# STUDY OF PROTEASE ACTIVITY OF BACTERIA ISOLATED FROM SOLID WASTE

A DISSERTATION SUBMITTED TO THE CENTRAL DEPARTMENT OF MICROBIOLOGY TRIBHUVAN UNIVERSITY

#### IN PARTIAL FULFILLMENT OF THE REQUIREMENTS THE AWARD OF THE DEGREE OF MASTER OF SCIENCE IN MICROBIOLOGY (ENVIRONMENT AND PUBLIC HEALTH)

BY SMRITI MAINALI

CENTRAL DEPARTMENT OF MICROBIOLOGY TRIBHUVAN UNIVERSITY KIRTIPUR, KATHMANDU, NEPAL 2009

### RECOMMENDATION

This is to certify that Ms **Smriti Mainali** has completed this dissertation work entitled **"Study of protease activity of bacteria isolated from solid waste"** as a partial fulfillment of M. Sc. degree in Microbiology under my supervision. To the best of my knowledge this is an original research work of her and this work has not been submitted for award of any other degree.

.....

Mr. Binod Lekhak Lecturer Central Department of Microbiology Tribhuvan University Kirtipur, Kathmandu, Nepal

Date: .....

### **CERTIFICATE OF APPROVAL**

On the recommendation of **Mr. Binod Lekhak**, this dissertation work of Ms **Smriti Mainali** entitled **"Study of protease activity of bacteria isolated from solid waste"** has been approved for the examination and is submitted to Tribhuvan University in partial fulfillment of the requirements for Masters of Science in Microbiology.

.....

Dr. Dwij Raj Bhatta (M. Sc., Ph.D. in Microbiology) Head of the Department Central Department of Microbiology Tribhuvan University Kirtipur, Kathmandu, Nepal

Date: .....

### **BOARD OF EXAMINERS**

#### **Recommended by:**

Mr. Binod Lekhak Supervisor

Approved by:

# Dr. Dwij Raj Bhatta, M.Sc, Ph.D.

Head of the Department

\_\_\_\_\_

**Examined by:** 

#### Dr. Tika Bahadur Karki

Professor and Head of the Department Department of Biotechnology Kathmandu University External Examiner

#### Dev Raj Joshi

Lecturer Internal Examiner

Date: \_\_\_\_\_

### ACKNOWLEDGEMENTS

I would like to express my sincere gratitude and appreciation to my supervisor **Mr. Binod Lekhak,** Lecturer, Central Department of Micorbiology for his valuable suggestions, comments, continuous guidance and support throughout this work.

I am equally indebted to **Dr. Dwij Raj Bhatta**, Associate Professor and Head, Central Department of Microbiology for providing me the laboratory facilities to carry out this work.

My special regards goes to **Mr. Dev Raj Joshi**, Lecturer, Central Department of Microbiology for his guidance, constant inspiration and valuable suggestions during the whole study period. I would like to acknowledge **Mrs. Reshma Tuladhar**, Lecturer, Central Department of Microbiology and **Mr. Janardan Pandey**, Central Department of Biotechnology for their valuable suggestions and guidance during this work. I am specially thankful to **Mr. Ramesh Khadka**, **Mr. Madhukar Thapa**, **Mr. Navaraj Karki**, **Mr. Raiman Shakya** and all the laboratory staffs of Central Department of Microbiology for their kind co-operation for accomplishment of this dissertation work.

I am indebted to Ratnashree, Mrs. Niru Rana, my parents and family members, my husband and in-laws for their continuous support, inspiration, encouragement and co-operation during this study period.

I would like to acknowledge my friends Nanu Maiya Khadka, Bijaya Laxmi Maharjan, Ram Prasad Awal, Esha Shrestha, Saurav Aryal, Manita Aryal, Shiv Nandan Sah and Geeta Pandey for their supportive contribution and suggestions during the study period.

.....

Smriti Mainali

Date: \_\_\_\_\_

### ABSTRACT

The study was carried out at Central Department of Microbiology, Tribhuvan University, Kirtipur from September 2008 to May 2009. Random sampling was adopted to collect altogether 20 samples of municipal solid waste from different sites within Kahtmandu Valley. The temperature of the solid waste was recorded at site using mercury thermometer while pH was recorded after transporting the samples to the laboratory. The indigenous proteolytic bacteria were isolated by spread plate technique on mineral base agar supplemented with 1% gelatin. Out of 113 isolates obtained, only 22 degraded gelatin incorporated in agar media and the rest were chemolithotrophs. On the basis of degree of hydrolysis, 3 potent gelatinase producers (laboratory code: K2.2i, R3.3i and R5.2iv) with diameter of zone of hydrolysis more than 20 mm were selected and identified on the basis of their morphological, cultural and biochemical characteristics. The isolates K2.2i and R5.2iv were *Micrococcus* spp. while R3.3i remained unidentified. Enzyme was extracted from these most potent isolates by fermentation. The crude enzyme extracts were assayed for secondary screening by cup plate assay and the isolates K2.2i and R3.3i exhibited similar gelatinase activity while R5.2iv was slightly less active. The enzymes from K2.2i and R3.3i were purified by acetone precipitation. The kinetics (temperature and pH) was studied and enzyme from K2.2i was found optimally active at 4<sup>o</sup>C and pH 8 while that from R3.3i was optimally active at  $37^{\circ}$ C and pH 9. The enzymes were further purified by ammonium sulphate fractionation. The 30%, 60% and 90% fractions of enzyme from K2.2i and only 30% and 60% fractions of enzyme from R3.3i revealed gelatinase activity during assay at 37<sup>o</sup>C and pH 7. The activities of 60% fractions of both the enzymes were the highest while studied at different temperatures and pH. The obtained data was analyzed for analysis of variance (ANOVA) and paired samples t-test using statistical software (SPSS version 11.5). From these analyses, significant effect of temperature (P=0.045 and P=0.014for enzymes from R3.3i and K2.2i respectively) and pH (P=0.032 and P=0.078 for enzymes from R3.3i and K2.2i respectively) was observed on the enzyme activity. Also, ammonium sulphate fractionation can be significantly applied for purification of the enzymes. From the study of the temperature and pH condition of the piled up solid waste and the activity profile of the obtained enzymes, it can be concluded that both of these enzymes are suitable for their application in solid waste management; enzyme from K2.2i in the cold seasons while that from R3.3i at in warmer seasons. The study could be extended to field trial.

#### Key words: Solid waste, protease activity, proteolytic bacteria

# **TABLE OF CONTENTS**

	Page No.
Title Page	i
Recommendation	ii
Certificate of Approval	iii
Board of Examiners	iv
Acknowledgement	V
Abstract	vi
Table of Contents	vii – ix
List of Abbreviations	X
List of Tables	xi
List of Figures	xii
List of Photographs	xiii
List of Appendices	xiv
<b>CHAPTER – I INTRODUCTION</b>	1 – 2
CHAPTER – II OBJECTIVES	3
CHAPTER – III LITERATURE REVIEW	4 – 26
3.1 Solid Waste	4
3.2 Enzymes	7
3.3 Classification of Enzymes	8
3.4 Protease	9
3.5 Classification of Protease	10 – 13
3.5.1.1 Exopeptidases	10
3.5.1.1.1 Aminopeptidases	10
3.5.1.1.2 Carboxypeptidases	10
3.5.1.2 Endopeptidases	11 - 12
3.5.1.2.1 Serine Proteases	11
3.5.1.2.1.1 Serine Alkaline Proteases	11
3.5.1.2.1.2 Subtilisins	11
3.5.1.2.2 Aspartic Proteases	12

3.5.1.2.3 Cysteine/thiol Proteases	12
3.5.1.2.4 Metalloproteases	12
3.5.2.1 Acid Proteases	13
3.5.2.2 Alkaline Proteases	13
3.5.2.3 Neutral Proteases	13
3.6 Sources of Proteases	13 – 18
3.6.1 Plant Proteases	14
3.6.2 Animal Proteases	15
3.6.3 Microbial Proteases	16 – 18
3.6.3.1 Bacteria	17
3.6.3.2 Fungi	17
3.6.3.3 Actinomycetes	17
3.6.3.4 Viruses	18
3.7 Purification and Characterization of Proteases	18 – 24
3.7.1 Effect of Temperature	19
3.7.2 Effect of pH	21
3.7.3 Effect of Substrate Concentration	22
3.8 Application of Proteases	24 - 26
3.8.1 Detergents	24
3.8.2 Leather Industry	24
3.8.3 Food Industry	25
3.8.4 Pharmaceutical Industry	25
3.8.5 Other Applications	25
CHAPTER – IV MATERIALS AND METHODS	27 – 30
4.1 Materials	27
4.2 Methods	27 - 30
4.2.1 Study Design and Sampling Site Selection	27
4.2.2 Sample Collection and Transportation	27
4.2.3 Laboratory Processing	27 – 29
4.2.3.1 Primary Screening of Gelatin Degrading Bacteria	27
4.2.3.2 Identification of Screened Bacteria	28
4.2.3.3 Enzyme Extraction	28
4.2.3.4 Assay of Enzyme Activity	28
4.2.3.5 Secondary Screening of Bacteria	28

4.2.3.6 Study of Enzyme Kinetics	29
4.2.3.6.1 Enzyme Activity at Different Temperatures	29
4.2.3.6.2 Enzyme Activity at Different pH	29
4.2.3.7 Salt Fractionation of the Protease	29
4.2.4 Statistical Analysis	29
CHAPTER – V RESULTS	31 – 37
5.1 Distribution of Sampling Sites	31
5.2 Screening and Identification of Bacteria	32
5.3 Kinetics of Partially Purified Enzymes	33
CHAPTER – VI DISCUSSION AND CONCLUSION	38 – 49
6.1 Discussion	38
6.2 Conclusion	<b>49</b>
CHAPTER – VII SUMMARY AND RECOMMENDATION	50 – 52
7.1 Summary	50
7.2 Recommendations	51
CHAPTER – VIII REFERENCES	53 - 62
APPENDICES	I – XII

# LIST OF ABBREVIATIONS

- AIDS Acquired Immuno Deficiency Syndrome
- ANOVA Analysis of Variance
- CBS Central Bureau of Statistics
- ICIMOD International Centre for Integrated Mountain Development
- IUCN International Union for Conservation of Nature
- KMC Kathmandu Metropolitan City
- MOPE Ministry of Population and Environment
- WHO World Health Organization

## **LIST OF TABLES**

- Table 3.1
   Solid waste types associated with various source classification
- Table 3.2Waste material by kind and composition
- Table 3.3
   International classification of enzymes
- Table 5.1pH and temperature records of samples collected
- Table 5.2Identification of the potent producers of gelatinase
- Table 5.3Secondary screening
- Table 5.4
   Effect of temperature on the activity of salt fractionated enzymes
- Table 5.5Effect of pH on the activity of salt fractionated enzymes

# **LIST OF FIGURES**

- Figure 3.1 Effect of substrate concentration on the rate of enzyme activity
- Figure 4.1 Flowchart for the isolation and characterization of substrate degrading bacteria
- Figure 5.1 Primary screening of the isolates for extracellular protease on 1% gelatin agar
- Figure 5.2 Effect of temperature on the activity of partially purified enzyme
- Figure 5.3 Effect of pH on the activity of partially purified enzyme

### LIST OF PHOTOGRAPHS

- Photograph 1 Isolated colonies of bacteria on mineral base agar with 1% gelatin.
- Photograph 2 Secondary screening of gelatin degrading bacteria on gelatin agar
- Photograph 3 Culture of isolate R3.3i on nutrient agar
- Photograph 4 Microphotograph showing gram staining of isolate R3.3i
- Photograph 5 Culture of isolate K2.2i on nutrient agar
- Photograph 6 Microphotograph showing gram staining of isolate K2.2i

## LIST OF APPENDICES

- Appendix I Equipments and Media/Chemicals/Reagents
- Appendix II Composition and preparation of media and reagents
- Appendix III Test procedures
- Appendix IV Identification charts
- Appendix V Ammonium sulphate fractionation chart
- Appendix VI Data analysis by SPSS 11.5 version