

**ASSESSMENT OF ARSENIC TOLERANT BACTERIA FROM
ARSENIC CONTAMINATED GROUNDWATER IN NAWALPARASI
DISTRICT OF NEPAL**

**A
DISSERTATION
SUBMITTED TO THE
CENTRAL DEPARTMENT OF MICROBIOLOGY
TRIBHUVAN UNIVERSITY**

**IN PARTIAL FULFILLMENT OF THE REQUIREMENT FOR THE
AWARD OF THE DEGREE OF MASTER OF SCIENCE IN
MICROBIOLOGY
(ENVIRONMENT AND PUBLIC HEALTH)**

**BY
SHREE KRISHNA SHRESTHA**

**CENTRAL DEPARTMENT OF MICROBIOLOGY
TRIBHUVAN UNIVERSITY
KIRTIPUR, KATHMANDU, NEPAL
2008**

RECOMMENDATION

This is to certify that **Mr. Shree Krishna Shrestha** has completed this dissertation work entitled “**ASSESSMENT OF ARSENIC TOLERANT BACTERIA FROM ARSENIC CONTAMINATED GROUNDWATER IN NAWALPARASI DISTRICT OF NEPAL**” as a partial fulfillment of M. Sc. Degree in Microbiology under our supervision. To our knowledge, this work has not been submitted for any other degree.

Dr. Anjana Singh
Head
Central Department Microbiology
Tribhuvan University

Dr. Linda S. Smith
Country Director
Filters for Families
Lalitpur

Mr. Makhan Maharjan
Program Manager
Environment and Public Health
Organization, Kathmandu

Date:

CERTIFICATE OF APPROVAL

On the recommendation of **Dr. Anjana Singh, Dr. Linda S. Smith** and **Mr. Makhan Maharjan**, this dissertation work of **Mr. Shree Krishna Shrestha**, entitled “**ASSESSMENT OF ARSENIC TOLERANT BACTERIA FROM ARSENIC CONTAMINATED GROUNDWATER IN NAWALPARASI DISTRICT OF NEPAL**” has been approved for the examination and is submitted to the Tribhuvan University in the partial fulfillment of the requirement for Master’s Degree of Science in Microbiology.

Dr. Anjana Singh, Ph.D.

Head

Central Department of Microbiology

Tribhuvan University, Kirtipur

Kathmandu, Nepal

Date:

BOARD OF EXAMINERS

Recommended by:

.....
Dr. Anjana Singh
Supervisor

.....
Dr. Linda S. Smith
Supervisor

.....
Mr. Makhan Maharjan
Supervisor

Approved by:

.....
Dr. Anjana Singh
Head of Department

Examined by:

.....
Dr. Alenka H. Radojic, Ph. D.
External Examiner

.....
Mr. Binod Lekhak
Internal Examiner

Date

ACKNOWLEDGEMENTS

It is my heartiest pleasure to express my sincere gratitude to my supervisor **Dr. Anjana Singh**, Head, Central Department of Microbiology, Tribhuvan University whose expert guidance and encouragement led me to the completion of this thesis work.

I am also indebted to my supervisors **Dr. Linda S. Smith**, Country Director, Filters for Families, Bakundole, Lalitpur and **Mr. Makhan Maharjan**, Program Manager, Environment and Public Health Organization, New Baneshwor for supervision and constant encouragement for the completion and accomplishment of this work.

I am thankful to Dr. Shreekant Adhikari, Dr. Dwij Raj Bhatta, Mr. Binod Lekhak and Ms. Shaila Basnyat for their valuable idea, moral support, and kind co-operation during this study. I am much obliged to Mr. Megha Raj Banjara, Mr. Dev Raj Joshi, Mr. Kiran Babu Tiwari and all the teachers of Central Department of Microbiology for their tremendous support and valuable suggestions during the research work.

I acknowledge profound gratitude to the Parasi Hospital and its staff for providing me laboratory and other facilities. I am immensely grateful to Mr. Somai Kurmi, Mr. Arjun Gupta, Mr. Rajan Poudel, Mr. Sushil Tuladhar and all the families of FFF. I am also gratefully indebted to Ms. Dikshya Pokharel, College of Applied Science.

I am deeply grateful to Mr. Ramesh Khadka, Mr. Rayman Shakya, Mr. Madhukar Thapa, Mr. Nawaraj Karki and other staffs of Central Department of Microbiology for their help. I highly appreciate for kind cooperation and genuine support by my colleagues Narendra Maden, Sanjiv Neupane, Pankaj Baral, Manoj Khadka, Kashi Ghimire and Anil Shrestha.

I sincerely appreciate cooperation received from all my family members especially Ram Krishna Shrestha, Pancha Maya Shrestha, Dr. Ramesh Makaju, Sanu Maya Makaju and my brother Rabi Shrestha.

At last, I would like to dedicate this thesis work to the peoples of Nawalparasi district, especially peoples of Ramgram Municipality; Sunwal, Sarawal, Sukrauli, Swathi, Pratappur and Jahada VDCs.

Date:

Shree Krishna Shrestha

ABSTRACT

Arsenic, considered a human carcinogen, contamination in groundwater drinking water is a worldwide public health issue, including Nepal. To assess arsenic tolerant bacteria from arsenic contaminated drinking water, this research study was carried out from June 2007 to January 2008. Eighty-six water samples were collected from one municipality and six VDCs' wards in Nawalparasi district of Nepal having arsenic concentration above WHO guidelines (10 ppb).

Physico-chemical and microbiological parameters of water quality were determined to be poor. The pH of 11.63% water samples was found below WHO guideline. Sixty two percent (53/86) samples in Nawalparasi district showed arsenic level above Nepal drinking water quality standard (NDWQS) (50 ppb). Filtered water samples showed that 30.16% (19/63) was above NDWQS guideline. Comparison between arsenic concentration of influent and effluent water showed significant difference ($p < 0.05$). Within the last 10 years, 66.27% (57) tubewells were constructed whereas 52.63% (30/57) tubewells constructed within 6-10 years. Moderately positive correlation ($r = 0.23$, $p = 0.83$) was found between arsenic concentration and age of tubewells. At 36-95 feet depth range, 50 (58.14%) tubewells were above NDWQS and 70 (81.39%) tubewells above the WHO guideline. Results showed weakly negative correlation ($r = -0.12$, $p = 0.24$) indicating arsenic concentration of groundwater decreases as the depth of tubewell increases.

Skin manifestations were observed in 12 (34%) females and 18 (33%) males, skin manifestations and gender difference was found statistically insignificant ($p > 0.05$). Arsenicosis cases were highest (33.3%, 10/30) in 32-41 age groups. Most of the patients (30%) suffered from diffused arsenical melanosis and 20% patients from both diffused arsenical melanosis and keratosis. Mild skin manifestation were observed in most, 66.7% (20/30) patients followed by 7 (23.3%) moderate and 3 (10%) severe. Percentage of arsenicosis cases was higher among respondents using tubewell that were built >10 years (41%) than that were built 10 years (32%).

The microbiological analysis of groundwater revealed the presence of total coliforms in 98.84% samples (85) of influent water from tubewell and 71.43% (45/63) effluent water from filter. Among the isolates, *E. coli* (37%) was found to be predominant in influent water which was followed by *P. aeruginosa* (16%), *Citrobacter* spp. (16%) and *K. pneumoniae* (10.5%). Similarly, *E. coli* (39.21%) was found predominant in effluent water which was followed by *P. aeruginosa* (19.61%), *K. oxytoca* (13.72%) and *Citrobacter* spp. (13.72%). Among thirteen arsenic tolerant bacteria isolated from plate count agar incorporated with As (V) >50 ppm, the most predominant isolates were unidentified NP1 (Bluish green) (19.51%) followed by *Pseudomonas* spp. (16.38%), *Bacillus cereus* (11.15%). The mean colony count of samples (<300 cfu/ml) was highest in 10 ppm (216 cfu/ml) and lowest in 200 ppm (70 cfu/ml). The highest MIC value of As (III) was obtained for *E. coli* (mean=15 ppm) followed by NP1 (Bluish green) (mean=10.71ppm). Similarly the highest MIC value of As (V) was obtained for *Pseudomonas* spp. (mean=560 ppm) followed by *E. coli* (mean=360ppm).

Plasmids that were extracted from 8 arsenic tolerant *E. coli* and one arsenic sensitive *E. coli* and run in agarose gel which showed 8 types of plasmid profiles of size ranging from 3.1 to 16.21 kb. The most frequent band was 14.17 kb (in four isolates) followed by 6.16 kb (two isolates) and 8.5 kb (two isolates). Apparently a size difference was observed in overall plasmid profiling. Large size bands were obtained from arsenic tolerant bacteria than arsenic sensitive bacteria (4.36Kb).

Key words: Arsenic, groundwater, arsenic tolerant bacteria, water quality, arsenicosis, plasmid profiles

TABLE OF CONTENTS

	Page No.
Title page	i
Recommendation	ii
Certificate of approval	iii
Board of examiners	iv
Acknowledgement	v
Abstract	vii
Table of contents	viii
List of abbreviations	xiii
List of tables	xv
List of figures	xvi
List of photographs	xvii
List of appendices	xviii
CHAPTER I: INTRODUCTION	1-4
CHAPTER II: OBJECTIVES OF THE STUDY	5
2.1 General objective	5
2.2 Specific objectives	5
CHAPATER III: LITERATURE REVIEW	6-30
3.1 Arsenic, its sources and occurrences in the environment	6
3.2 History of guideline development	8
3.3 Global scenario of arsenic poisoning of groundwater	8
3.4 National scenario of arsenic poisoning of groundwater	9
3.5 Interim Nepal guidelines and policies for arsenic in drinking water	11
3.6 VDCs vulnerability to arsenic	11

3.7 Physico-chemical parameters of water	11
3.7.1 pH	12
3.7.2 Temperature	12
3.7.3 Arsenic	12
3.8 Microbiological parameters of water	13
3.8.1 Coliform bacteria	13
3.8.2 Some intestinal pathogens found in contaminated drinking water	14
3.9 Health effects of arsenic	17
3.9.1 Route of entry	17
3.9.2 Acute toxicity of arsenic (III) and (V)	17
3.9.3 Chronic toxicity of arsenic (III) and (V)	18
3.9.3.1 Dermal	18
3.9.3.2 Vascular effects	18
3.9.3.3 Cancer	19
3.9.4 Characteristics of arsenicosis	19
3.9.5 Treatment	20
3.10 Mitigation of arsenic in drinking water	20
3.10.1 3-Kolshi filter	21
3.10.2 SONO filter	21
3.10.3 Arsenic bio-sand filter (Kanchan Arsenic Filter)	22
3.10.3.1 Arsenic removal unit	22
3.11 Testing and investigation	23
3.11.1 Hydride Generator Atomic Absorption Spectrophotometer	23
3.11.2 Blanket tubewell testing	23
3.12 Biological properties of arsenic	24
3.13 Microbial arsenic tolerance mechanisms	26
3.13.1 Chromosomal-determined resistance	27
3.13.2 Plasmid-mediated resistance	27
3.14 Minimum inhibitory concentration	28
3.15 Plasmid profiling	28

CHAPTER IV: MATERIALS AND METHODS	29-38
4.1 Materials	29
4.2 Methods	29
4.2.1 Study period	29
4.2.2 Study area	29
4.2.3 Study sites	29
4.2.4 Study population	30
4.2.5 Tools for data collection and analysis	30
4.2.6 Collection of samples for water analysis	30
4.2.6.1 Sample collection from tubewell	30
4.2.6.2 Sample collection from effluent water (filter)	31
4.2.7 Study of physico-chemical parameters of water samples	31
4.2.7.1 Temperature	31
4.2.7.2 pH	31
4.2.7.2 Measurement of arsenic concentration by kit method	32
4.2.7.3 Measurement of total arsenic concentration by AAS	32
4.2.8 Physical examination for skin manifestations of chronic arsenic ingestion	32
4.2.9 Microbial examination water sample	33
4.2.9.1 Total coliform count	33
4.2.9.1.1 Enumeration of coliforms	34
4.2.9.2 Enrichment	34
4.2.9.2.1 Detection of <i>Salmonella</i> and <i>Shigella</i> spp.	34
4.2.9.2.2 Detection of <i>Vibrio cholerae</i>	34
4.2.9.3 Isolation and identification of enteric bacteria	34
4.2.9.4 Enumeration of arsenic tolerant bacteria	35
4.2.9.5 Isolation and identification of arsenic tolerant bacteria	35
4.2.9.6 Minimum inhibitory concentration (MIC)	36
4.2.10 Extraction of plasmid DNA	36
4.2.11 Electrophoresis of plasmid DNA from arsenic tolerant <i>E. coli</i>	37

4.2.12 Determination of size of DNA by semi log plot	37
4.2.13 Decontamination of Ethidium Bromide	37
4.2.14 Data analysis	37
CHAPTER V: RESULTS	39-54
5.1 Physico-chemical parameters of water	39
5.1.1 Temperature	39
5.1.2 pH	39
5.1.3 Arsenic	40
5.1.3.1 Arsenic concentration in different VDCs/Municipality above and below NDWQS	40
5.1.3.2 Arsenic concentration before and after filtration	41
5.1.3.3 Relation of arsenic concentration with age of tubewell	42
5.1.3.4 Relation of arsenic concentration with depth of tubewell	44
5.1.4 Clinical manifestations of arsenicosis patient	46
5.1.4.1 Types of arsenicosis among people	46
5.1.4.2 Age and sex-wise distribution of arsenicosis cases	47
5.1.4.3 Sex and stage-wise distribution of arsenicosis cases	48
5.1.4.4 Distribution of arsenicosis vs. duration of tubewell use	48
5.2 Bacteriological analysis of water	49
5.2.1 Total coliform count	49
5.2.2 Isolation and identification of enteric bacteria by MF technique	50
5.2.3 Isolation and identification of arsenic tolerant bacteria by PCA	51
5.2.4 Colony count in PCA in different concentration of As (V)	51
5.2.5 Minimum inhibitory concentration of arsenic tolerant bacteria	52
5.3 Plasmid analysis	53
5.3.1 Profiles of plasmids from arsenic tolerant <i>E. coli</i>	53
CHAPTER VI: DISCUSSION AND CONCLUSION	55-63
6.1 Discussion	55

6.1.1 Physico-chemical parameters of water	55
6.1.2 Age of installation of tubewell	56
6.1.3 Depth of tubewell	57
6.1.4 Health affects	58
6.1.5 Microbial quality of water	59
6.1.6 Enteric bacteria isolated from water samples	60
6.1.7 Arsenic tolerant bacteria isolated from PCA	61
6.1.8 Colony count in PCA in different concentration of As (V)	61
6.1.9 Minimum inhibitory concentration of arsenic tolerant bacteria	62
6.1.10 Plasmid of arsenic tolerant and sensitive <i>E. coli</i>	62
6.2 Conclusion	62
CHAPTER VII: SUMMARY AND RECOMMENDATION	64-67
7.1 Summary	64
7.2 Recommendations	67
CHAPTER VIII: REFERENCES	68-76

LIST OF ABBREVIATIONS

AAS	:	Atomic Absorption Spectrophotometer
APHA	:	American Public Health Association
As (III)	:	Trivalent Arsenic
As (V)	:	Pentavalent Arsenic
As	:	Arsenic
ATP	:	Adenosine Tri-Phosphate
BFD	:	Black Food Disease
CBS	:	Central Bureau of Statistics
CFU	:	Colony Forming Unit
CIM	:	Composite Iron Matrix
DNA	:	Deoxyribonucleic Acid
DMAA	:	Dimethyl Arsinic Acid
DoHS	:	Department of Health Services
DOL	:	Department Of Irrigation
DWQIP	:	Drinking Water Quality Improvement Program
DWSS	:	Department of Water Supply and Sewage
ENPHO	:	Environment and Public Health Organization
EtBr	:	Ethidium Bromide
ETPAM	:	Environment Technologies Programs for Arsenic Mitigation
FFF	:	Filters for Families
FINNIDA	:	Finnish International Development Agency
GIS	:	Geographic Information System
H ₂ S	:	Hydrogen Sulphide
HGAAS	:	Hydride Generation Atomic Absorption Spectrophotometer
IARC	:	International Agency for Research on Cancer
IDWSS	:	International Decade of Water Supply and Sanitation
JRCS	:	Japanese Red Cross Society

KAF	:	Kanchan Arsenic Filter
Kbp	:	Kilo base pair
LD ₅₀	:	Lethal Dose-50
LF	:	Lactose Fermenting
mg/l	:	milligram per liter
µg/L	:	microgram per liter
MA	:	MacConkey Agar
MIC	:	Minimum Inhibitory Concentration
MIT	:	Massachusetts Institute of Technology
MMAA	:	Monomethyl Arsonic acid
MR	:	Methyl Red
NA	:	Nutrient Agar
NASC	:	Nepal Arsenic Steering Committee
NDWQS	:	National Drinking Water Quality Standard
NRCS	:	Nepal Red Cross Society
ppb	:	parts per billion
ppm	:	parts per million
RNA	:	Ribonucleic Acid
rpm	:	revolution per minute
RWSSSP	:	Rural Water Supply and Sanitation Support Program
SIM	:	Sulphide Indole Motility
SPSS	:	Statistical Package for Social Sciences
UNDP	:	United Nation Development Programme
UNICEF	:	United Nation Children's Fund
US-EPA	:	United State-Environment Protection Agency
VDC	:	Village Development Committee
VP	:	Voges Proskauer
WHO	:	World Health Organization

LIST OF TABLES

- Table 1:** Distribution of temperature of water samples
- Table 2:** Arsenic concentration in different VDCs/Municipality
- Table 3:** Arsenic concentration before filtration
- Table 4:** Arsenic concentration after filtration
- Table 5:** Arsenic concentration paired t-test analysis
- Table 6:** Classification of arsenic concentration by age of tubewell
- Table 7:** Classification of arsenic concentration by depth of tubewell
- Table 8:** Distribution of skin manifestation and gender
- Table 9:** Distribution of arsenicosis by types and sex
- Table 10:** Age and sex-wise distribution of arsenicosis cases among people
- Table 11:** Sex and stage-wise distribution of arsenicosis cases among people
- Table 12:** Distribution of arsenicosis cases versus duration of tubewell use
- Table 13:** Source wise distribution of enteric bacterial isolates of water samples
- Table 14:** Types of bacteria isolated from arsenic contaminated groundwater
- Table 15:** Colony count in Plate Count Agar in different concentrations of arsenic (V)
- Table 16:** Minimum inhibitory concentration of arsenic tolerant bacteria to arsenic species (As III and As V)
- Table 17:** Plasmid profiles of selected arsenic tolerant *E. coli*

LIST OF FIGURES

- Figure 1:** Kanchan™ arsenic filter cross section
- Figure 2:** Illustration of arsenic removal mechanism
- Figure 3:** The environmental and global cycle of arsenic and its compounds
- Figure 4:** Comparison of pH measurements of water samples with standards
- Figure 5:** Scattered diagram of age of installation of tubewells and arsenic concentration
- Figure 6:** Scattered diagram of depth of tubewells and arsenic concentration (influent water)
- Figure 7:** Analysis of water samples by total coliform count

LIST OF PHOTOGRAPHS

- Photograph 1: Arsenic tolerant *Bacillus subtilis* isolated on nutrient agar (NA) incorporated with arsenic concentration
- Photograph 2: Enumeration of arsenic tolerant bacteria in different concentrations of arsenic (V) by Pour plate method
- Photograph 3: Biochemical tests of *E. coli*
- Photograph 4: Colonies of total coliforms on M-endo agar
- Photograph 5: Plasmid profiles of arsenic tolerant and sensitive *E. coli*
- Photograph 6: Spotted melanosis on the back side of chest of an arsenicosis patient
- Photograph 7: Diffused keratosis on both soles of an arsenicosis patient
- Photograph 8: Keratosis on both palms of an arsenicosis patient
- Photograph 9: Arsenic concentration measurement by using Wagh tech kit
- Photograph 10: Water sample collection from shallow tubewell

LIST OF APPENDICES

Appendix-I	Questionnaire
Appendix-II	Maps of study area
Appendix-III	List of the equipment and materials used during the study
Appendix-IV	I. Composition and preparation of different culture media II. Composition and preparation of different biochemical media
Appendix-V	Composition and preparation of different staining and test reagents
Appendix-VI	Staining methods; methodology of biochemical tests used for identification of bacteria
Appendix-VII	Biochemical characteristics of different bacteria
Appendix-VIII	Minimum inhibitory concentration of different arsenic concentration
Appendix-IX	Protocols
Appendix-X	Number of KAF distributed in Nawalparasi District
Appendix-XI	Standard curve of known DNA bands Vs distance
Appendix-XII	Arsenic test data from different VDCs/Municipality of Nawalparasi District
Appendix-XIII	Method of application as described in Wagtech international arsenic test kit leaflet