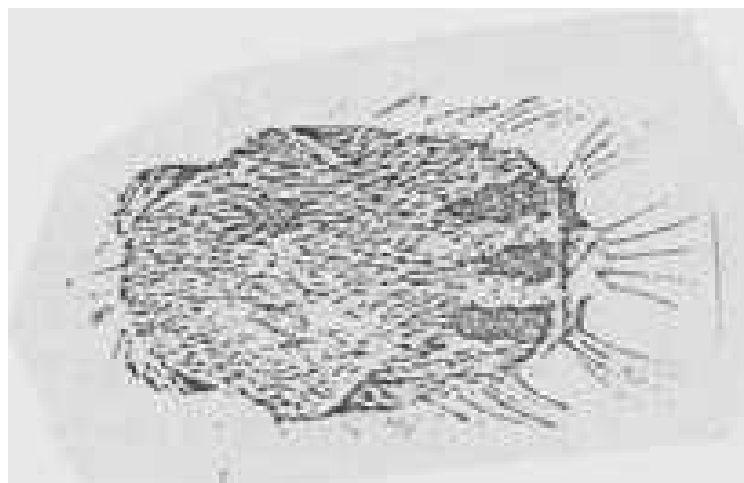


Figure l:
Anterior surface of fore and mid femora extensively speckled with pale scale

Figure
Scales on
scutal
wing



m:
lateral
area behind
base,
prescutellar

arean and on scutellum predominantly pale

Pictorials of *Culex barraudi*

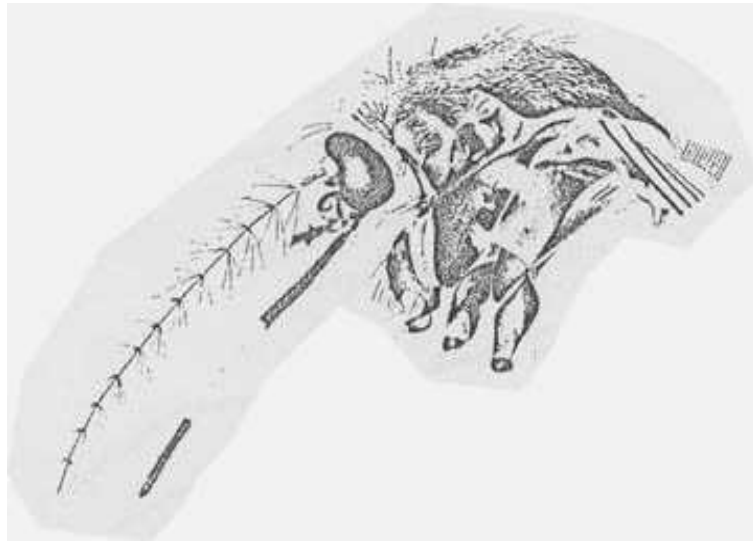
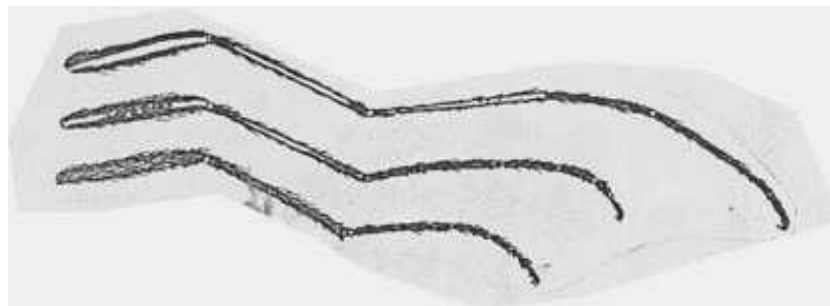


Figure n:
Postspiracular
with smooth
of semi erect
scales on lower
aspect

area
patch



o:

Figure

Longitudinal pale-scaled stripe on anterior surface of midfemur
broken into small spots at middle

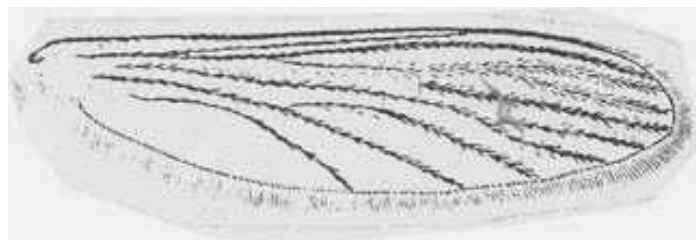


Figure p:
Costal vein
-scales

entirely dark

Pictorials of the vector i.e. *Culex tritaeniorhynchus*

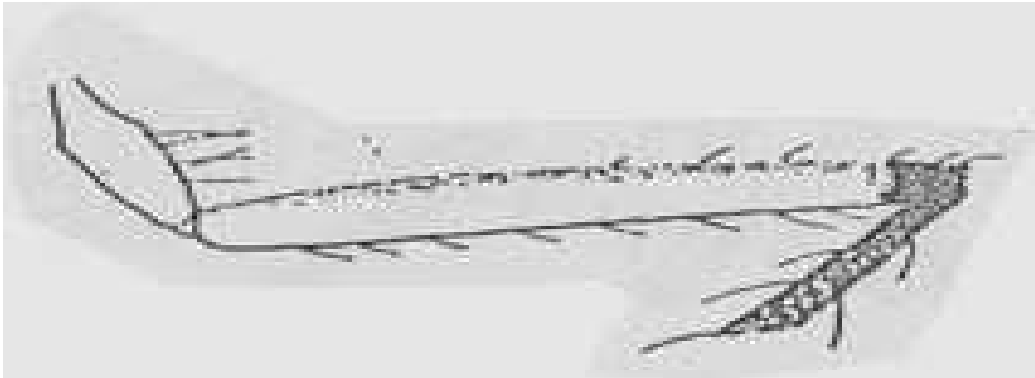


Figure q:
Anterior surface of hind femur pale scaled with narrow black scaled ring apically

Pictorials of the vector i.e. *Culex psuedovishnui*

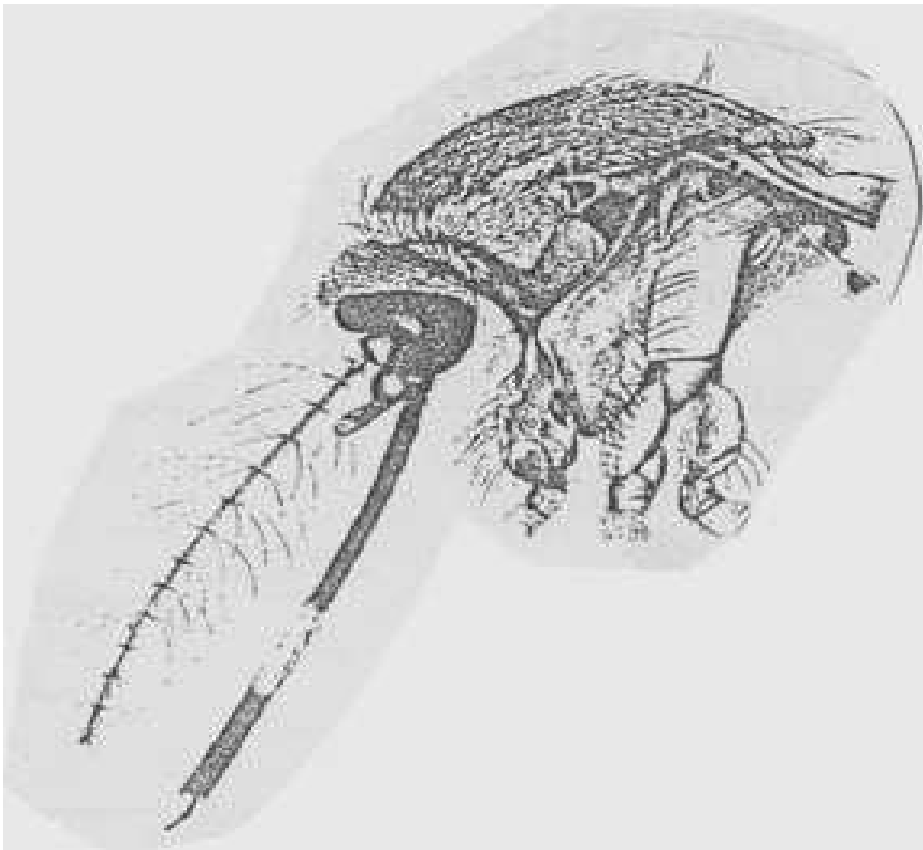
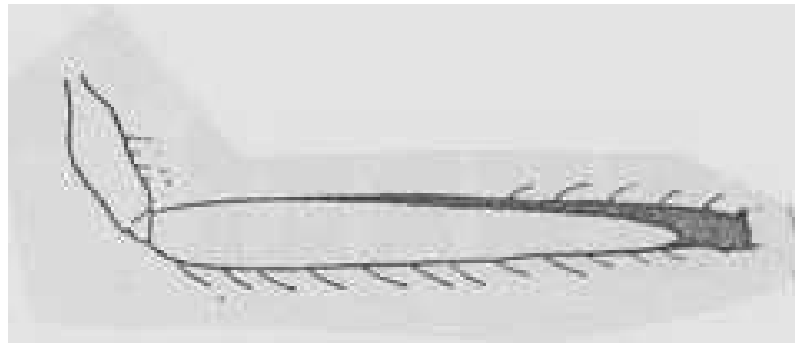


Figure r:
scutum with yellow to silvery scales

Figure s:
Femora
proboscis
speckled
pale scales



and
never
with

Pictorials of *Culex mimulus*

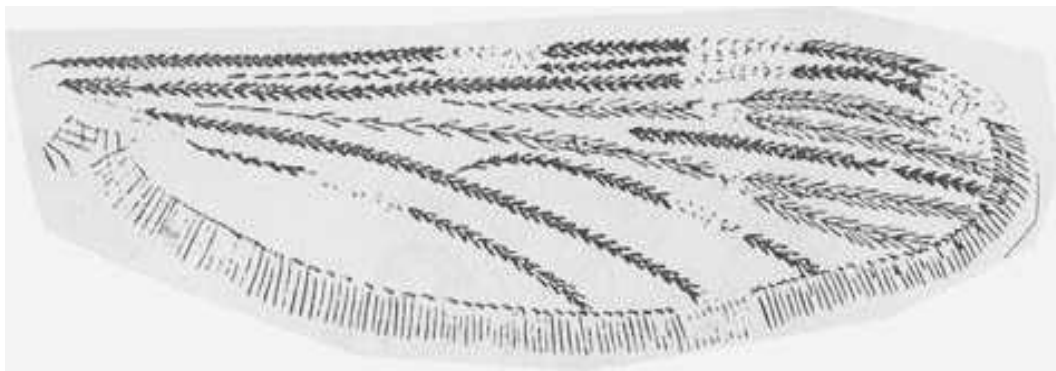
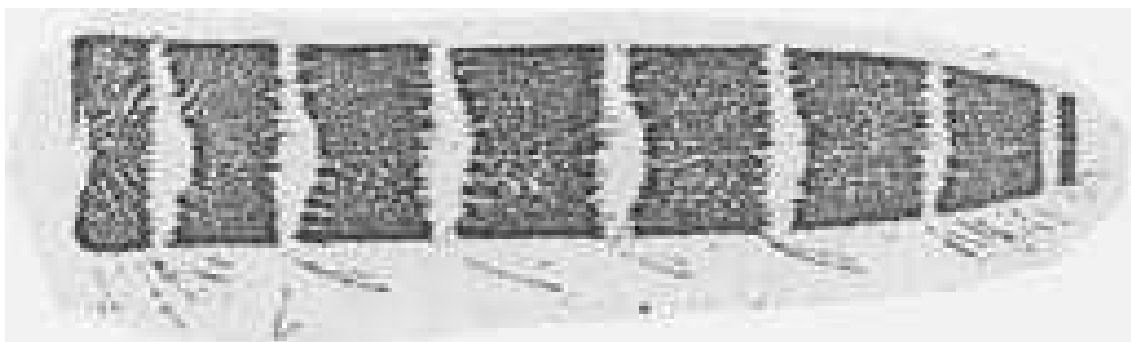


Figure t:
Second pale scaled costa spot involves veins C, Sc, R and sometimes R_s
and Cu

Figure u:
Basal pale bands of abdominal terga narrow, usually less than 0.25 length of



segment

